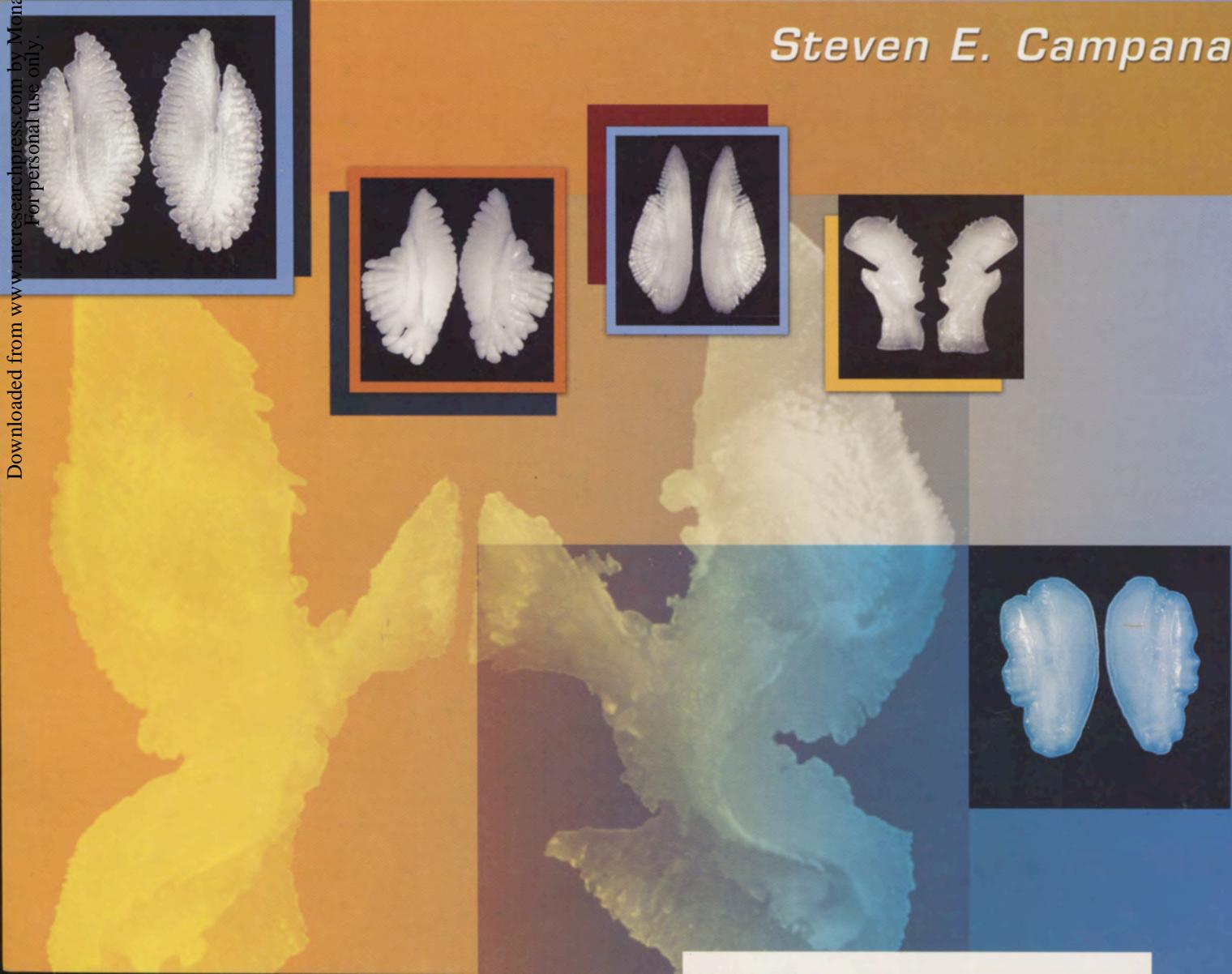


Photographic Atlas of **Fish Otoliths** of the Northwest Atlantic Ocean

Canadian Special Publication of Fisheries and Aquatic Sciences 133

Steven E. Campana



Photographic Atlas of Fish Otoliths of the Northwest Atlantic Ocean

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Steven E. Campana

Marine Fish Division
Bedford Institute of Oceanography
P.O. Box 1006, Dartmouth
Nova Scotia B2Y 4A2, Canada

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Contents

Abstract/Résumé	vii
Acknowledgements.....	viii
Introduction	1
Otolith location and function	2
Otolith composition	3
Otolith morphology	3
Biological factors affecting otolith morphology	6
Effects of preservation on otolith morphology	6
Methods	7
Using the atlas	8
References	10
Photographic plates	13
Alphabetical species list	279

Abstract/Résumé

The shape of fish otoliths is highly species specific. Since otoliths resist degradation better than most other tissues, the shape and size of preserved or undigested otoliths recovered from fossilized sediments, native middens, and the stomachs and droppings of fish predators can be used to reconstruct the species composition of the diet or fish assemblage. This photographic atlas presents light and (or) scanning electron micrographs of 580 pairs of sagittal otoliths representing 288 species, 97 families, and 27 orders of fish from the northwest Atlantic. For most species, multiple individuals across a range of sizes are presented in order to highlight changes in otolith shape with increased size. For 72 of the families, photographs of the lapillar and asteriscal otoliths are also presented.

Chez les poissons, la forme des otolithes est très particulière à chaque espèce. Puisque les otolithes résistent mieux à la dégradation que la plupart des autres tissus, la forme et la taille des otolithes, préservés ou non digérés, prélevés des sédiments fossilisés, des dépotoirs autochtones et des estomacs ou des excréments de prédateurs peuvent être utilisés pour reconstituer la composition taxinomique du régime alimentaire d'un poisson ou de l'habitat dans lequel il vit. Cet atlas photographique propose des images, obtenues à partir de microscopie optique ou de microscopie électronique à balayage, de 580 paires d'otolithes sagittaux, représentant 288 espèces, 97 familles et 27 ordres de poissons de l'Atlantique Nord-Ouest. Pour la plupart des espèces, plusieurs spécimens d'une gamme de tailles variées sont présentés pour mettre en évidence les changements dans la forme des otolithes en fonction de la taille du poisson. Des photographies des asteriscii et des lapilli sont également offertes pour 72 familles.

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Introduction

Otoliths (“earstones”) are small, white structures found in the head of all fishes other than sharks, rays, and lampreys. Although they are located within the skull adjacent to the brain, they are not attached to the skull, but are retained within the transparent tubular canals of the inner ear. Otoliths provide a sense of balance to fish in much the same way that the inner ear provides balance in humans. Fish otoliths also aid in hearing (Popper and Lu 2000).

To the fisheries biologist, the otolith is one of the most important tools for understanding the life of fish and fish populations. Growth rings (annuli) not unlike those of a tree record the age and growth of a fish from the date of hatch to the time of death (Casselman 1987). Daily growth increments formed in the first year of life record daily age and growth patterns in surprising, albeit microscopic, detail (Campana and Neilson 1985). In addition, chemical and elemental assays allow the reconstruction of everything from the year of hatch, to migration pathways, to population identity, to the temperature of the water (Campana 1999). Indeed, virtually the entire lifetime of the fish is recorded in the otolith. For that reason, otoliths are used and studied in almost every fisheries laboratory in the world, and form the basis for most age-structured analyses of fish populations around the world (Summerfelt and Hall 1987; Secor et al. 1995; Fossum et al. 2000). Recent estimates indicate that more than 800 000 otoliths were aged worldwide in 1999, with many more analyzed for shape, chemical composition, and other applications (Campana and Thorrold 2001).

Otoliths have a distinctive shape which is highly species specific, but varies widely among species (Maisey 1987). Thus fish, seal, and seabird biologists, as well as taxonomists and archaeologists, often rely on the shape and size of preserved or undigested otoliths to reconstruct the species and size composition of the diet of fish predators (Murie and Lavigne 1985; Jobling and Breiby 1986; Barrett et al. 1990). Preserved otoliths may also serve to identify fossil fish assemblages for phylogenetic or climatological studies (Nolf 1985;

Andrus et al. 2002). Identification is aided by the fact that otoliths resist degradation better than most other tissues (Cottrell et al. 1996), and are often the only identifiable animal remains recovered from stomachs and droppings, as well as from Indian middens.

Reference collections of otoliths now exist for several locations around the world, although none claim to be comprehensive. The best published descriptions are those of Smale et al. (1995) for South African fishes, Harkonen (1986) for northeast Atlantic fishes, Morrow (1976) for the Bering Sea, and Nolf (1985) for fossil fishes. Regional collections also exist for Argentina (Volpedo and Echeverria 2000), Antarctica (Hecht 1987; Williams and McElroy 1990), and the northeastern Pacific (Harvey et al. 2000). There are no published reference collections for the northwest Atlantic, nor do any of the published works contain photographs of the non-sagittal otoliths (the lapilli and asterisci).

The intention of this book is to provide a photographic reference book for those using or studying the otoliths of fishes of the northwest Atlantic. This work will be of particular interest to those who reconstruct the diet of fish, seals, seabirds, and other fish predators, as well as archaeologists, paleontologists, and taxonomists. Biologists preparing to age previously unstudied fish species will also find the book of value.

This otolith atlas presents light and (or) scanning electron micrographs (SEM) of 580 pairs of sagittal otoliths representing 288 species, 97 families, and 28 orders of fish. Although the species coverage in this book is extensive, it does not include all of the 538 species which have been documented in the northwest Atlantic (although many of these species are considered rare or transient). For most species, multiple individuals across a range of sizes are presented in order to highlight changes in otolith shape with increased size. However, the presentation is limited to post-juvenile stages on account of the relative absence of distinguishing features among larval fish otoliths. For 72 of the families, photographs of the lapillar and asteriscal otoliths are also presented.

Otolith location and function

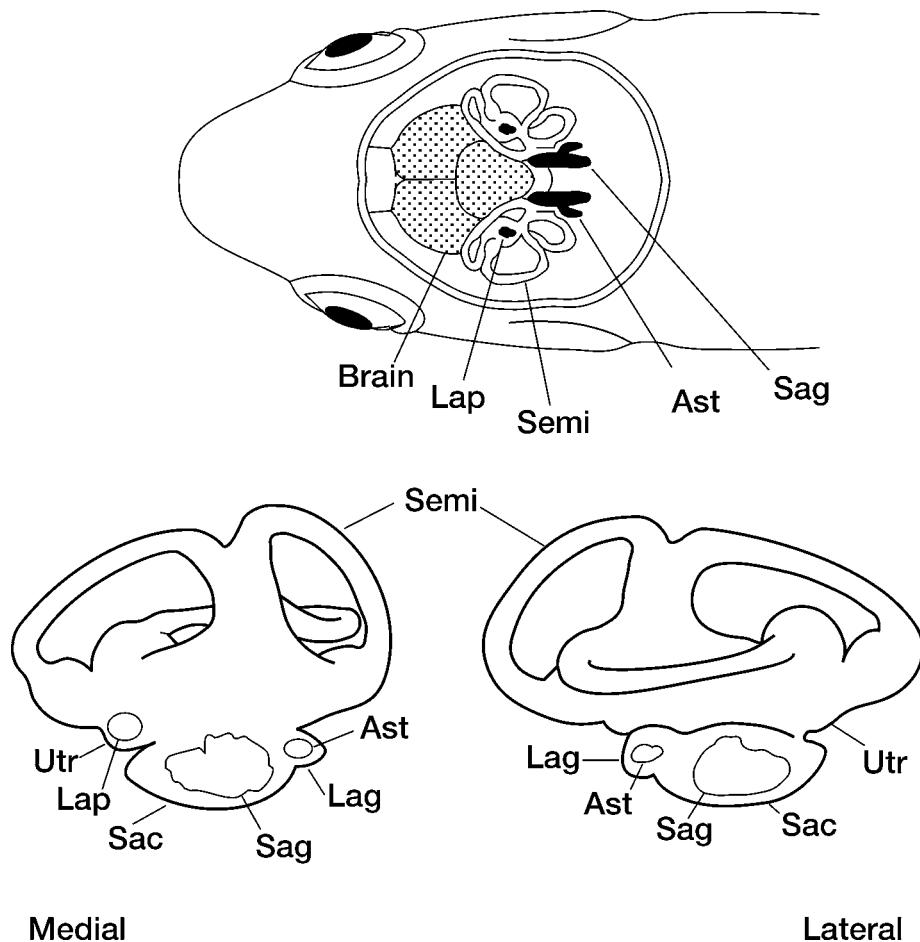
The inner ear is the primary vestibular organ in fishes and other vertebrates, responsible for balance and orientation in three dimensions (Popper and Lu 2000). The inner ear also aids in sound detection in most fishes. The inner ear is located adjacent to the brain, within and occasionally invaginated in the neurocranium (Fig. 1). Composed of a series of interconnecting semi-circular canals, the fluid-filled inner ear looks delicate and translucent but is surprisingly tough. Inner ear architecture varies somewhat among species, but a common feature is the presence of three pairs of chambers, each of which contains an otolith (Fig. 1). Since otoliths are the only solid bodies within the endolymph fluid, changes in orientation and acceleration are detected by slight shifts in the location of the otoliths relative to the surrounding chamber. A sensory epithelium in the form of a macula lies on one wall of the chamber, coupled to the otolith via an otolithic membrane (Popper and Platt 1993). This sensory epithelium is thought to be responsible for the detection of both sound and changes in posture. Thus all otoliths appear to share a vestibular and sound detection function, although the balance between the

two functions may vary with the otolith. In general, the lapilli appear to be more associated with posture and the sagittae with sound detection.

The three pairs of otoliths tend to have a size and shape representative of the chamber within which they are held (Fig. 1). Each sacculus contains a sagitta (plural: sagittae), which is often the largest otolith in all but the ostariophysian fishes. The sacculus and sagitta on a given side are usually ventral to the posterior part of the brain, lateral to but close to the midline of the brain. Slightly posterior to each sacculus is a lagena, containing an asteriscus (plural: asteriscii). The asteriscus is so close to the sagitta in many fishes that they are often removed together when the sacculus is pulled out. Considerably more anterior and dorsal on each side is the utricle containing the lapillus (plural: lapilli). The lapillus is often the smallest of the otoliths.

In ostariophysian (otophysan) fishes, a chain of Weberian ossicles connecting the swimbladder to the inner ear enhances sound detection. In these fishes, the asteriscii are usually larger than the sagittae.

Fig. 1. Schematic of the location of the inner ear and three pairs of otoliths in the skull of a generalized teleost. *Top*, Dorsal view of the inner ear and otoliths in relation to the brain in a cutaway of a fish skull (modified from Secor et al. 1992). *Bottom*, Position of otoliths and otolith chambers in the inner ear of the teleost *Trichogaster* (modified from Popper and Hoxter 1981). Ast, asteriscus; Lag, lagena; Lap, lapillus; Sac, sacculus; Sag, sagitta; Semi, semi-circular canal of labyrinth; Utr, utricle.



Otolith composition

Otoliths are very pure compared to most biological and mineralogical structures, with the composition being dominated by calcium carbonate in an organic matrix. Most otoliths contain more than 95% by weight of calcium carbonate, with 3–5% in the form of an organic matrix, and less than 1% as non-organic trace impurities. The trace element and stable isotope composition of the otolith has been given extensive study, owing to numerous applications in reconstructing the environmental history, migration, and population identity of fishes (Campana 1999).

Calcium carbonate can crystallize as any one of three crystal polymorphs: calcite, aragonite, or vaterite. However, the vast majority of sagittal and lapillar otoliths are composed of aragonite, which has a milky white appearance (Carlström 1963; Oliveira et al. 1996; Campana 1999). This is unlike the otoconia of mammals, which are composed of calcite.

Curiously, different polymorphs of calcium carbonate appear to be linked to the different otolith organs. While aragonite is the norm for sagittae and lapilli, most asteriscii are made of vaterite, thus accounting for their glassy appearance (Oliveira et al. 1996). Vaterite is also the principal polymorph in many aberrant, or “crystalline”, otoliths (Mugiya 1972). Calcitic regions in otoliths are much rarer.

The implications of an otolith composition dominated by calcium carbonate lie most clearly with otolith preservation and stability. Both calcium carbonate and otoliths are stable for many years when stored dry. However, calcium carbonate is acid soluble, so preservation in even weakly acidic solutions will result in dissolution of the otolith.

Otolith morphology

The three pairs of otoliths differ markedly in shape and appearance. In most adult fishes, the sagittae are the largest pair and the lapilli the smallest (Fig. 2). In contrast, asteriscii are larger than sagittae in ostariophysian fishes (a group which includes the minnows and catfish). Sagittal shape differs substantially among species, while lapillar shape is more uniform. The shape of the asteriscii shows intermediate inter-specific variability. Within an otolith pair, the left and right otoliths are very similar, but not identical, mirror images of each other. Interestingly, the left and right asteriscii can differ considerably more in shape than the other otolith pairs (Campana and Casselman 1993).

The orientation and major landmarks of a typical sagitta are shown in the labelled scanning electron microscope (SEM) photo of Fig. 3. The rostrum, antirostrum, and postrostrum are consistent features of all sagittae, although their size and extension varies substantially among species. The sulcus, which represents the point of attachment of the sensory macula, is also a consistent feature of sagittae. Although the fine details of sulcus morphology are not documented here, those details are defined elsewhere and can prove helpful in some species identifications (Nolf 1985; Smale et al. 1995). An intricate morphology is also evident in the SEM images of asteriscii, but less so in the case of the lapilli (Fig. 4). It is possible that inter-specific differences in the shape of asteriscii and lapilli could be used to complement the differences observed in the sagittae. However, in light of the relatively small size of the lapilli and sagittae, SEM would probably be required by the end user to observe all but the most gross of morphological differences.

In any given species, otolith size and shape often changes substantially with fish growth (Fig. 5). In virtually all young fish larvae, otoliths tend to be relatively featureless: spherical or smoothly oblate in most species, and discoid in species (such as salmonids) which hatch at a larger size. In most species, the sagittae and lapilli are present at hatch, while the asteriscii first appear at an age of 2–3 weeks. At this early stage of fish development, relative otolith sizes can be inverted, with lapilli being larger than sagittae (Campana 1989). Otoliths first acquire the main features of their mature shape in the juvenile stage. As the size-specific photos of this atlas demonstrate, otolith shape can remain diagnostic but still change in later life as the fish (and otolith) grows. As a result, otolith size must be taken into consideration, as well as shape, when identifying a species from an otolith. In particular, the otolith shape of very large fish can differ substantially from those of average-sized adults.

Because of their function in maintaining the balance of the fish, otoliths tend to grow as the fish grows. Therefore, there is almost always a strong relationship between otolith size and fish size (Hunt 1992). Given a measurement of otolith size (whether in terms of length or weight), it is possible to estimate the length of the fish from which the otolith was obtained (Fig. 6). These estimates provide useful approximations of fish length, but cannot be interpreted too strictly, since the fish-otolith regression often differs among populations or groups of fish with different growth rates (Campana 1990). It is also important to note that the relationship between fish and otolith length is not necessarily linear, and that the relationship for larvae is often very different from that for adults.

Fig. 2. Light micrograph of the three pairs of otoliths from a 23-cm adult white perch (*Morone americana*). The left-hand otolith of each pair is shown on the left side.

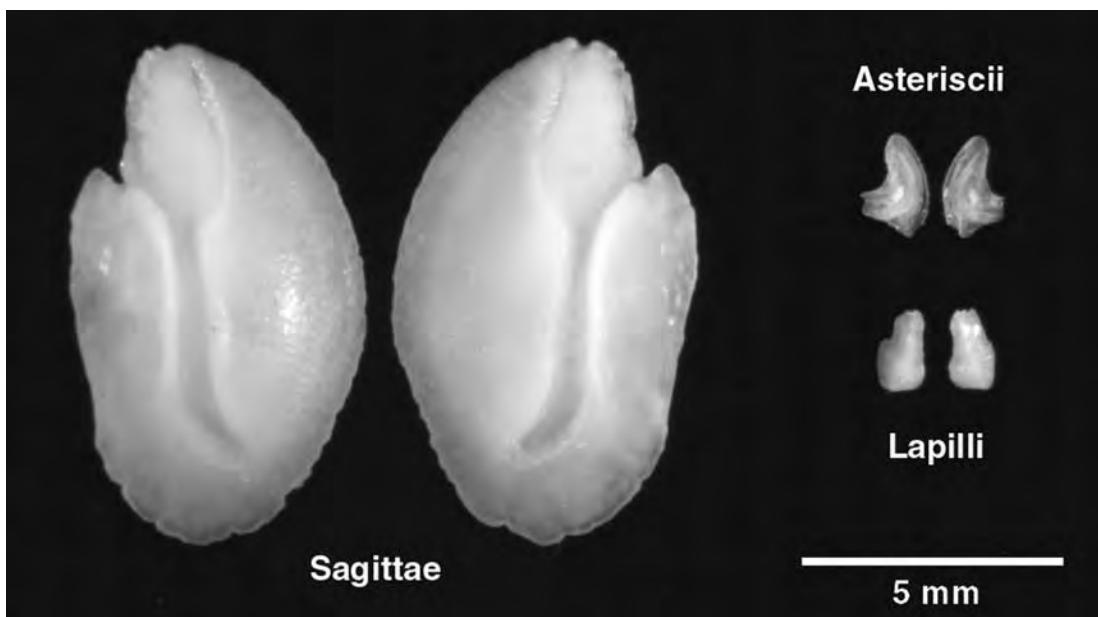


Fig. 3. Morphology of a haddock (*Melanogrammus aeglefinus*) sagitta evident in proximal (top sagitta) and distal (bottom sagitta) views with SEM.

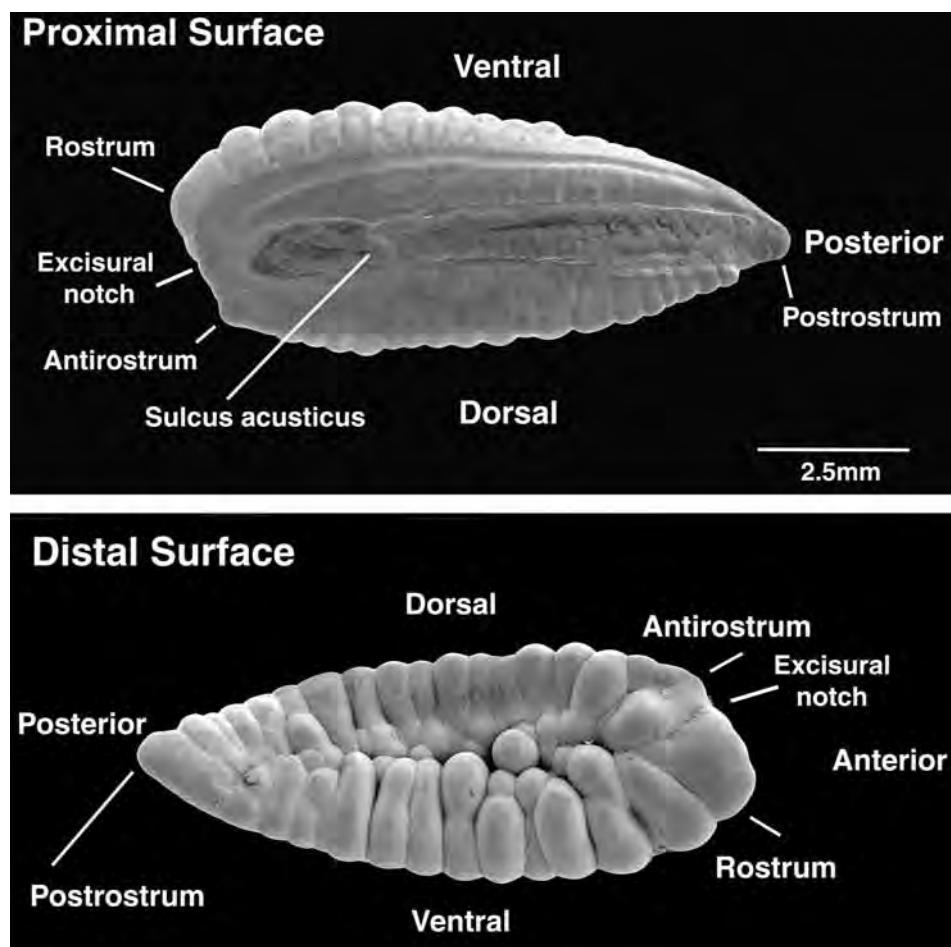


Fig. 4. Morphology of a cod (*Gadus morhua*) asteriscus (left panel) and lapillus (right panel) evident in top and bottom views with SEM.

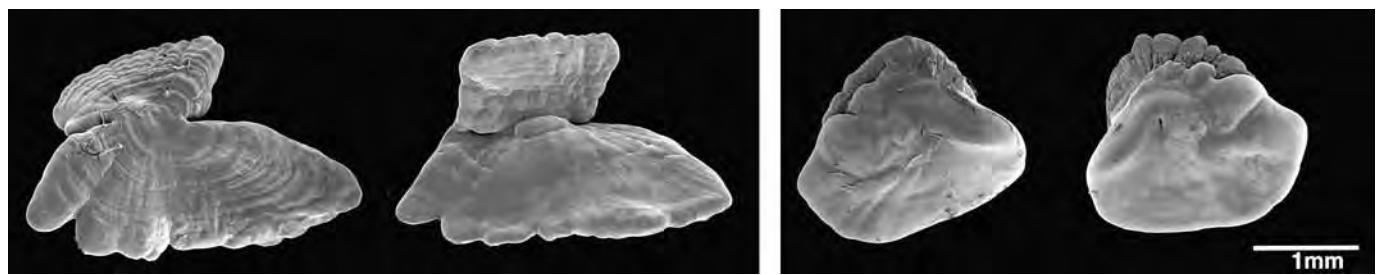


Fig. 5. Ontogenetic sequence showing the change in size and shape of a haddock sagitta from the larval stage (37 days old) to that of a 6-year-old adult. Note that the individual images have been re-scaled to allow presentation in a single figure; use scale bars for correct size.

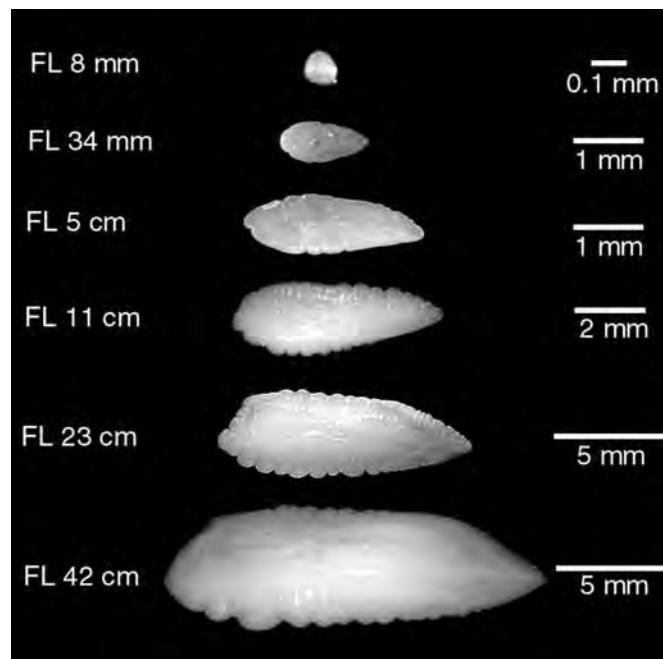
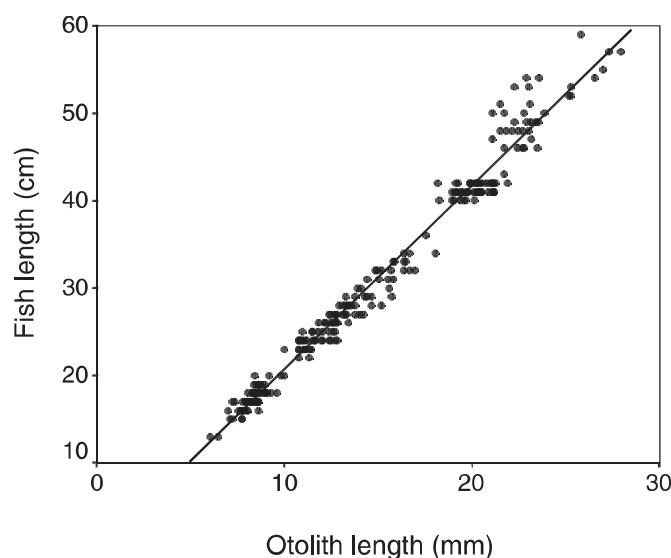


Fig. 6. Relationship between fish length and sagittal length in silver hake (*Merluccius bilinearis*; $n = 235$; $r^2 = 0.97$), which could be used to predict fish length based on the size of the recovered otolith. A linear relationship between fish and otolith size is characteristic of the juvenile and adult stages of most fish species, but the same relationship seldom applies to the larval stage.



Biological factors affecting otolith morphology

There are a broad range of biological factors which influence or moderate otolith shape. These factors can operate at a range of scales, from that of general phylogeny to the individual level. Few of these factors are well understood, but those that are known or suspected are mentioned here.

There are no broad phylogenetic principles which are known to guide otolith shape (Maisey 1987). Although family- or genus-level otolith characteristics are often present, it is often impossible to predict otolith shape for any given species. There may be functional relationships however. Based on numerous observations, I have noted that fast-swimming fishes capable of rapid acceleration and turning tend to have smaller otoliths than their slower swimming counterparts. The tunas and swordfish are good examples of this phenomenon, whereby the sagittal otolith of a 400-kg bluefin tuna (*Thunnus thynnus*) is smaller than that of a 1-kg cod. Species capable of good sound production (and presumably good sound detection) can also be expected to have large saggital otoliths. Members of the Sciaenidae (grunts and drums) are characteristic of this group; species such as the black drum (*Pogonias cromis*) have sagittae which are among the largest observed. In contrast, the families within the group Ostariophysii (such as cyprinids and catfishes), which possess a chain of Weberian

ossicles to enhance sound detection, have somewhat smaller sagittae and larger asteriscii than normal.

Within a species, otolith shape can vary with the sex, population, and growth rate, as well as the stage of ontogeny described in the previous section. The magnitude of shape differences due to ontogeny and fish size is considerably larger than that due to sex, population, and growth rate, since the effect of the latter factors may be detectable only through statistical analysis (Campana and Casselman 1993; Cardinale et al. 2003).

In general, otoliths within an otolith pair are very similar but non-identical mirror images of each other. However, left versus right asymmetry is common in flatfish, a fish in which the eyes migrate to the same side of the head at around the time of metamorphosis to the settled juvenile. The presence or degree of asymmetry seems to vary among individuals, and is most evident in large individuals. In general, however, the sagitta found on the upper side of the fish (the right otolith in right-eyed flatfish) is irregularly shaped or occasionally shorter and thicker than the sagitta which faces down in the adult fish. The functional significance of this otolith asymmetry is unknown, but is presumably related to a reduced or altered function in one of the two otoliths.

Effects of preservation on otolith morphology

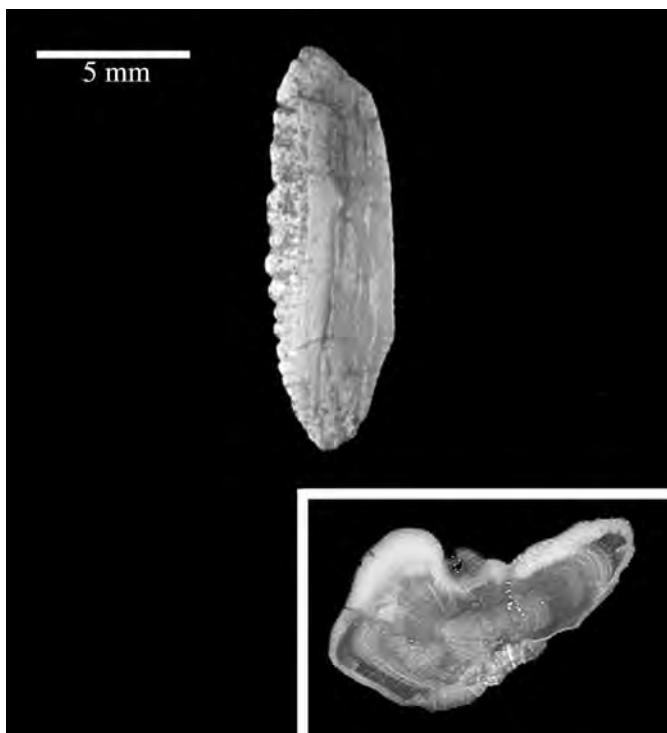
The shape and size of otoliths recovered from the stomach or feces of fish predators has long been used to reconstruct the species and size composition of the predator's diet (Murie and Lavigne 1985; Jobling and Breby 1986; Barrett et al. 1990; Pierce et al. 1991; Bowen et al. 1993; Dolloff 1993; Burns et al. 1998). In many cases, there are few alternatives, since otoliths are often the only animal remains that are recovered, let alone identified to species. Nevertheless, there are limitations to this application. Several studies have fed fish of known species and size composition to seals, and then recovered the ingested otoliths from the stomachs or feces (Dellinger and Trillmich 1988; Cottrell et al. 1996; Tollit et al. 1997; Bowen 2000). In all cases, sources of bias have been noted, associated primarily with the relatively rapid dissolution of small and (or) fragile otoliths in the acidic stomach environment. As a result, it seems likely that any dietary reconstruction could underestimate the contribution from fish species with small otoliths, or from smaller individuals. Even where complete otolith dissolution does not occur, partial dissolution can leave an otolith unrecognizable to species, or perhaps smaller than its original size. Examples of partial dissolution of otoliths recovered from seal droppings are shown in Fig. 7.

Preserved otoliths may also serve to identify fossil fish assemblages (Elder et al. 1996), date sedimentary strata (Gaebers 1984), reconstruct historical populations (Hales and Reitz 1992), prepare phylogenies (Nolf 1995), reconstruct ancient climates (Ivany et al. 2000; Andrus et al. 2002), and provide indicators of seasonal occupation for ancient peoples (Van Neer et al. 1993). Such otoliths may be recovered from aquatic sediments, fossil grounds, or archaeological middens, where they have been exposed to possibly acidic conditions or chemical leaching. Such conditions have the potential to bias reconstructions of past assemblages, if otoliths have been dissolved, or to alter climatic reconstructions if chemical leaching has occurred. However, other indicators can often be used to determine if otolith alteration has occurred. In the case of archaeological applications, bias is not usually a problem, since the growth increments used to determine seasonality are either present or absent. For reasons not fully understood, otoliths in some fossil middens may sustain little damage after thousands of years of preservation, while others are rendered illegible after only a few hundred years (Fig. 8).

Fig. 7. Morphology of silver hake (*Merluccius bilinearis*) sagittae recovered from seal droppings, reflecting increasing effects of digestion from left to right. The image on the far left is that of an undigested silver hake sagitta.



Fig. 8. Photo of an Atlantic tomcod (*Microgadus tomcod*) otolith recovered from a midden after about 600 years of burial. The otolith surface retains most of its detail, and thus can be used for species identification. However, a transverse section (inset) indicates that diagenesis has occurred, obscuring much of the age and seasonality information.



Methods

All identifications of common species were confirmed with established keys for the region (Scott and Scott 1988; Bigelow and Schroeder 1953). For unusual species, or for any fish where there was the slightest uncertainty concerning species identity, identification was confirmed by fish taxonomists. Uncertainty concerning the validity of existing classifications for the liparids, including the probable presence of undescribed species in the northwest Atlantic, suggest that caution should be used in the interpretation of this family (Lou Van Guelpen, Huntsman Marine Science Centre, St. Andrews, NB, personal communication). The taxonomic status of *Ammodytes* spp. is also unknown. All family associations and phylogenies were based on Nelson (1994).

Source fish used to provide otoliths were either frozen or preserved in 95% ethanol, so as to insure complete preservation of the otoliths. In the few instances where fish were fixed in formalin for a few days prior to otolith removal, the formalin was first made basic to a pH of 8–9 through addition of sodium carbonate. All three pairs of otoliths were removed wherever possible; removal of the inner ear canals made it easier to identify left and right pairs. Adhering tissue was removed immediately after removal. Otoliths were then stored dry until the time of examination.

The default method of photography for all otolith pairs was with reflected light under a dissecting microscope at a magnification of 3–40 times. Previous otolith atlases have used either SEM photography or line drawings, both of which provide excellent representations of otolith shape, but neither of which make the otolith appear as it would to the atlas user. Since most users rely on reflected light microscopy for otolith examination, light microscopic images can be particularly helpful in an atlas. Oblique lighting with a dual fibre optic light source helped provide visual contrast to the images. Sagittal otolith pairs were first photographed on the medial side, then turned over and photographed on the distal side. Lapillar and asteriscal pairs were photographed from a single side only. Images were captured with a digital video camera at a resolution of 1280×1024 . All images were automatically digitally enhanced to improve contrast or sharpen edges, but not at the expense of making the images appear artificial. Optimas software was used for image capture and automated enhancement, while Photoshop was used for any subsequent enhancements.

SEM images were prepared for sagittae of most species in order to capture morphological detail in and around the sulcus. Sagittae were prepared for SEM by coating with carbon in a vacuum evaporator. Photographs were taken at a magnification of 18–35 times.

Using the atlas

Fish systematics is an evolving science; hence species affiliations within a family are not necessarily stable. For this reason, this atlas is arranged alphabetically by order, rather than phylogenetically. Within each order, families are presented alphabetically, as are species within families. A key to identify unknown otoliths to the order or family level was attempted but was discontinued, owing to the difficulty of providing diagnostic keys which take into account the changing shape of many otoliths with increasing size. However, outlines of “representative” sagittae for each order are provided in Fig. 9. It is important to note that there can be wide variations in otolith shape within orders and families, and that “representative” outlines are not necessarily representative. This is particularly true for the Perciformes, which comprises dozens of families.

For most species, representative otolith images occupy the entire page. Where available, an SEM image of the sagitta heads the page in order to highlight details of the sulcus. For at least one species of each family, an accompanying pair of light microscopic images to the right of the SEM image shows the lapilli and asteriscii (note, however, that left- and right-side identifications of these two otolith pairs are not necessarily correct). Below these images are a series of light microscopic images showing multiple pairs of sagittae from fish of different sizes, arranged from smallest to largest. Image scales often vary across the images, so the accompanying scale bar should be used to estimate otolith size. Each sagittal pair is shown both medial side up (sulcus side) and distal side up, with the left otoliths arranged on the left side of the image panel wherever possible. In most cases, the rostrum and antirostrum are oriented up. However, it was not always possible to identify the rostrum in some sagittae; hence caution is required in interpreting the otolith orientations too strictly.

In most cases, external profile will be sufficient to identify an otolith to species. The key features include the relative size of the rostrum, antirostrum, postrostrum, and excisural notch. The length of the sulcus can also be diagnostic. Where mor-

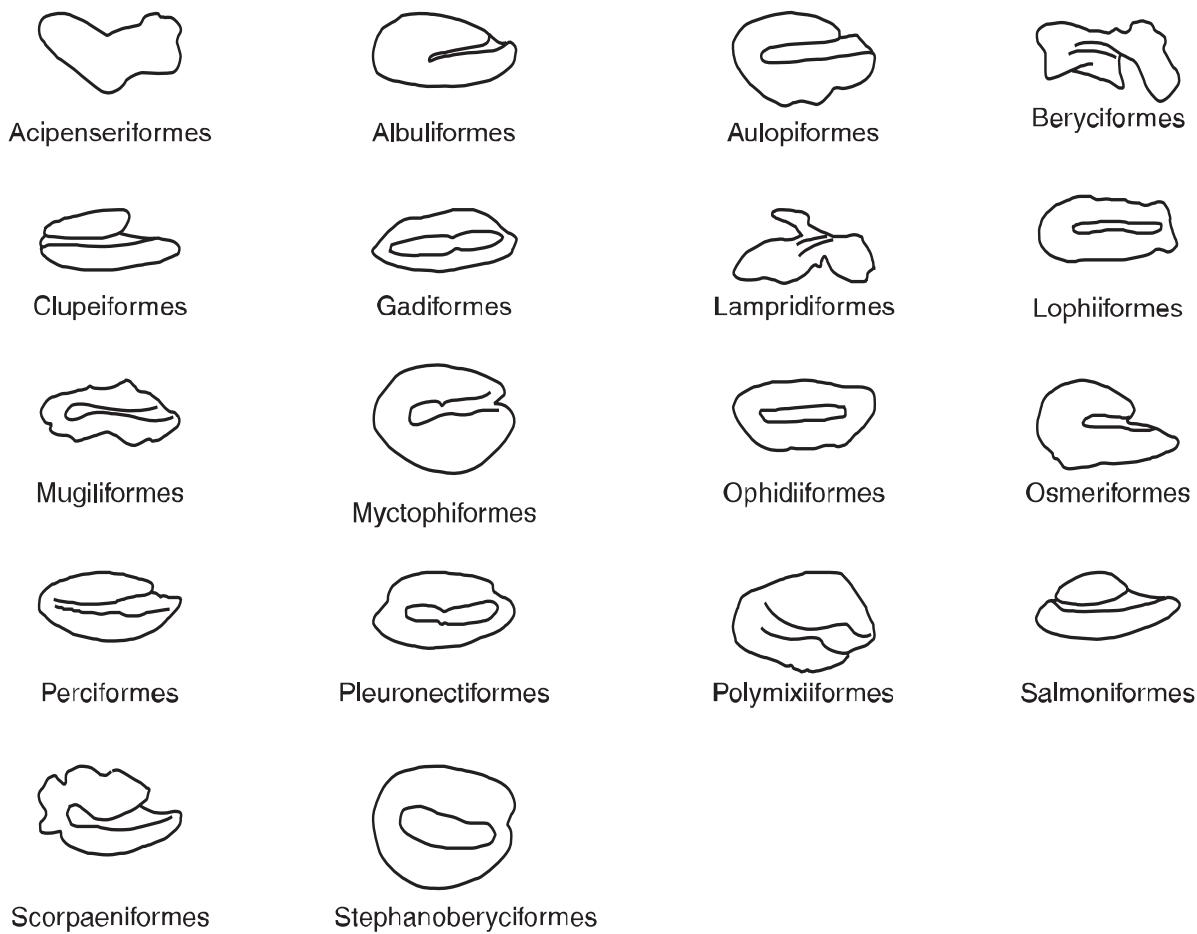
phological details of the sulcus are required, it can be difficult to view these details with a light microscope. However, use of oblique lighting is often helpful in providing visual contrast. Further contrast can be provided by sprinkling powdered graphite over the medial surface of the sagitta, and then lightly tapping the otolith on its side to remove any excess. Alternatively, a graphite pencil can rubbed over the sulcus region. If this is not sufficient, SEM may be required.

Clearly, intact and well-preserved otoliths will be easier to identify than will those which have been degraded or eroded. Although freshly removed otoliths provide excellent samples, otoliths which have been stored dry after removal remain in excellent condition almost indefinitely. Fish which have remained frozen after capture also provide well-preserved otoliths, as will those which have been preserved in 95% ethanol. Note, however, that ethanol becomes increasingly acidic as concentration drops, and that otoliths will dissolve in concentrations below about 70–80%. In addition, ethanol concentration declines soon after the fish carcass is added, owing to dilution from the water in the fish tissues. Complete ethanol replacement after 12–24 hours helps keep concentrations high. Formalin is not a recommended preservative for fish otoliths, since even buffered formalin is slightly acidic and will dissolve otoliths. However, short-term storage in formalin is possible if the formalin is first made basic with sodium carbonate to a pH of at least 8.

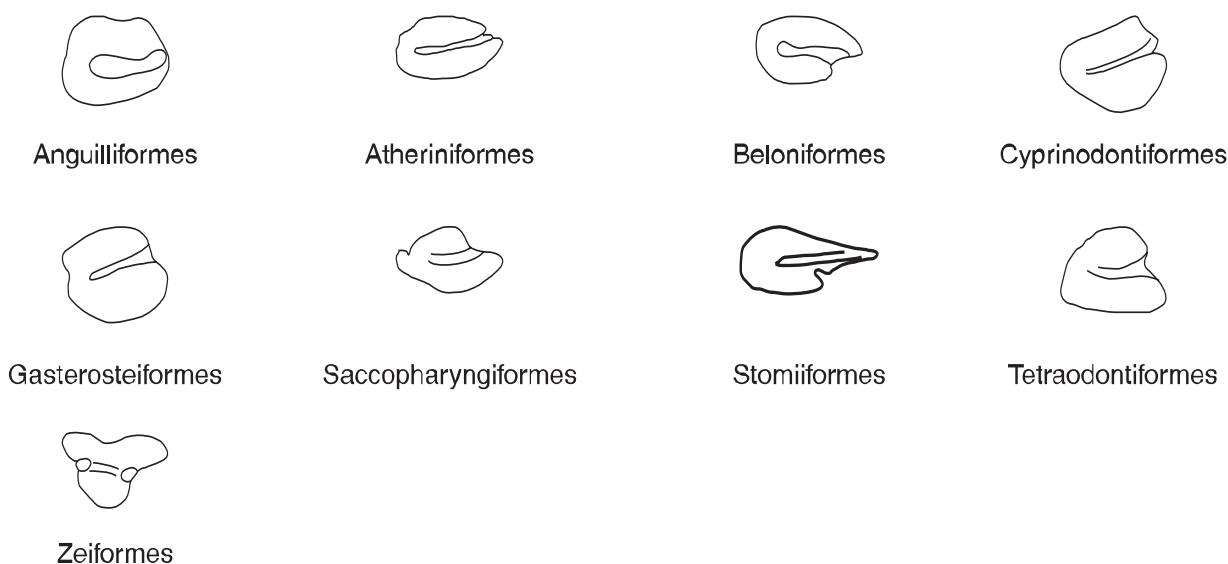
It is often possible to identify the characteristics of otolith dissolution or degradation. Rounded edges, particularly at the tips of the rostrum and postrostrum, often occur during digestion in an acidic stomach, and can signify an overall loss of material and size. Discolouration (usually brown or black) is a sign of degradation seen in both formalin-preserved material and in otoliths from archaeological sites. A chalky white appearance is a sign of exposure to mildly acidic conditions. None of these conditions should be confused with the irregular glassy appearance of “crystalline” otoliths, which are uncommon but natural occurrences in most species of fishes.

Fig. 9. Schematic outlines of adult sagittae representing each of the 27 orders of fish found in this atlas. The two sections of the figure separate orders in which the sagittae span a broad range of sizes (*top*) from those where the sagittae are typically ≤ 3 mm in diameter (*bottom*). Note that “representative” outlines are difficult to provide for orders such as the Perciformes, which contains many families.

Adult sagittae, all sizes



Adult sagittae, ≤ 3 mm



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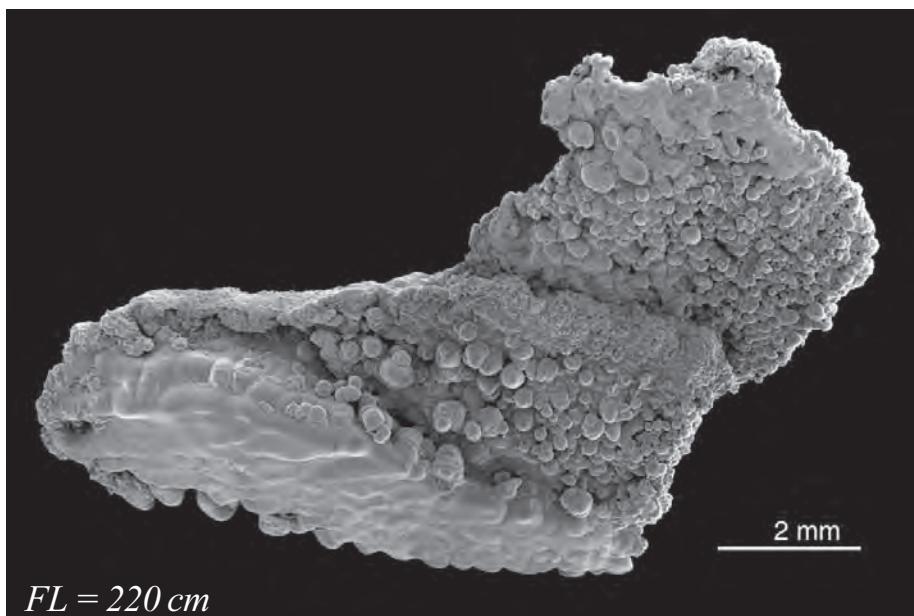
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Photographic plates

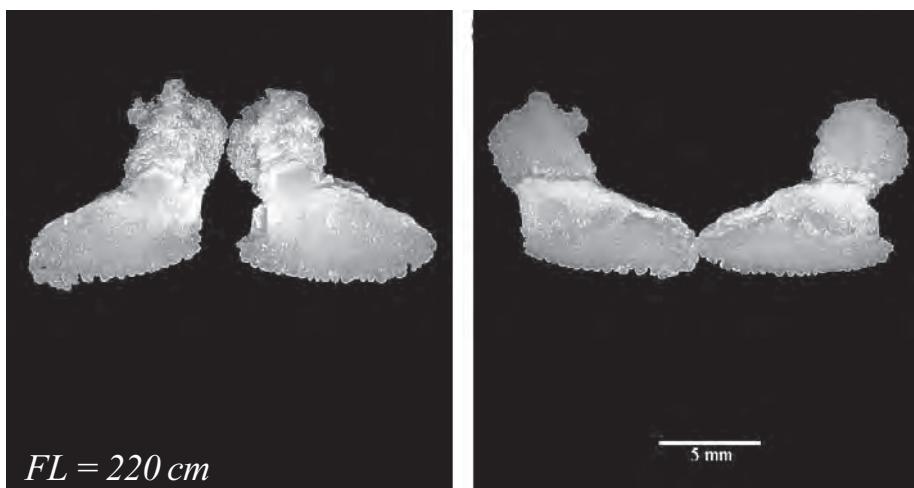
Acipenseriformes

15

Acipenseridae: *Acipenser oxyrinchus*



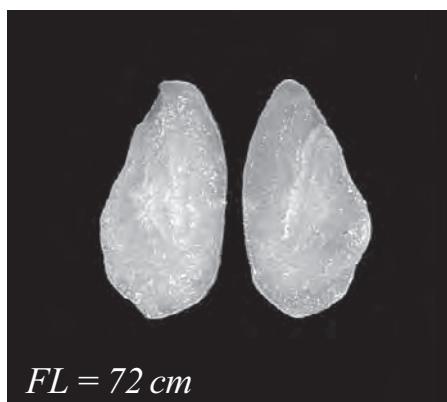
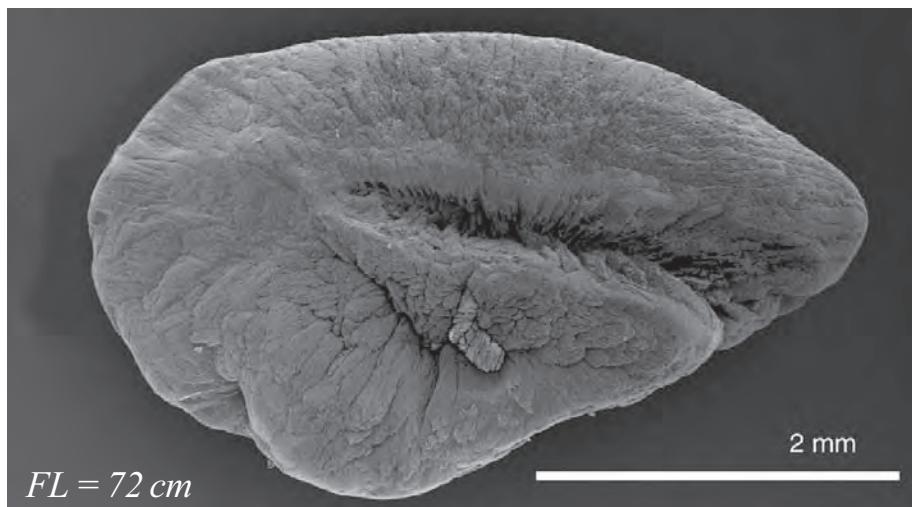
FL = 220 cm



FL = 220 cm

Albuliformes

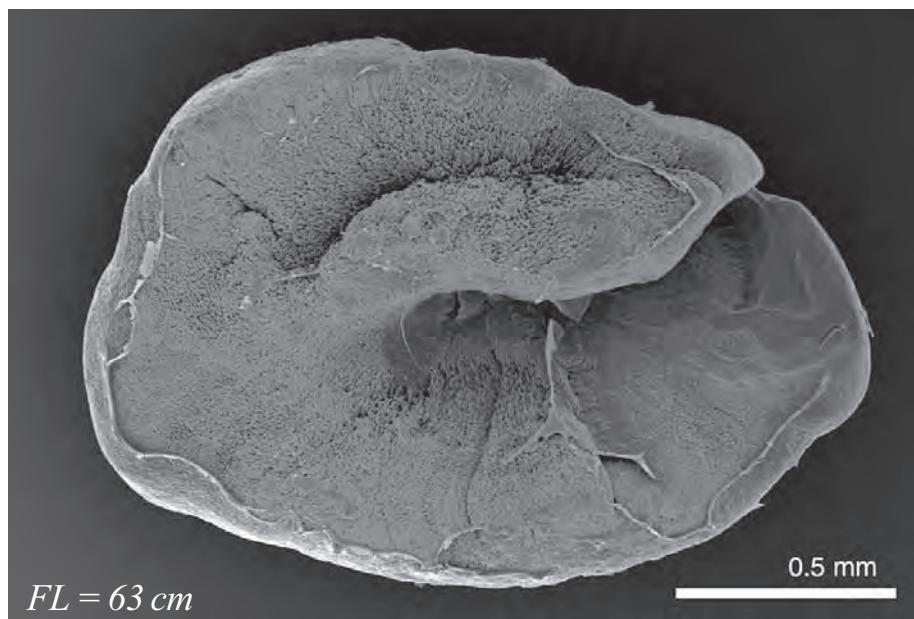
Halosauridae: *Halosauropsis macrochir*



Albuliformes

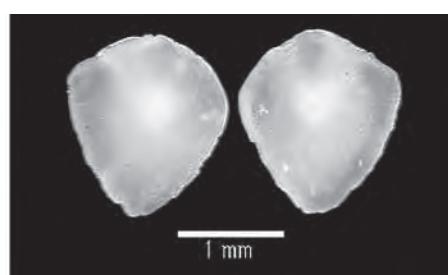
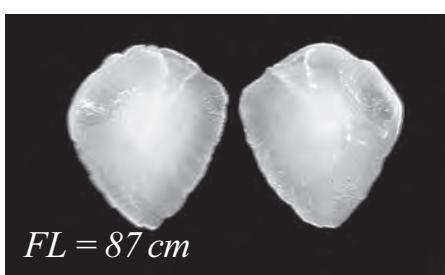
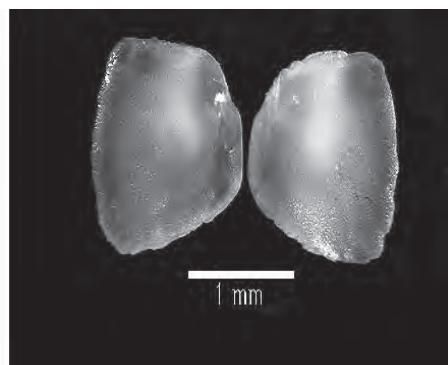
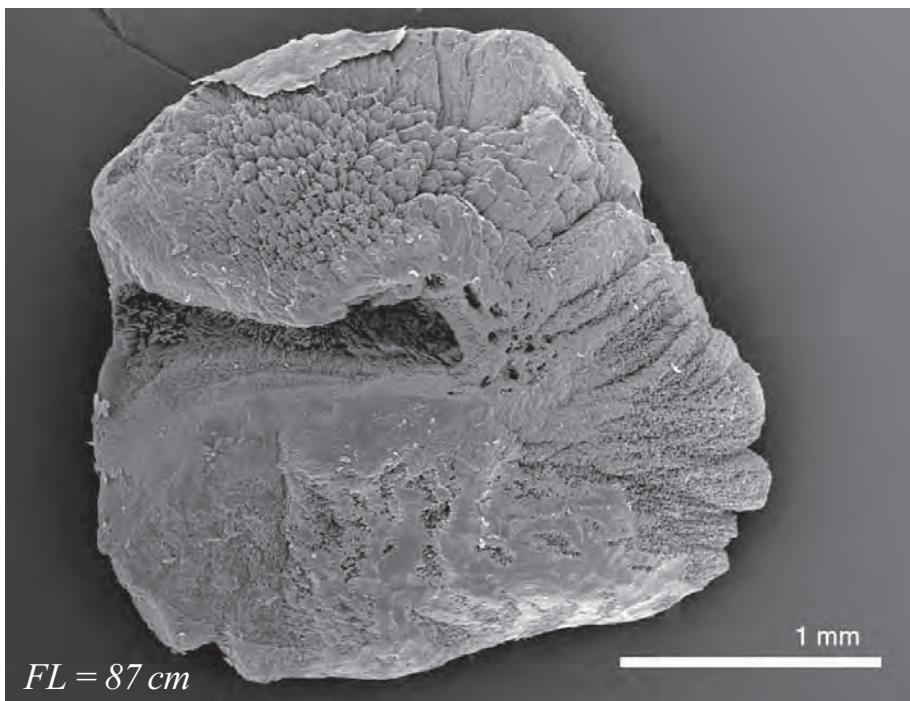
17

Notacanthidae: *Lipogenys gilli*



Albuliformes

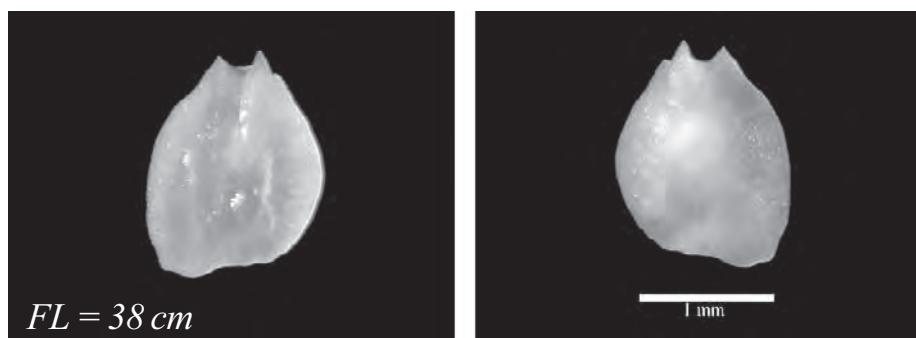
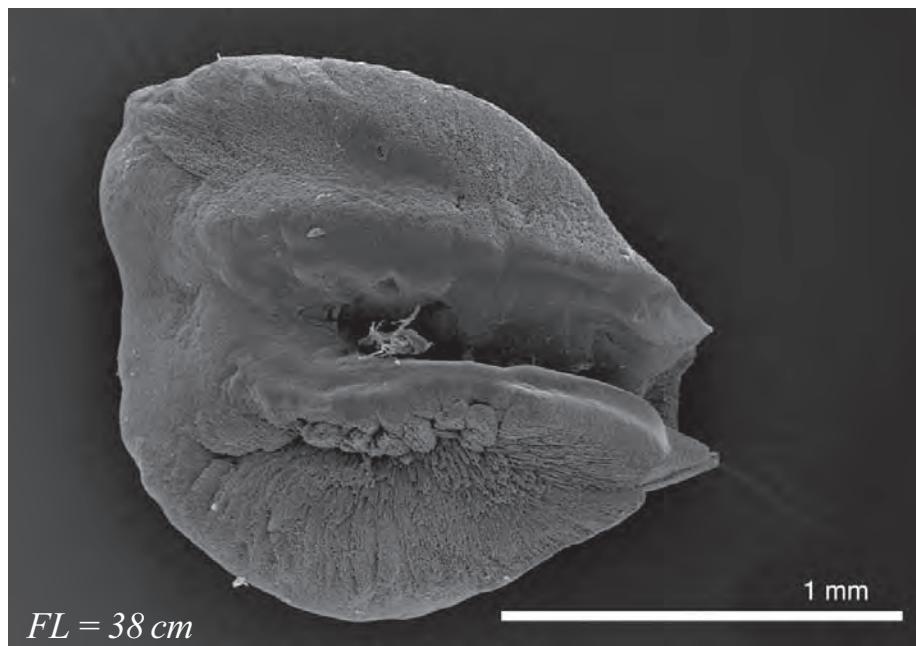
Notacanthidae: *Notacanthus chemnitzii*



Albuliformes

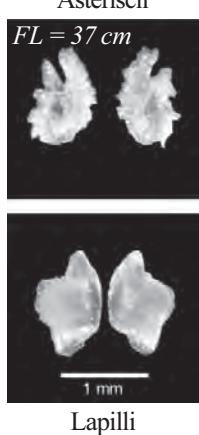
19

Notacanthidae: *Polyacanthonotus rissoanus*



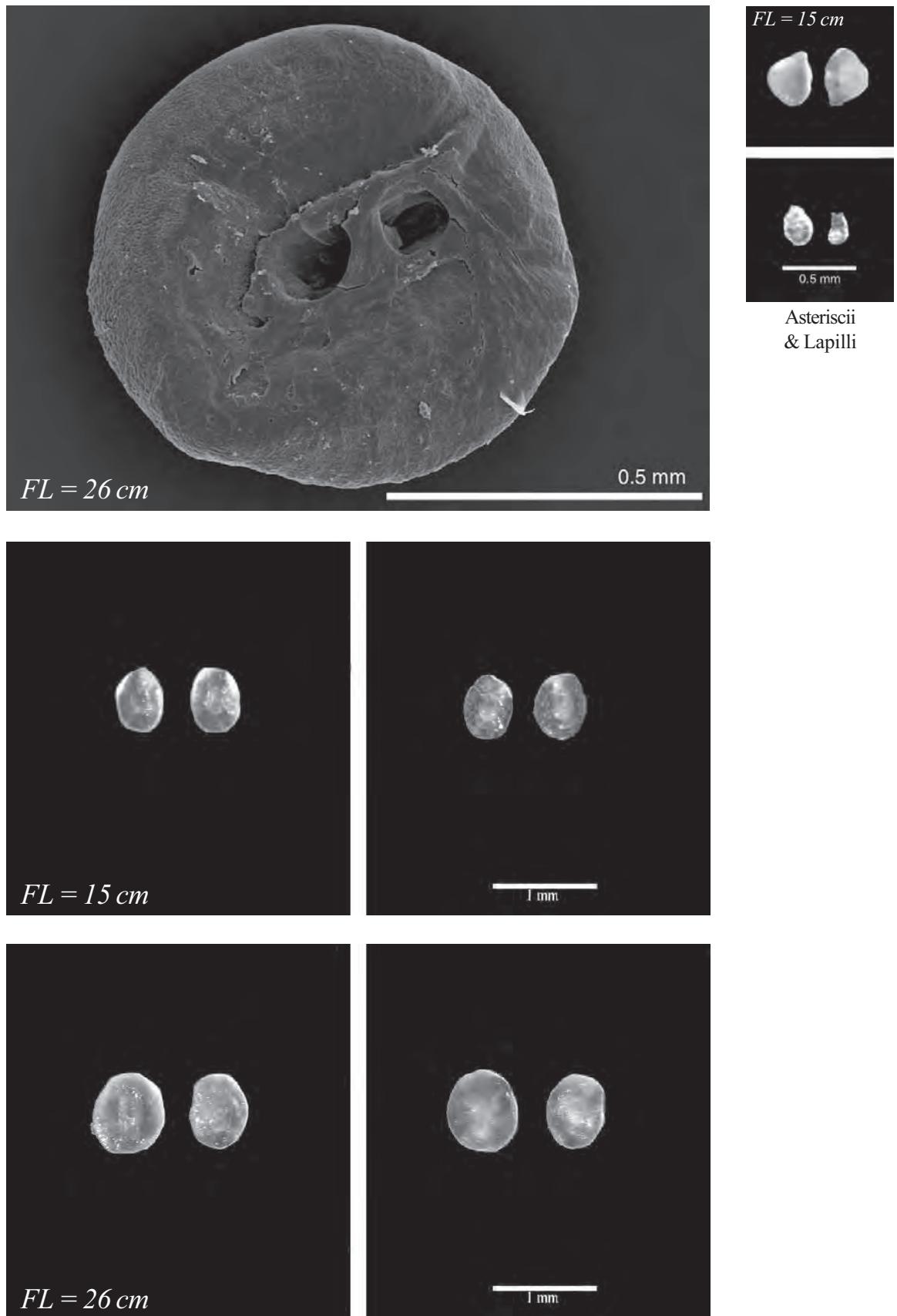
Anguilliformes

Anguillidae: *Anguilla rostrata*



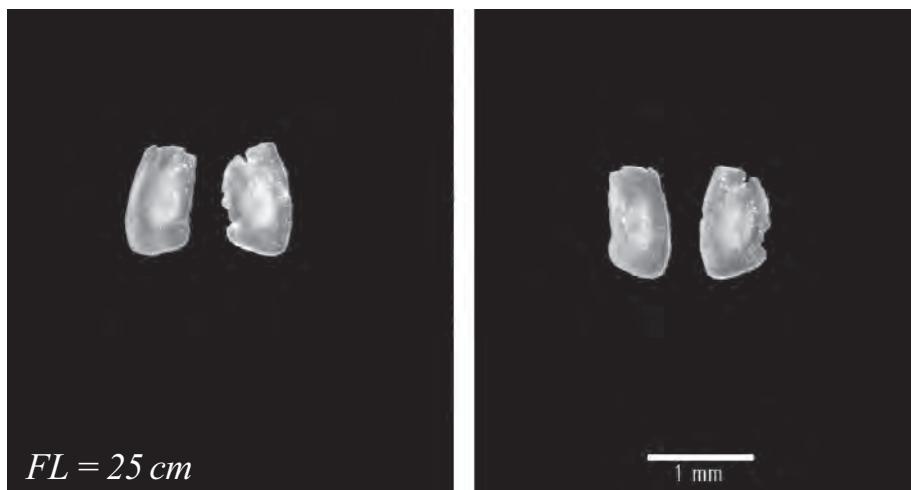
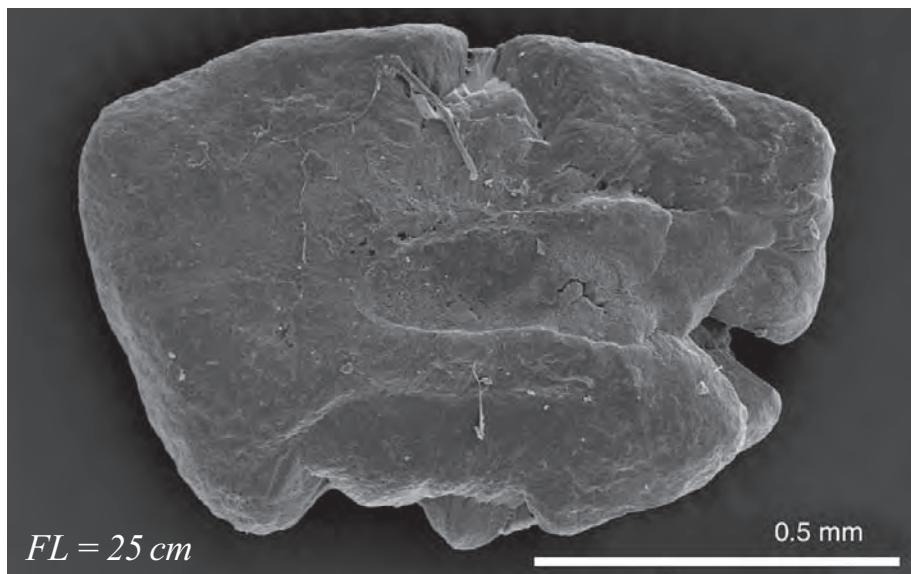
Anguilliformes

Derichthyidae: *Derichthys serpentinus*



Anguilliformes

Derichthyidae: *Nessorhamphus ingolfianus*

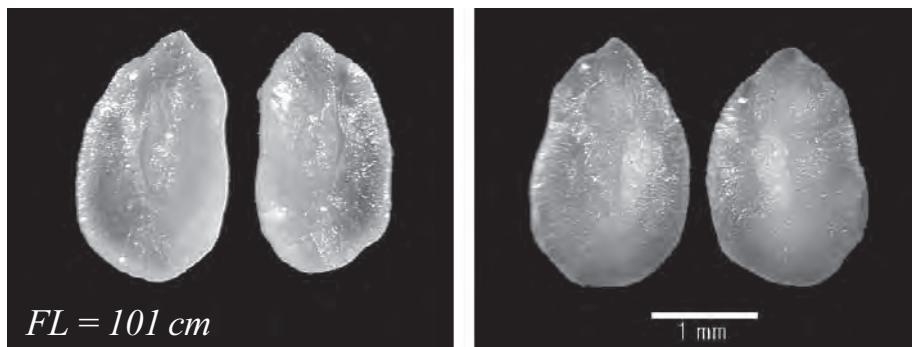
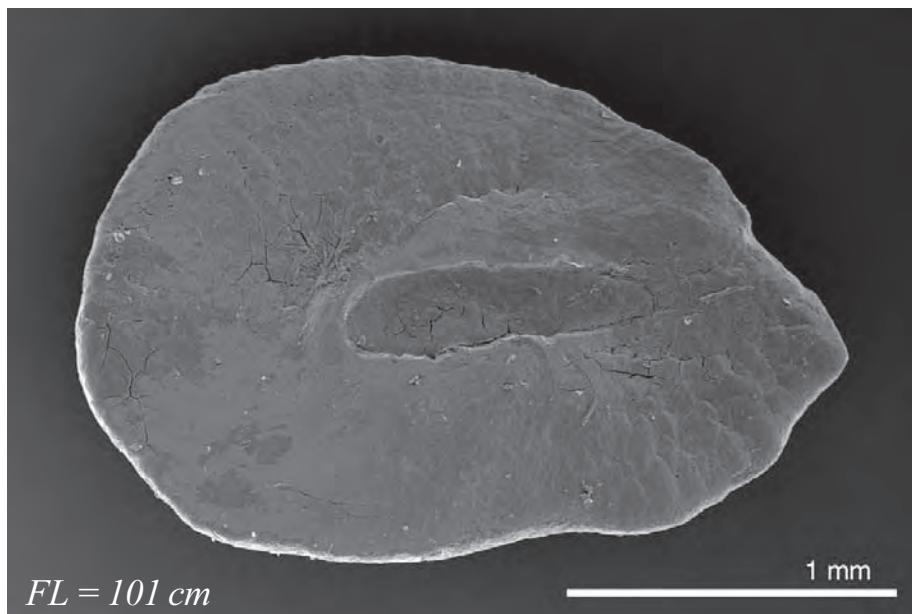


Anguilliformes
Nemichthyidae: *Nemichthys scolopaceus*



Anguilliformes

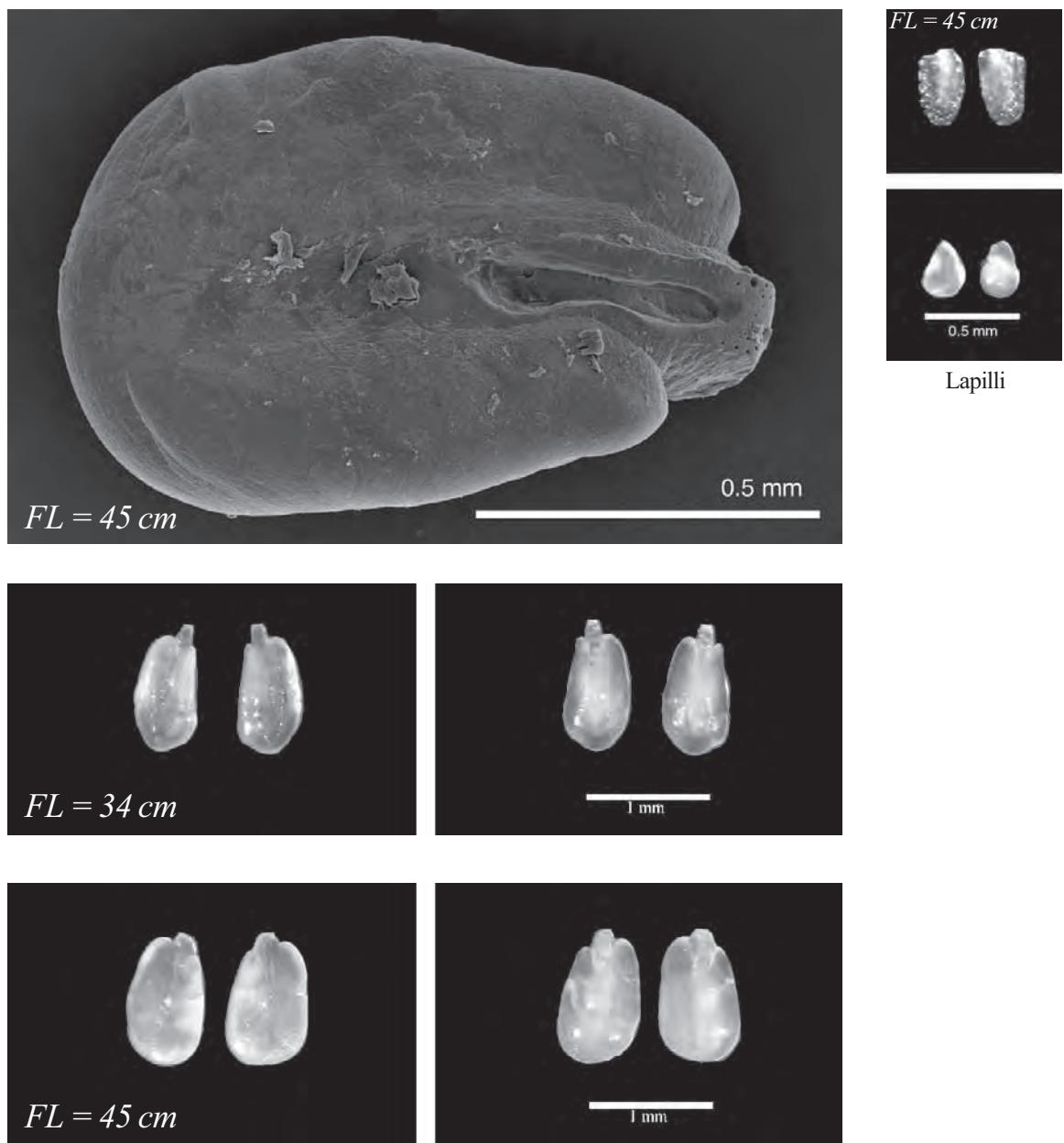
Nettastomatidae: *Venefica procera*



Anguilliformes

25

Serrivomeridae: *Serrivomer beani*



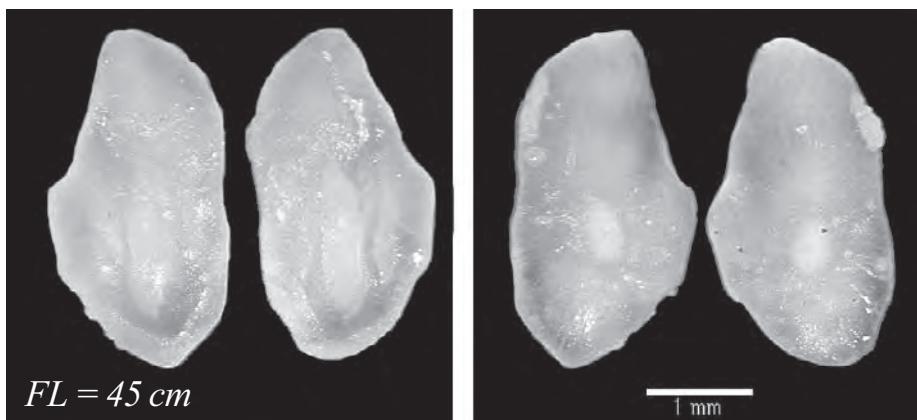
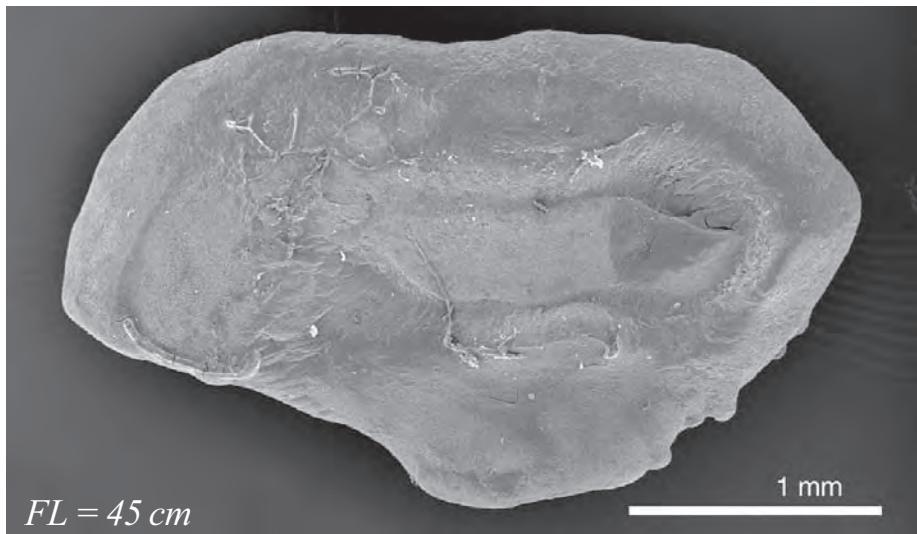
Anguilliformes

Synaphobranchidae: *Ilyophis brunneus*



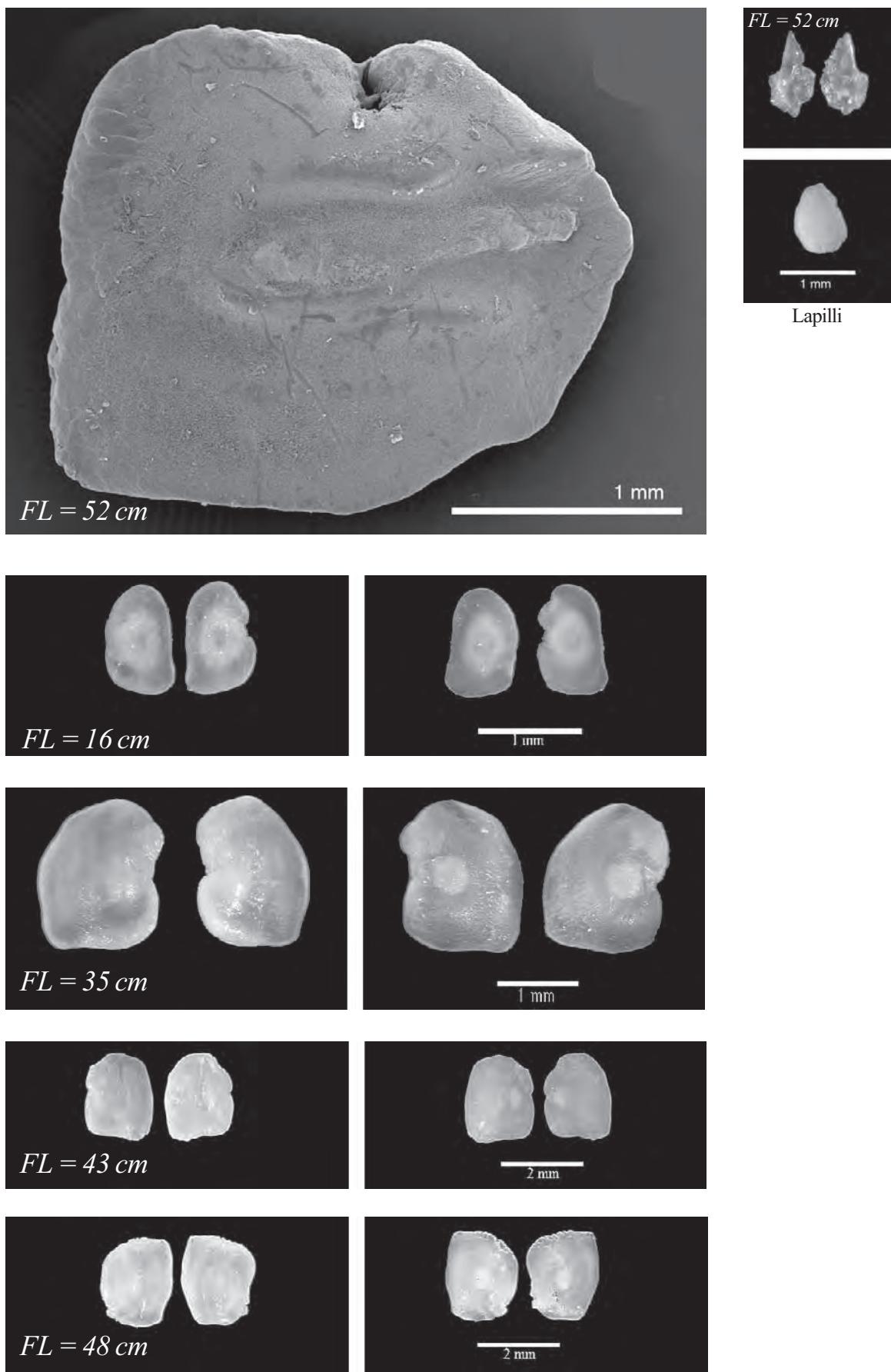
Anguilliformes

Synaphobranchidae: *Simenchelys parasiticus*



Anguilliformes

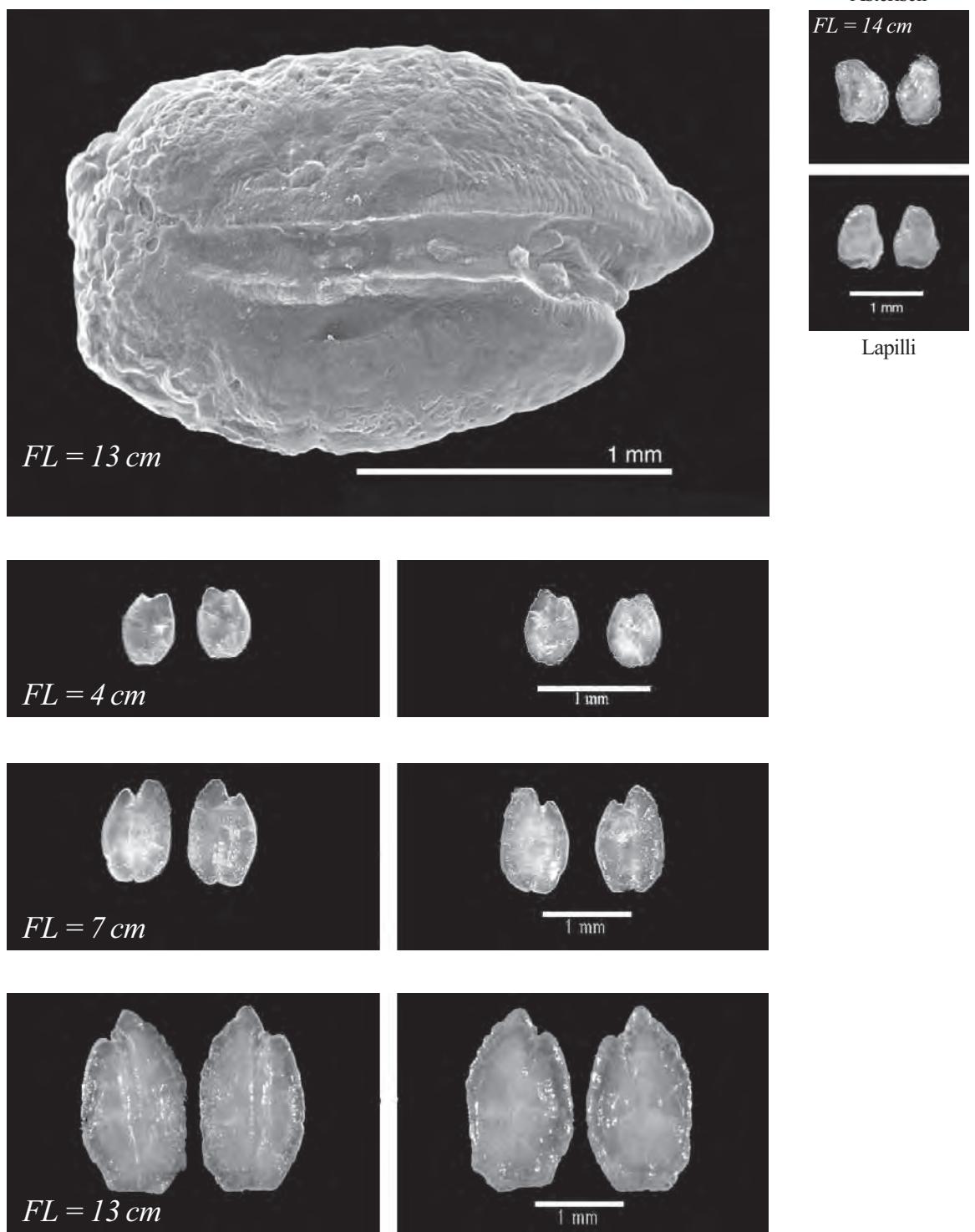
Synaphobranchidae: *Synaphobranchus kaupi*



Atheriniformes

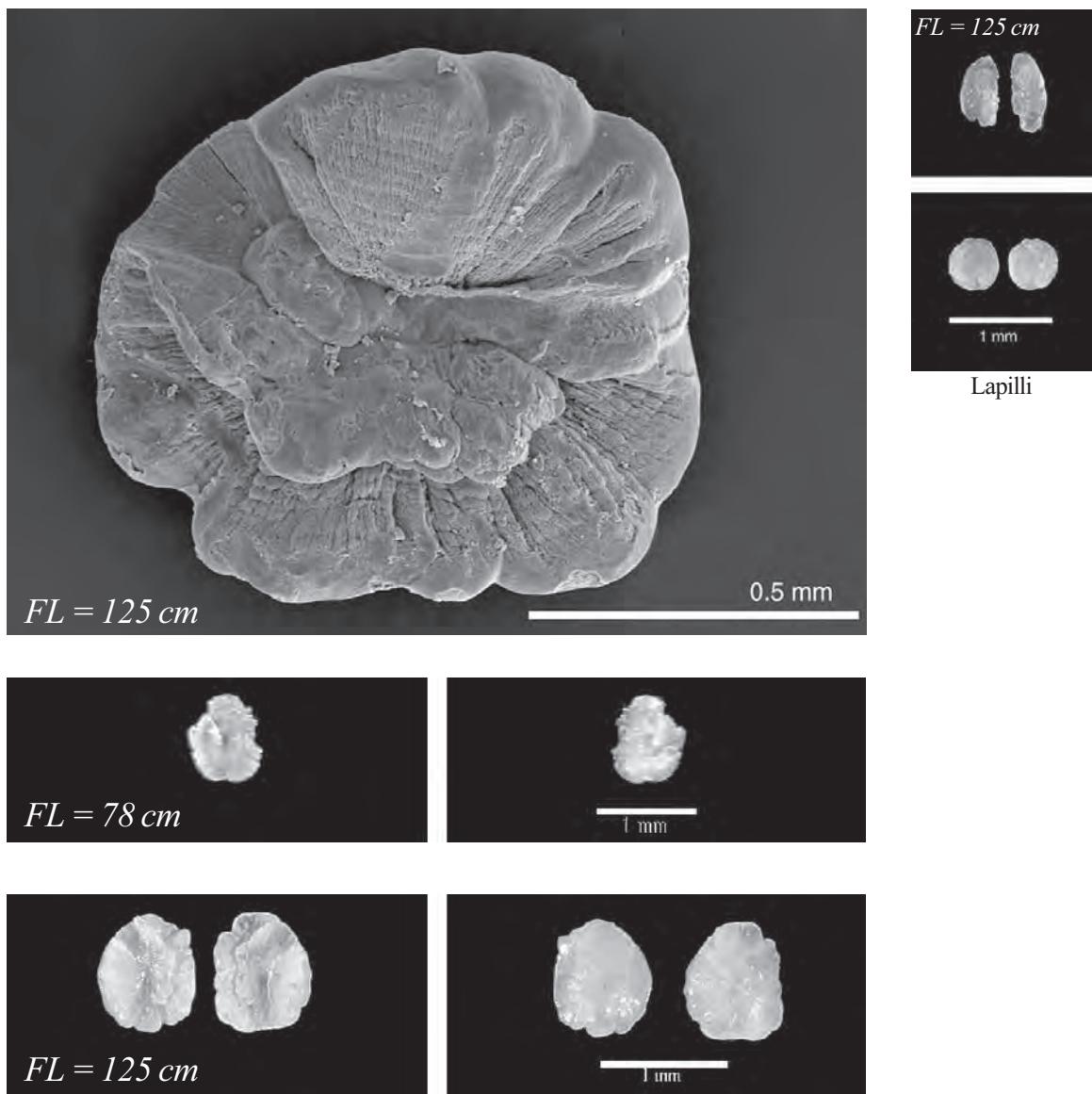
Atherinidae: *Menidia menidia*

29



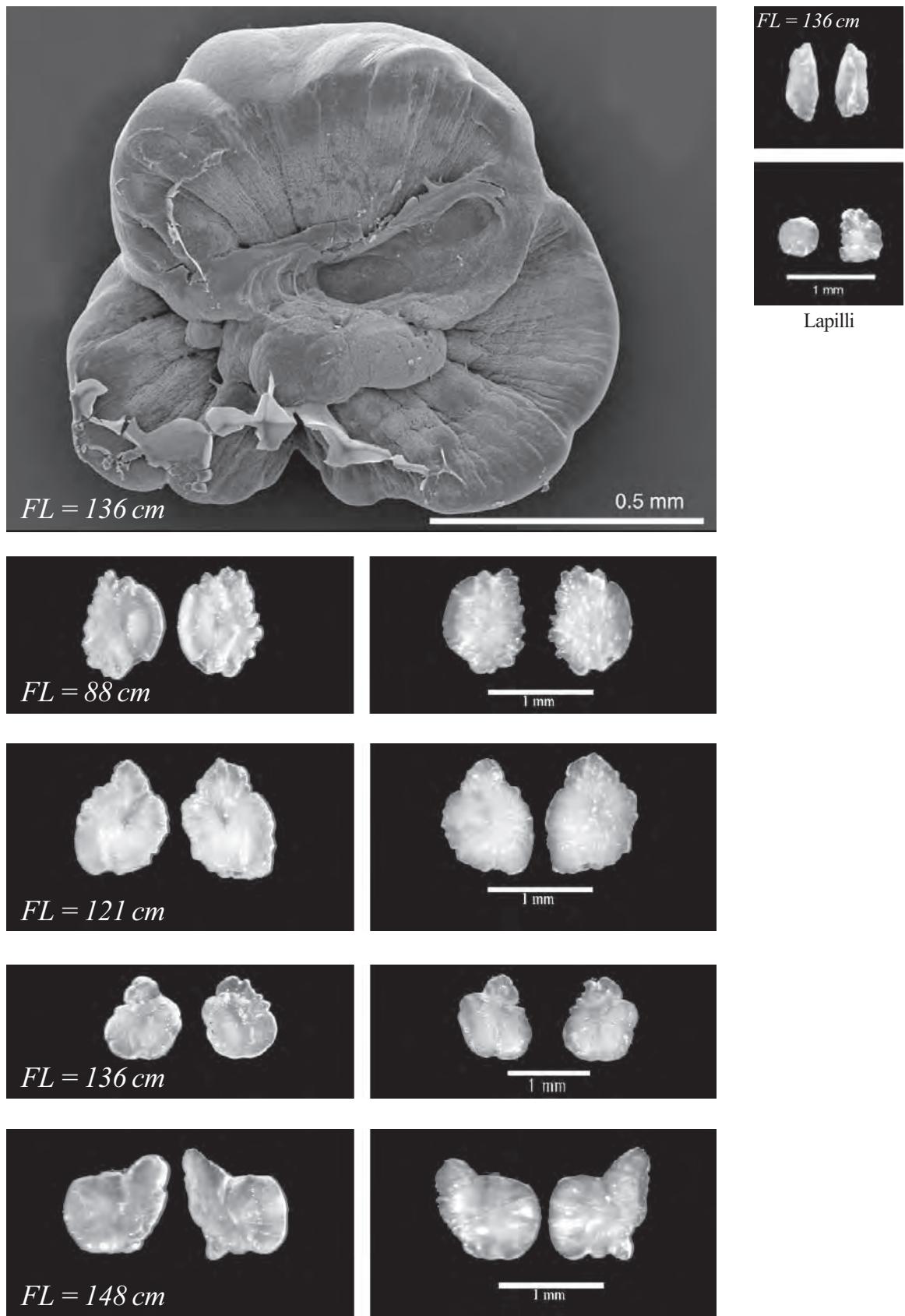
Aulopiformes

Alepisauridae: *Alepisaurus brevirostris*



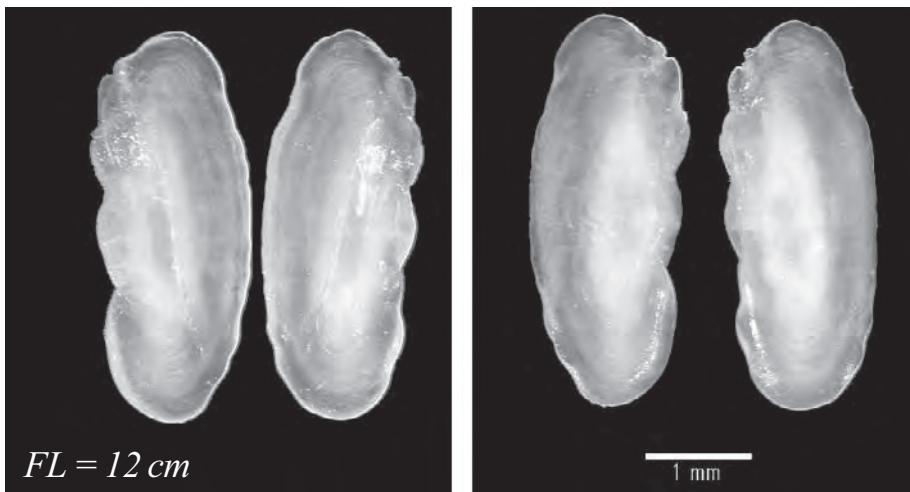
Aulopiformes

Alepisauridae: *Alepisaurus ferox*



Aulopiformes

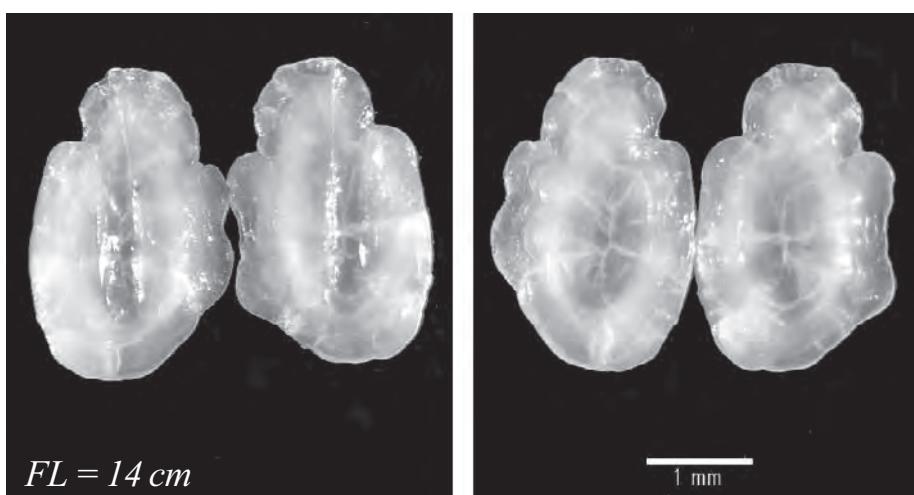
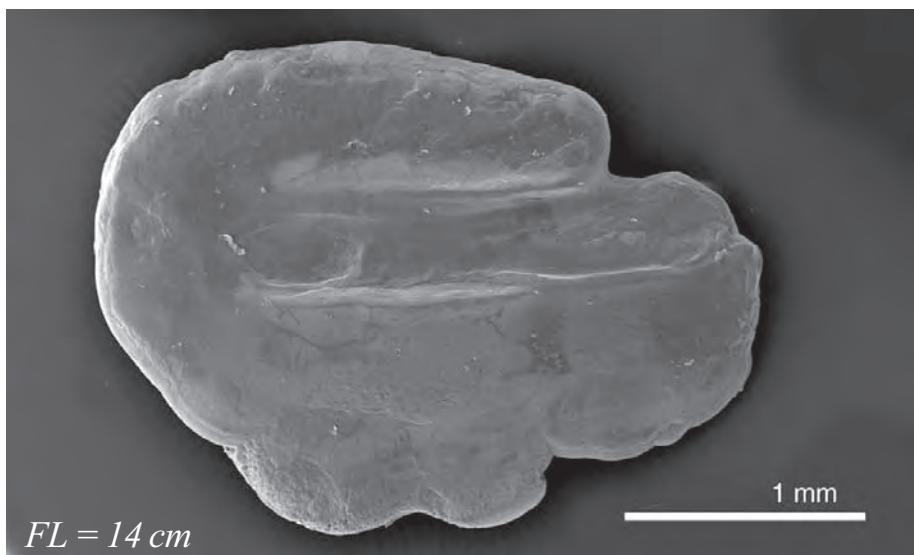
Chlorophthalmidae: *Chlorophthalmus agassizi*



Aulopiformes

33

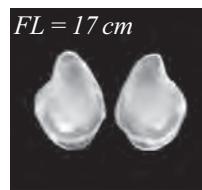
Chlorophthalmidae: *Parasudis truculenta*



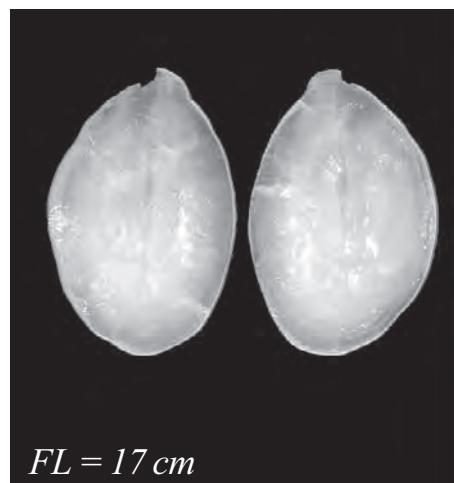
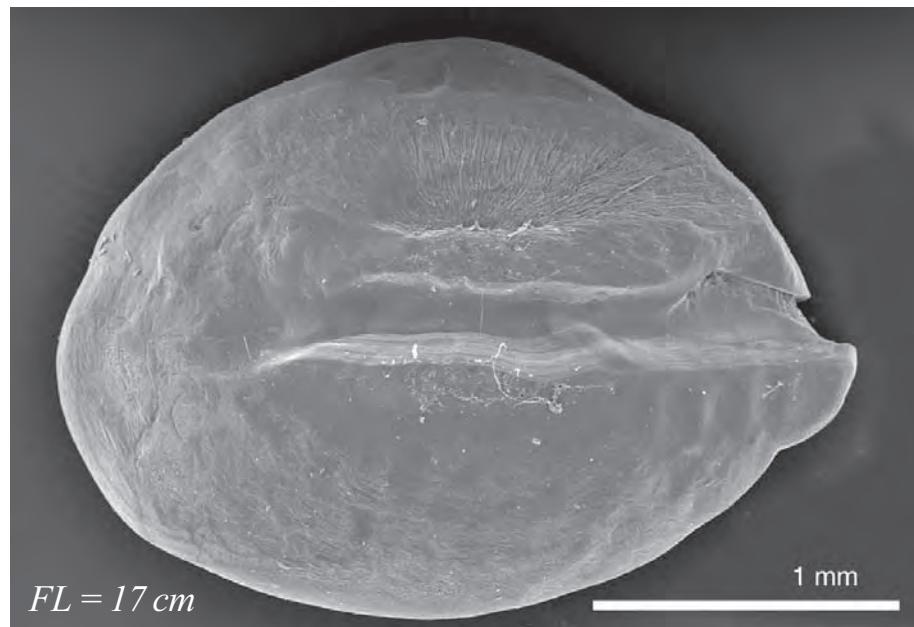
Aulopiformes

Ipnopidae: *Bathypterois quadrifilis*

Asterisci



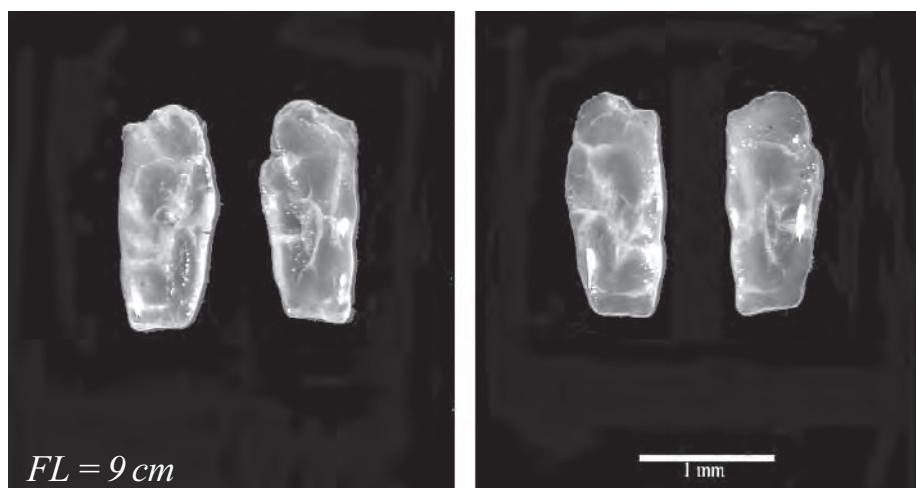
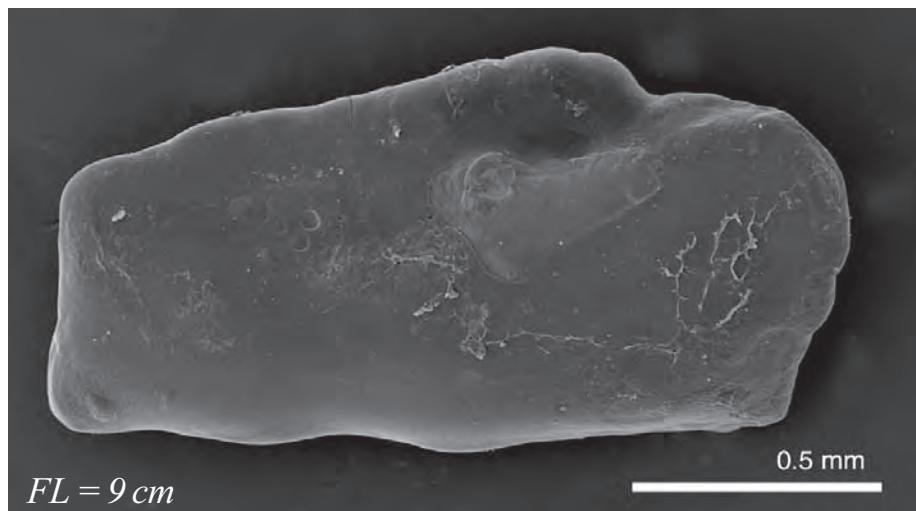
Lapilli



Aulopiformes

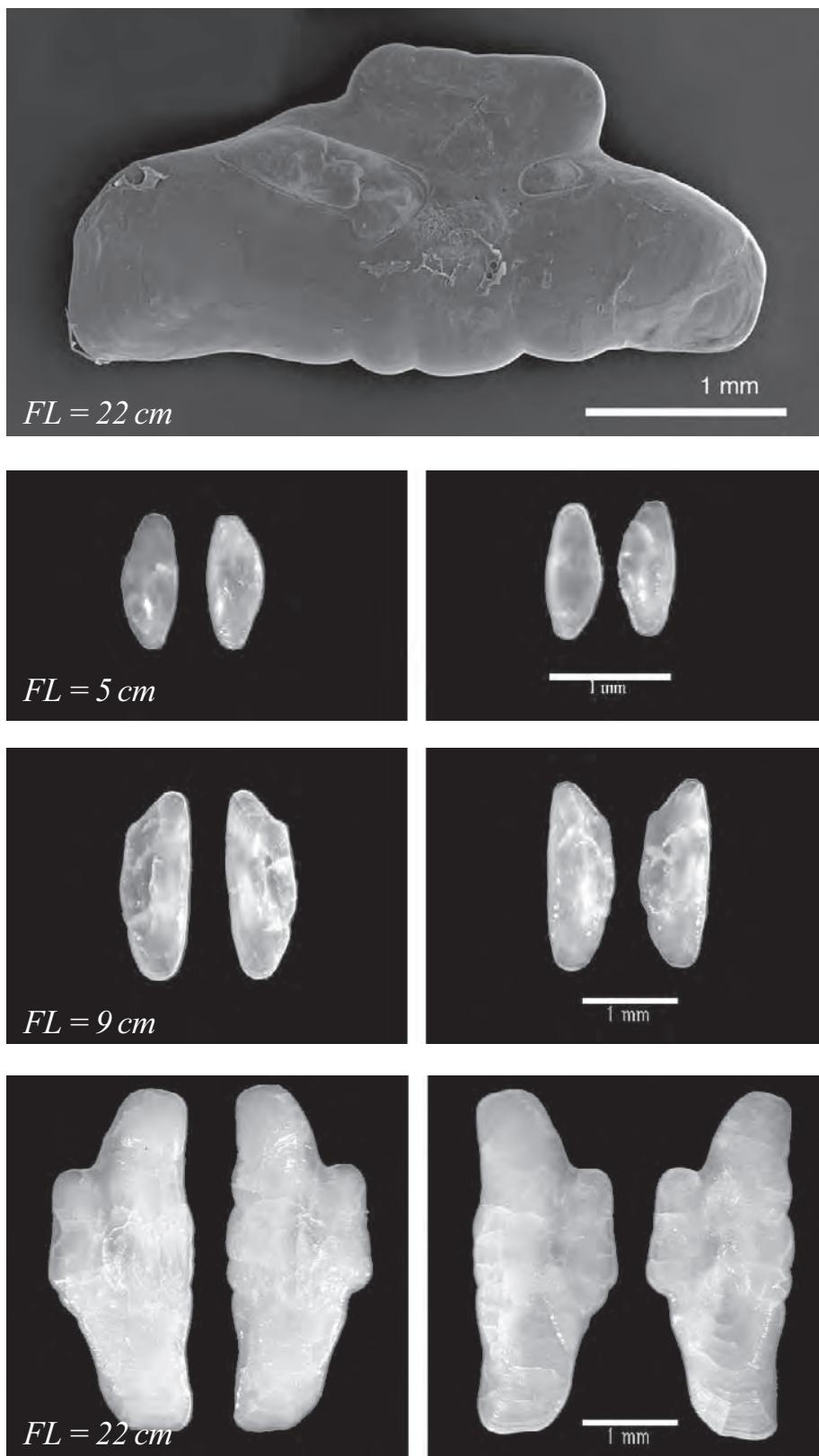
35

Paralepididae: *Lestidiops affinis*



Aulopiformes

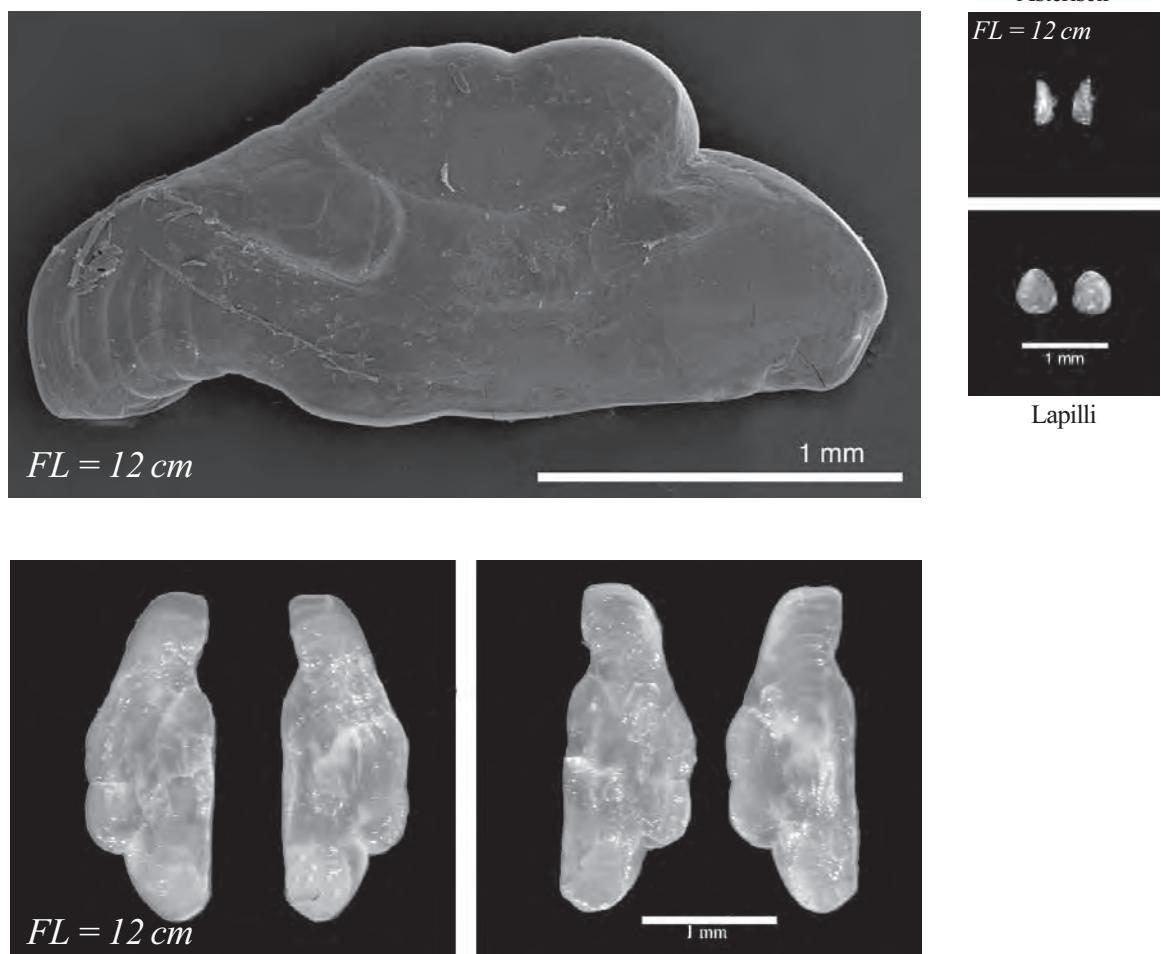
Paralepididae: *Notolepis rissoii*



Aulopiformes

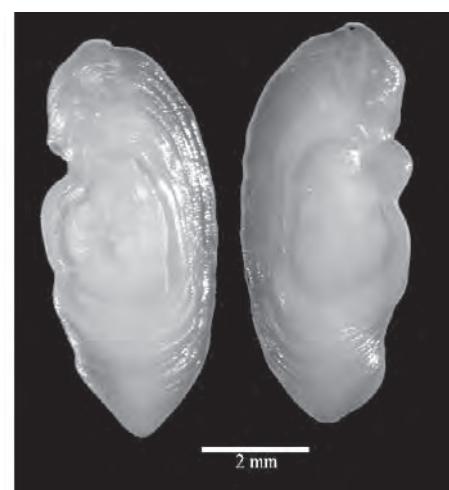
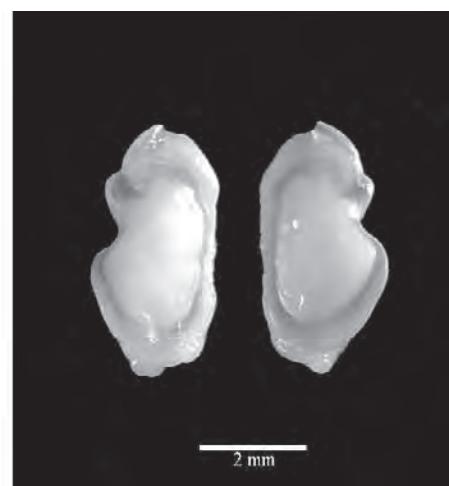
Paralepididae: *Paralepis atlantica*

37



Aulopiformes

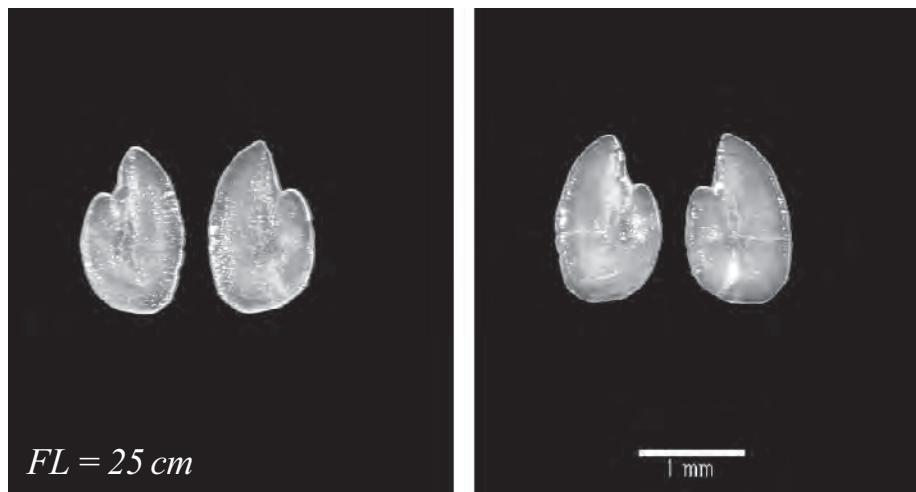
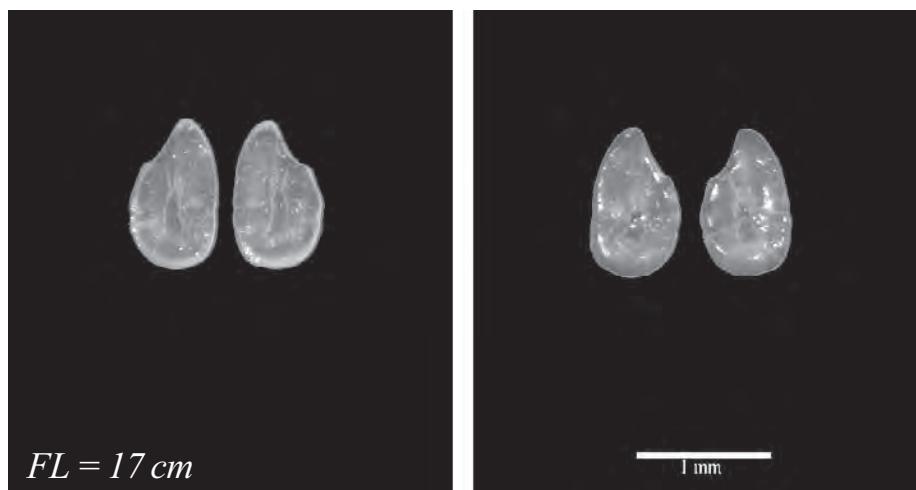
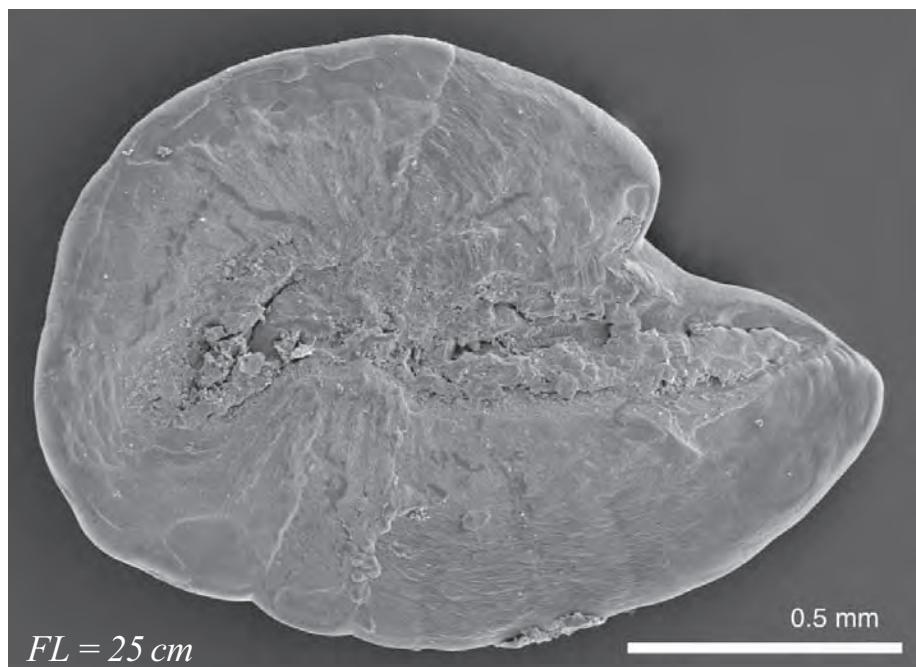
Synodontidae: *Bathysaurus ferox*



Beloniformes

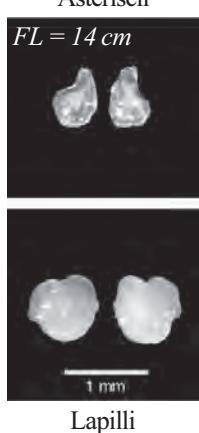
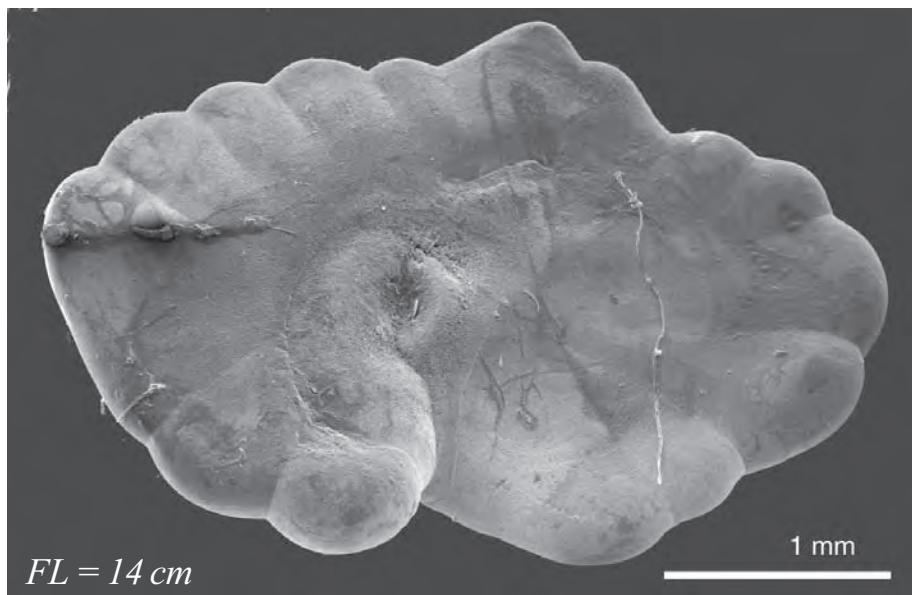
39

Scomberesocidae: *Scomberesox saurus*

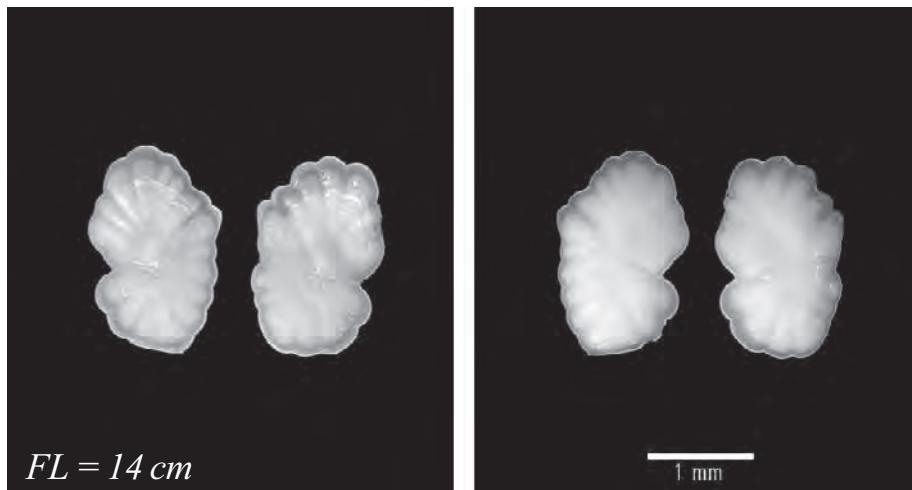


Beryciformes

Anoplogasteridae: *Anoplogaster cornuta*

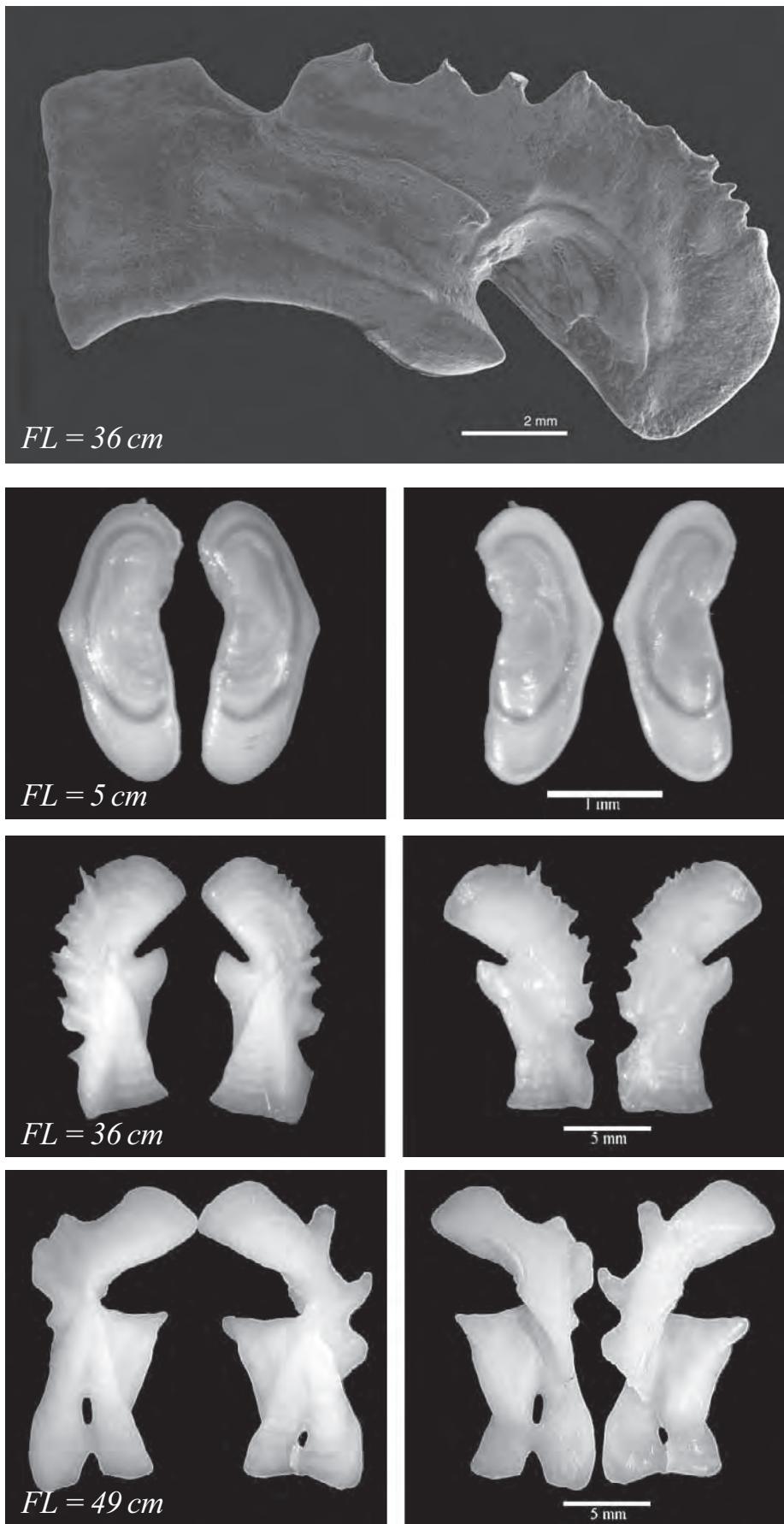


Lapilli



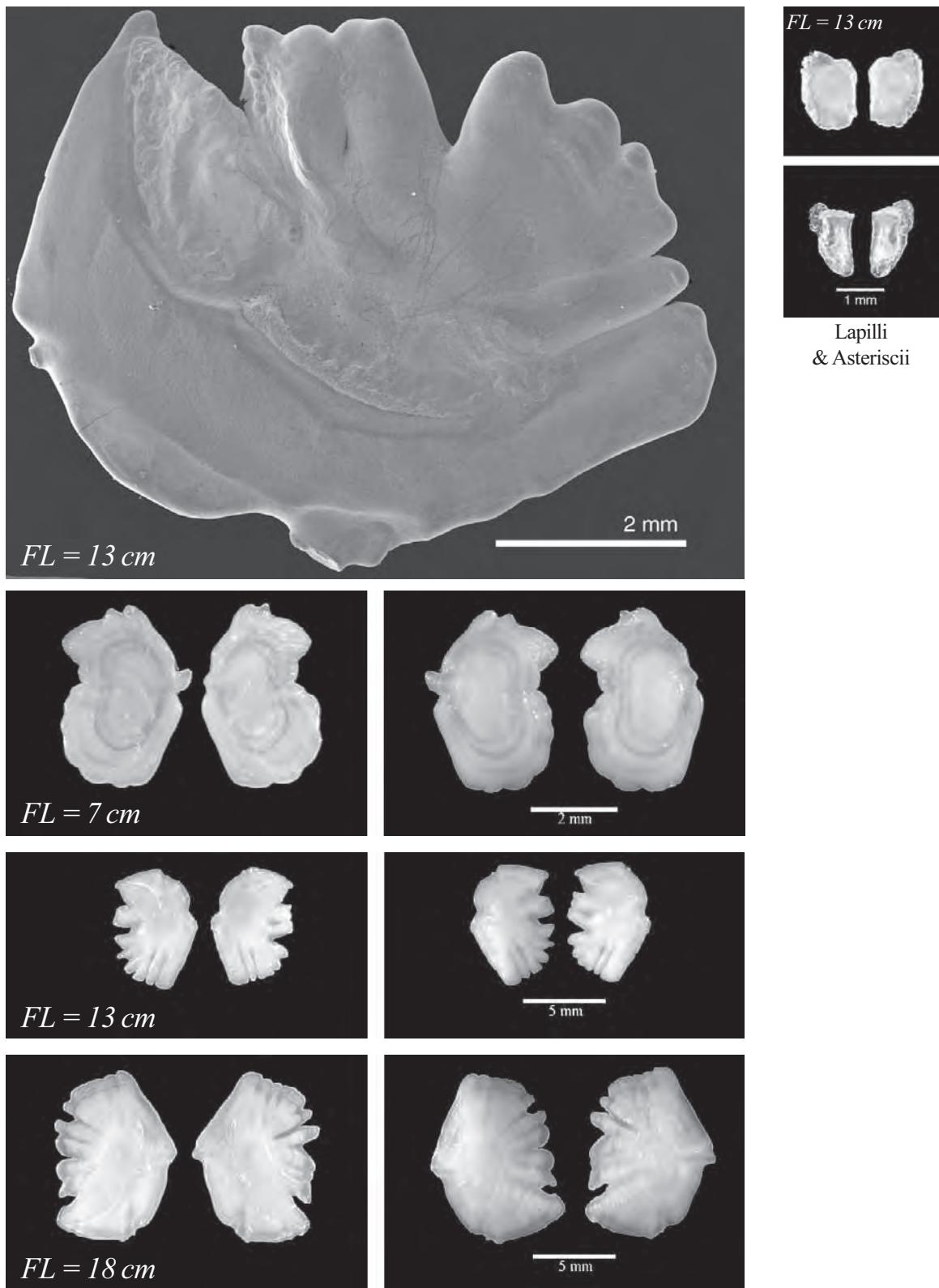
Beryciformes

Trachichthyidae: *Hoplostethus atlanticus*



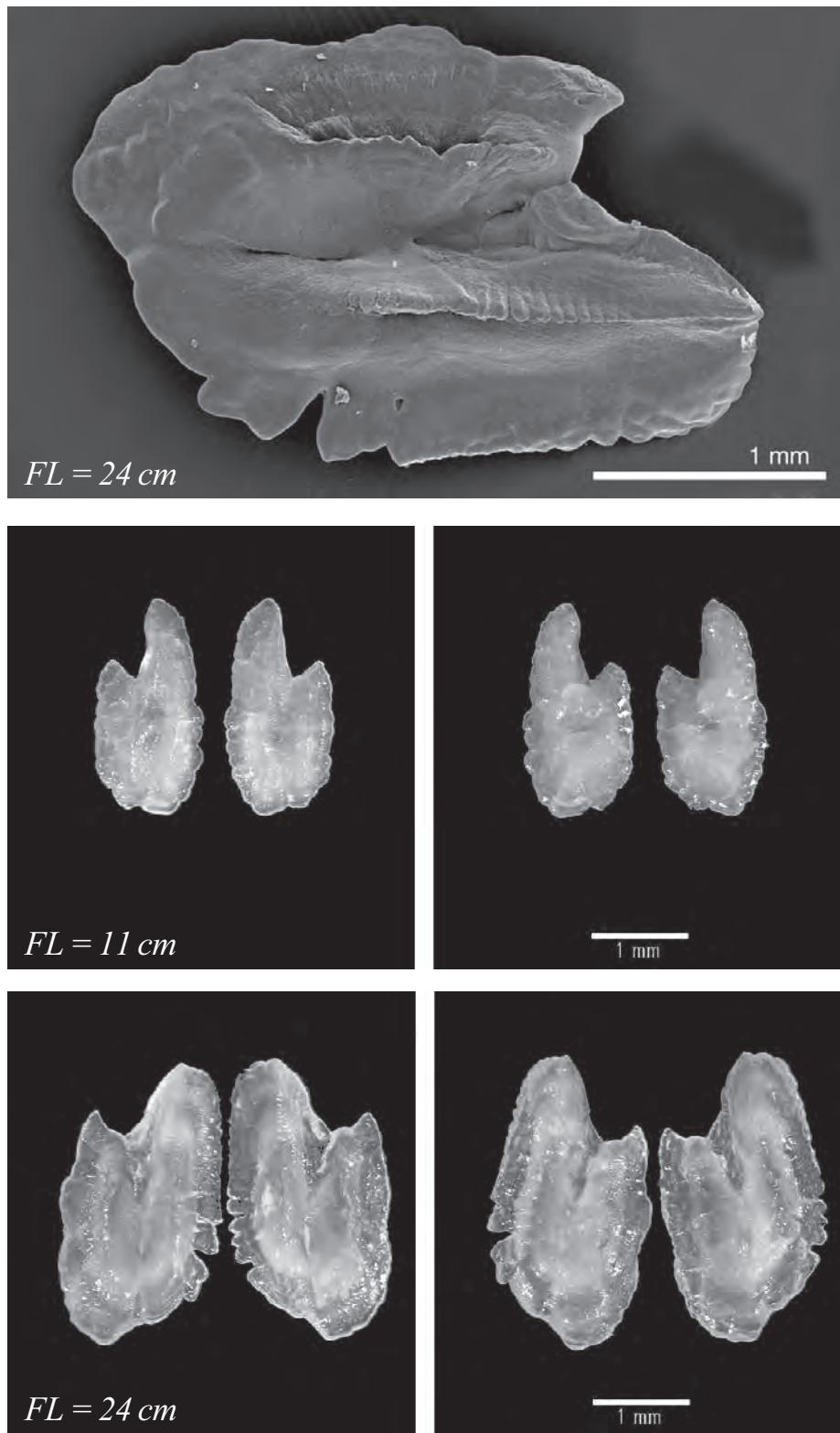
Beryciformes

Trachichthyidae: *Hoplostethus mediterraneus*



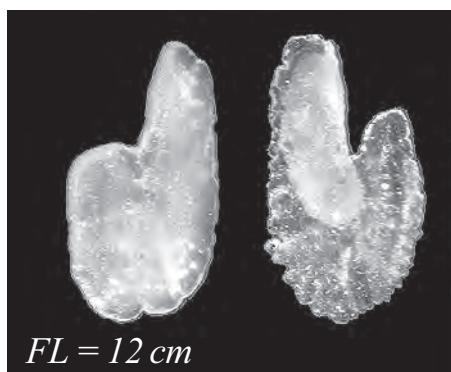
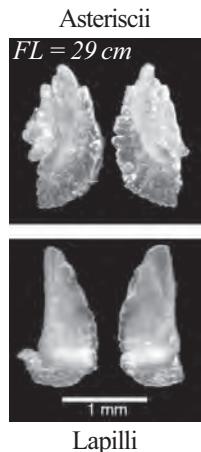
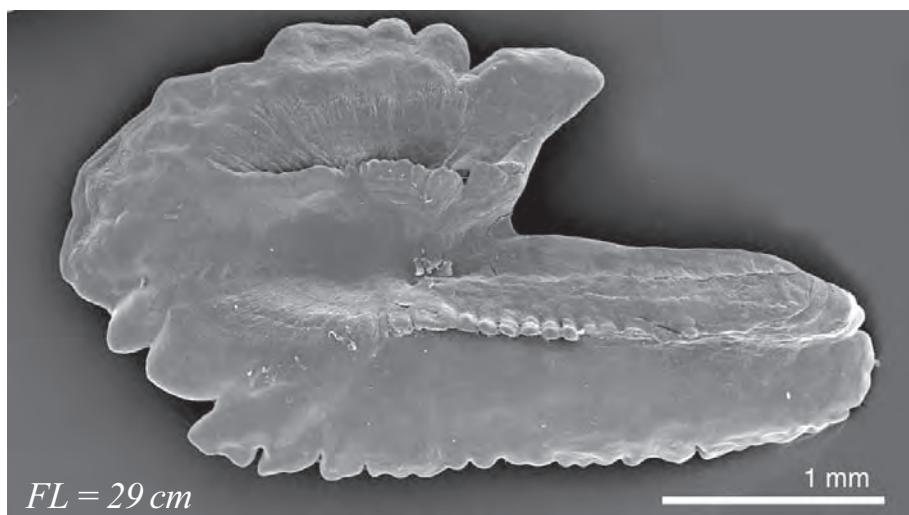
Clupeiformes

Clupeidae: *Alosa aestivalis*



Clupeiformes

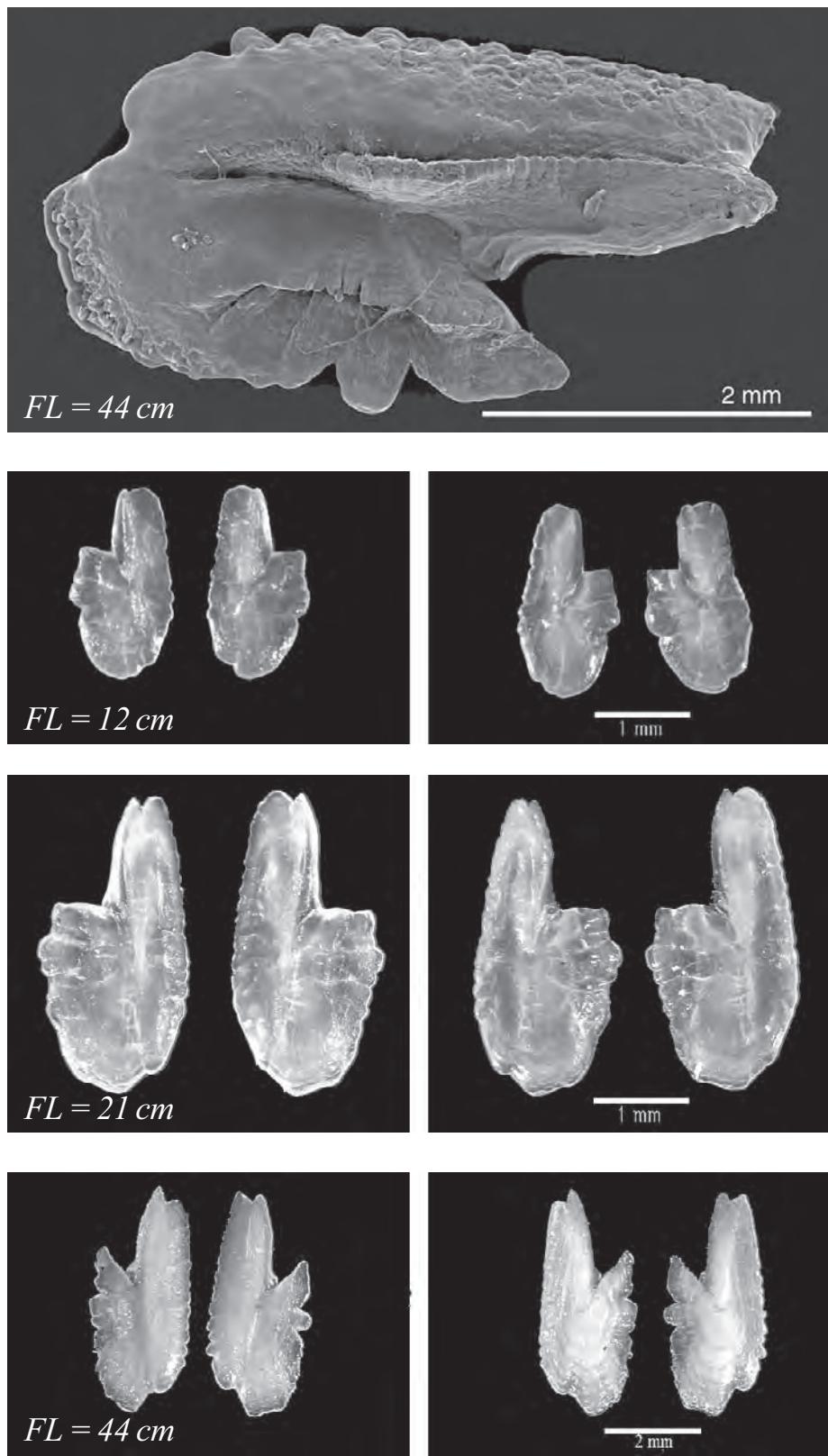
Clupeidae: *Alosa pseudoharengus*



Clupeiformes

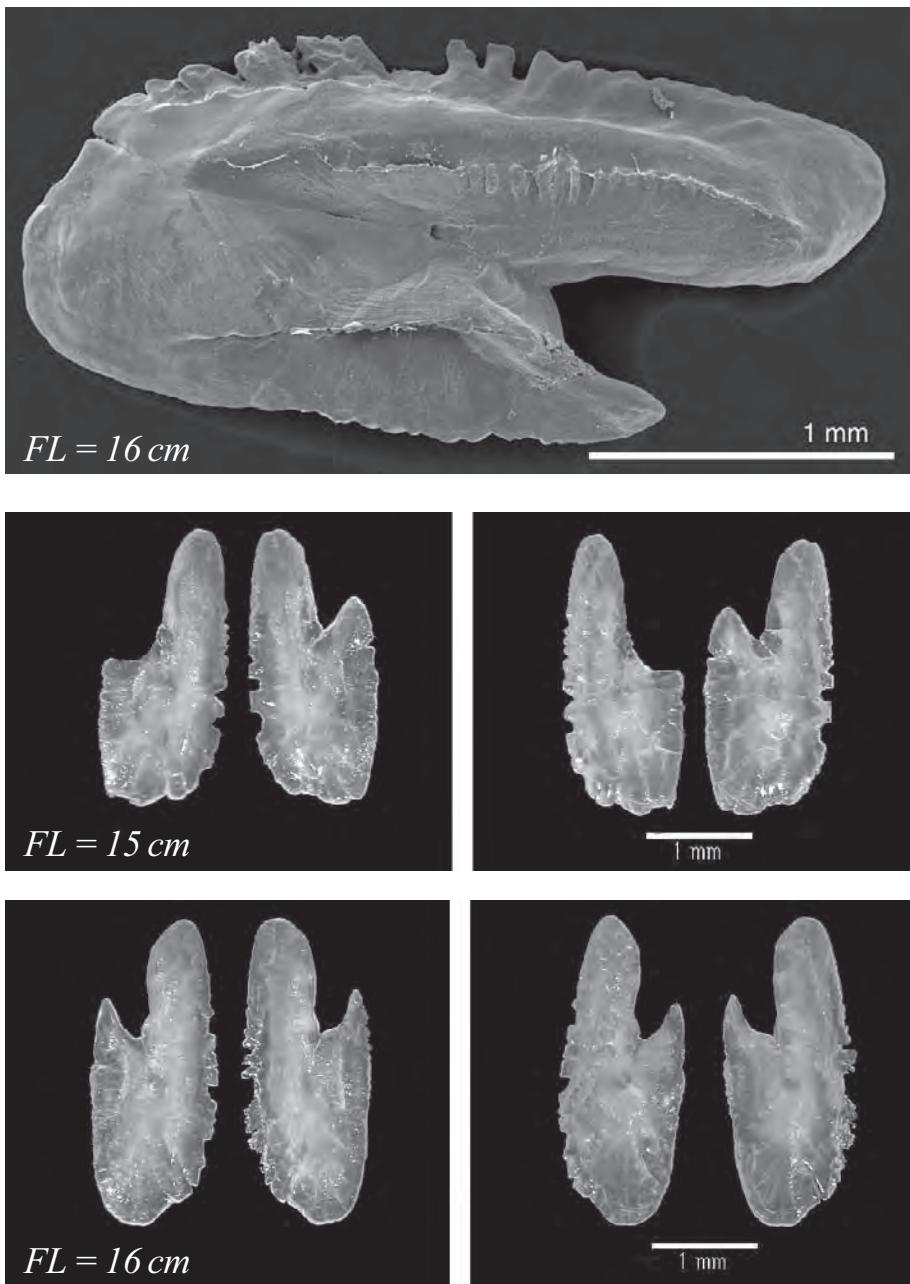
Clupeidae: *Alosa sapidissima*

45



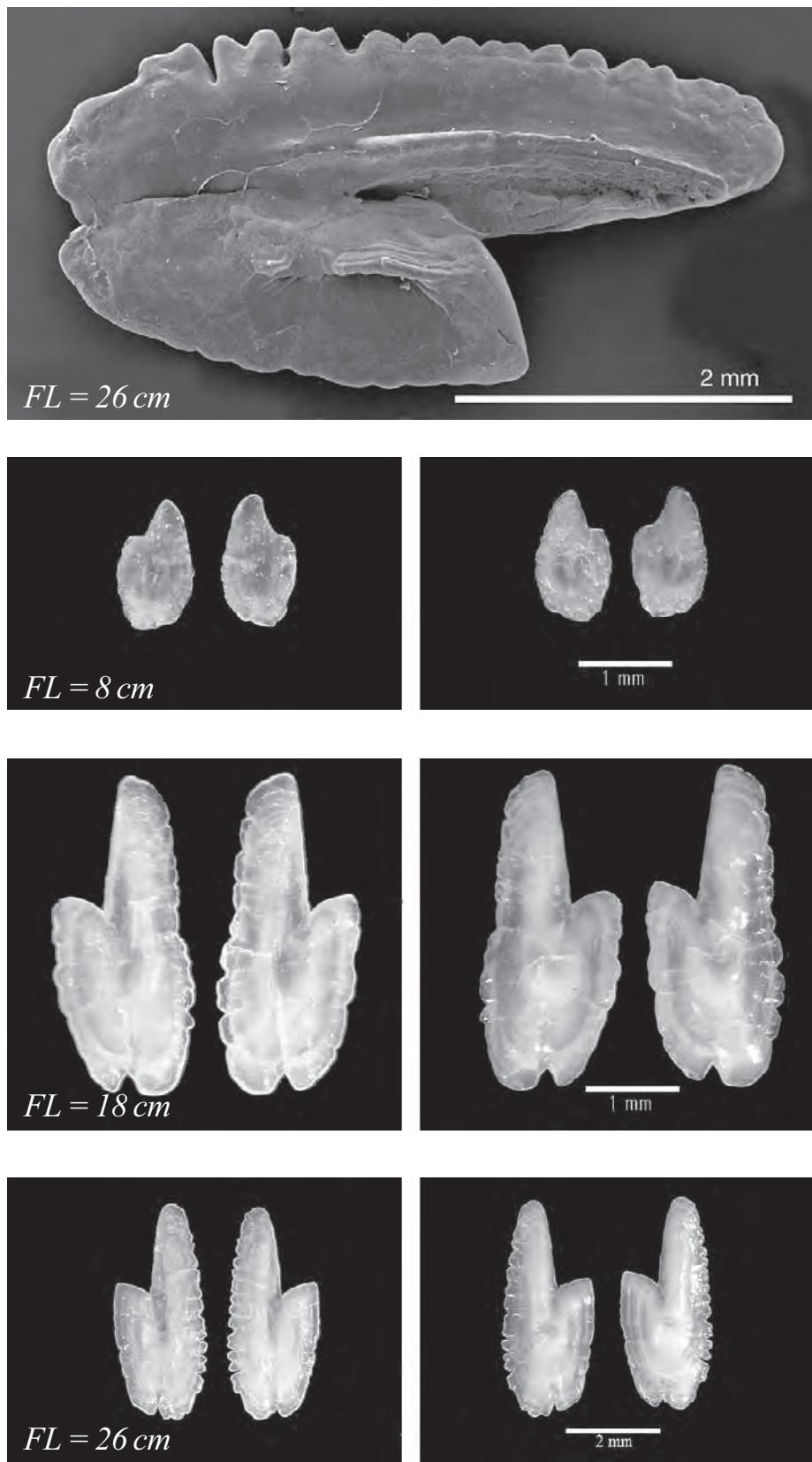
Clupeiformes

Clupeidae: *Brevoortia tyrannus*



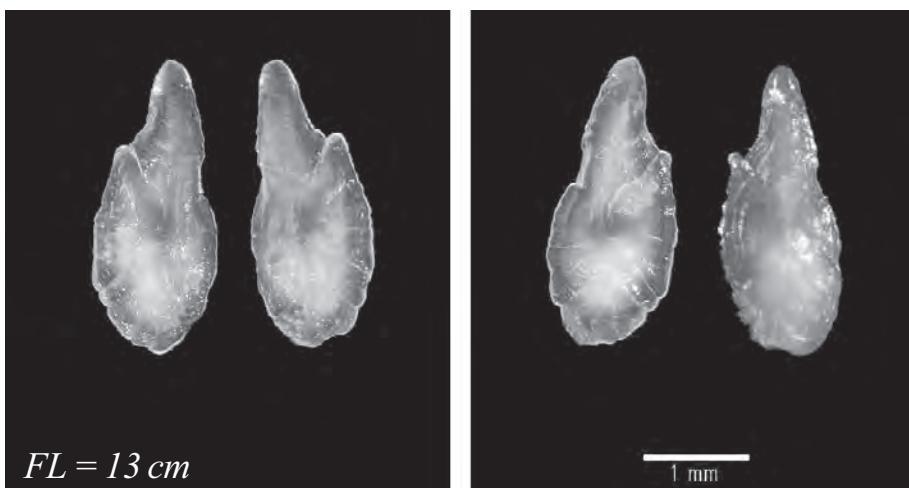
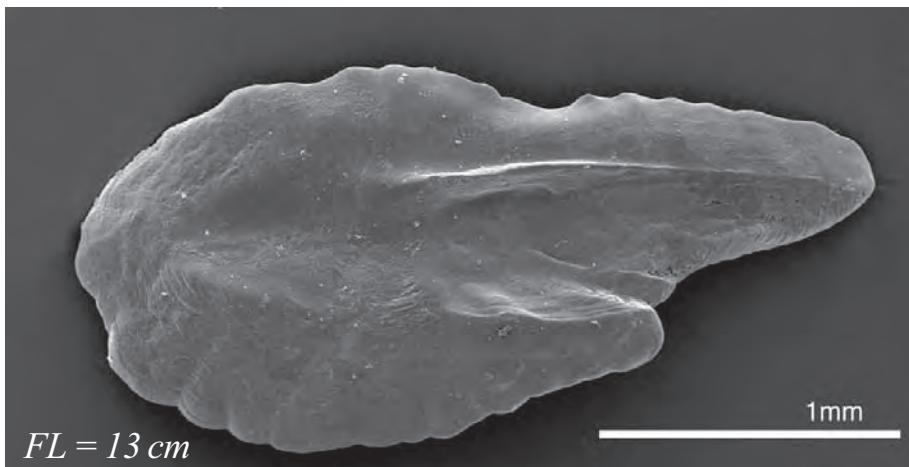
Clupeiformes

Clupeidae: *Clupea harengus* *harengus*



Clupeiformes

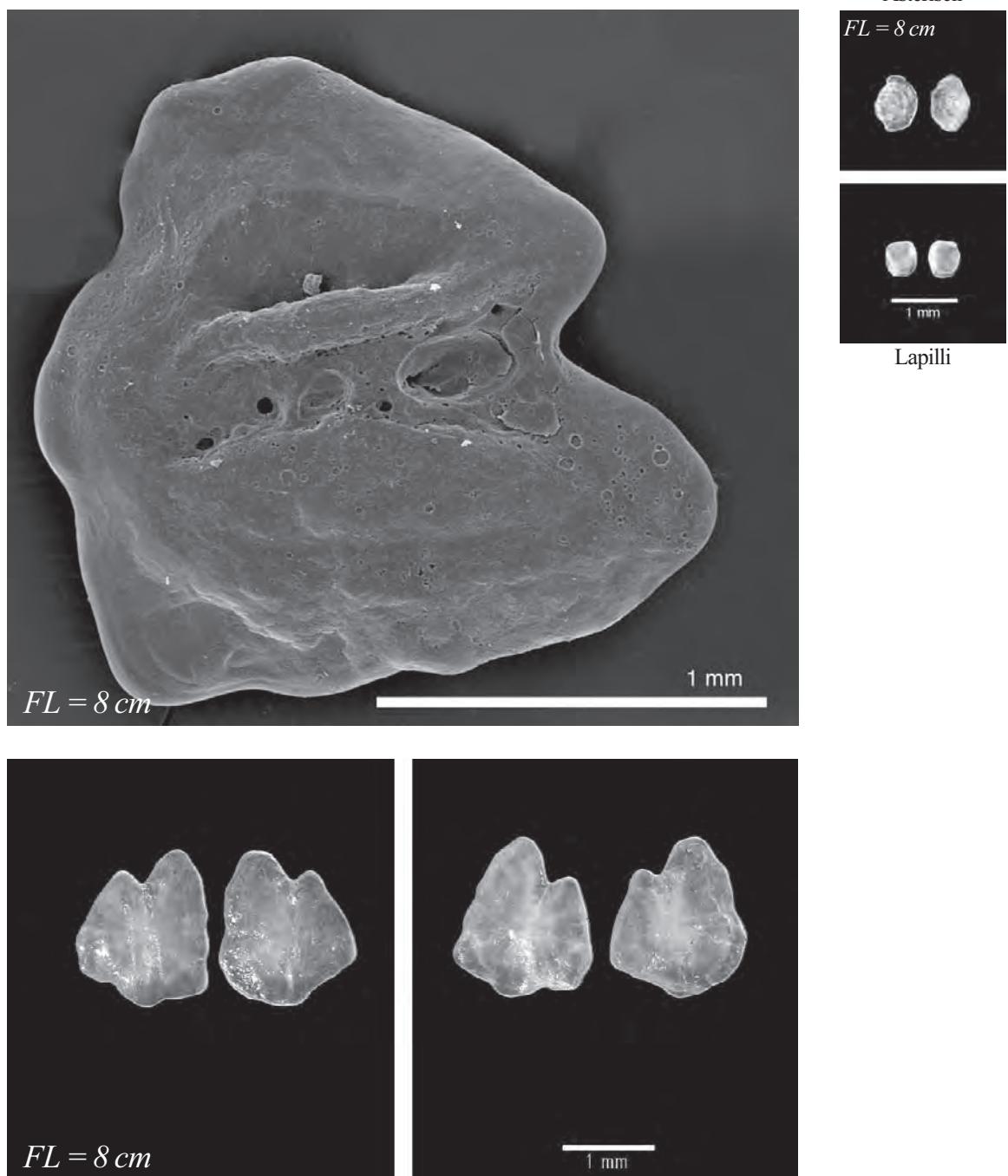
Clupeidae: *Etrumeus teres*



Cyprinodontiformes

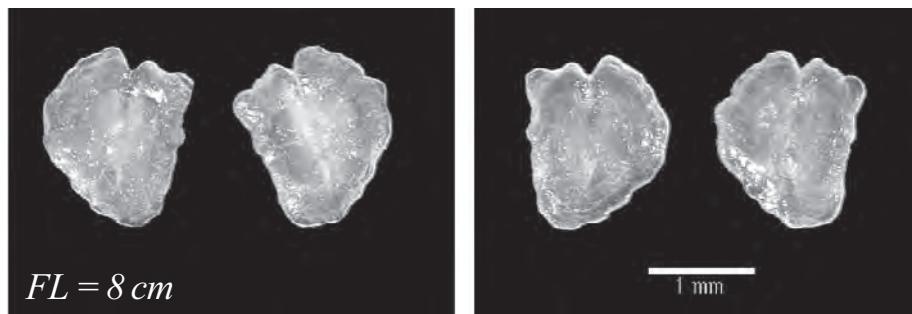
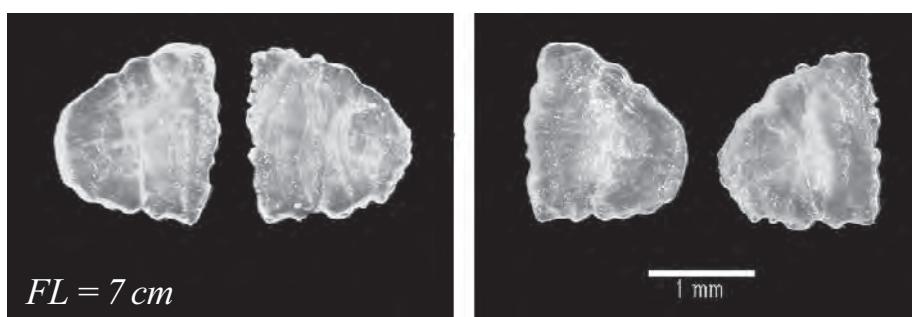
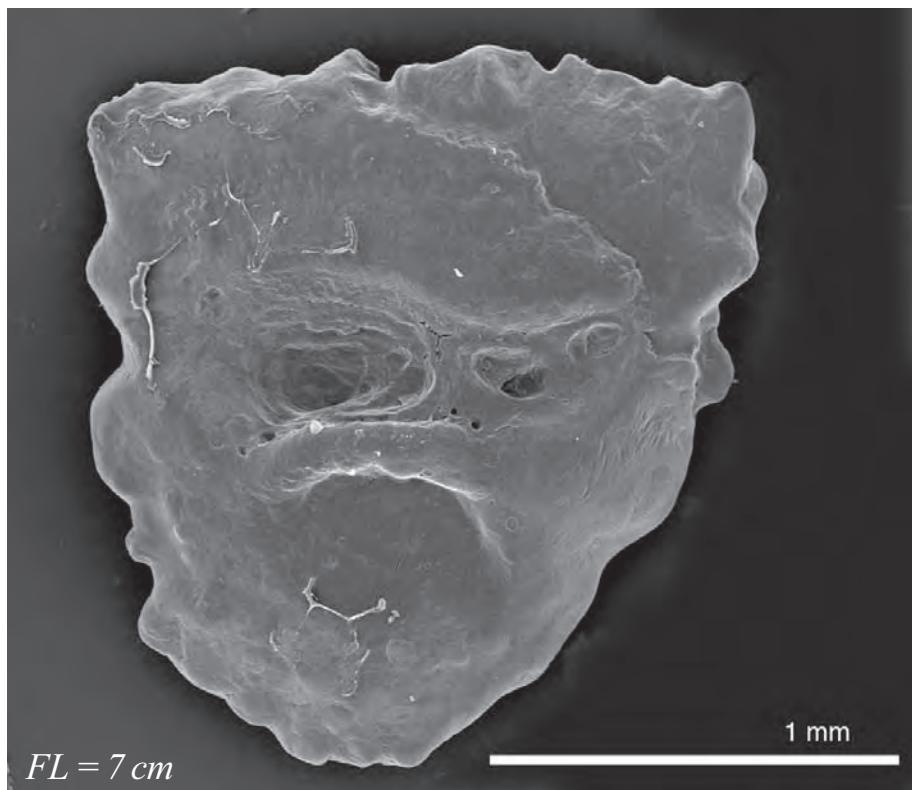
Fundulidae: *Fundulus diaphanus*

49



Cyprinodontiformes

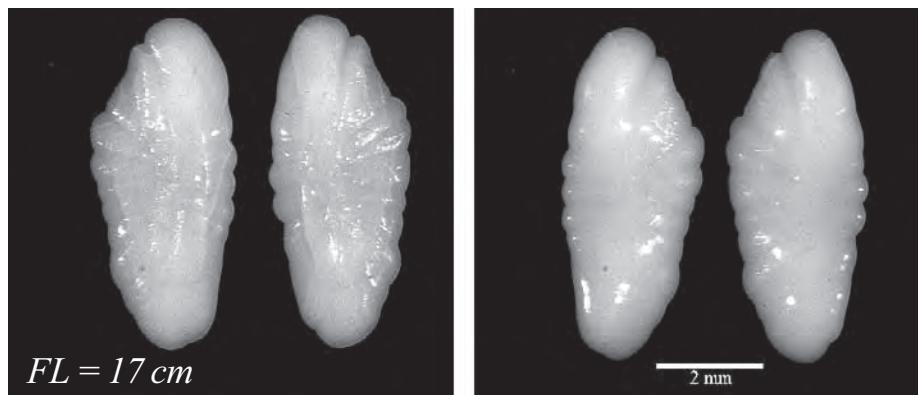
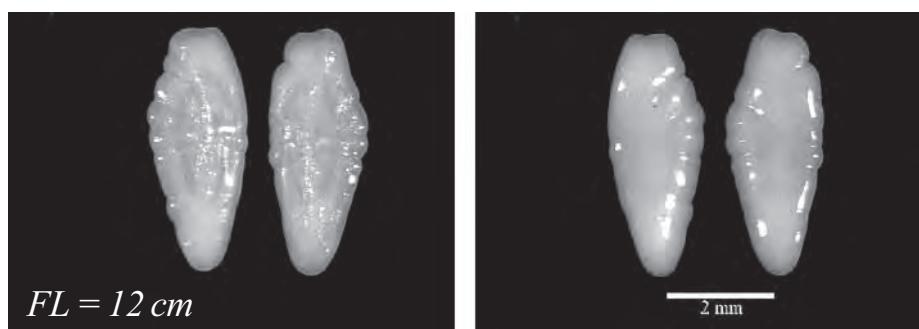
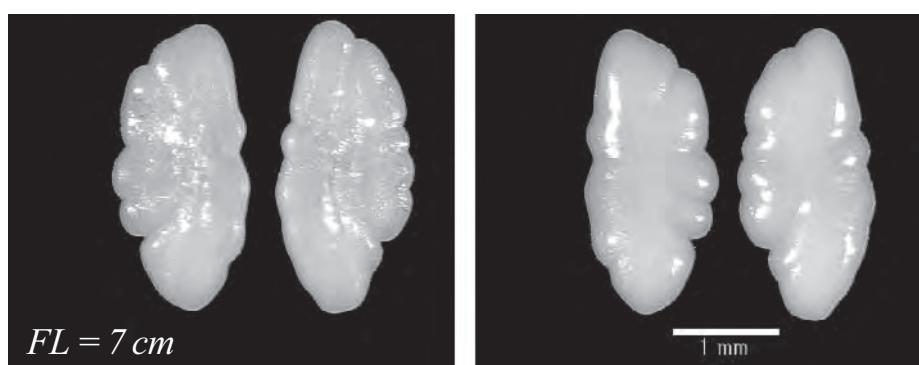
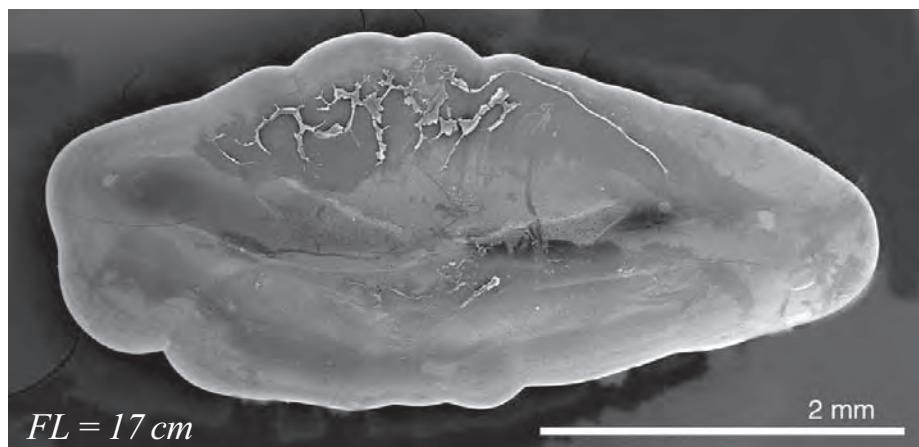
Fundulidae: *Fundulus heteroclitus*



Gadiformes

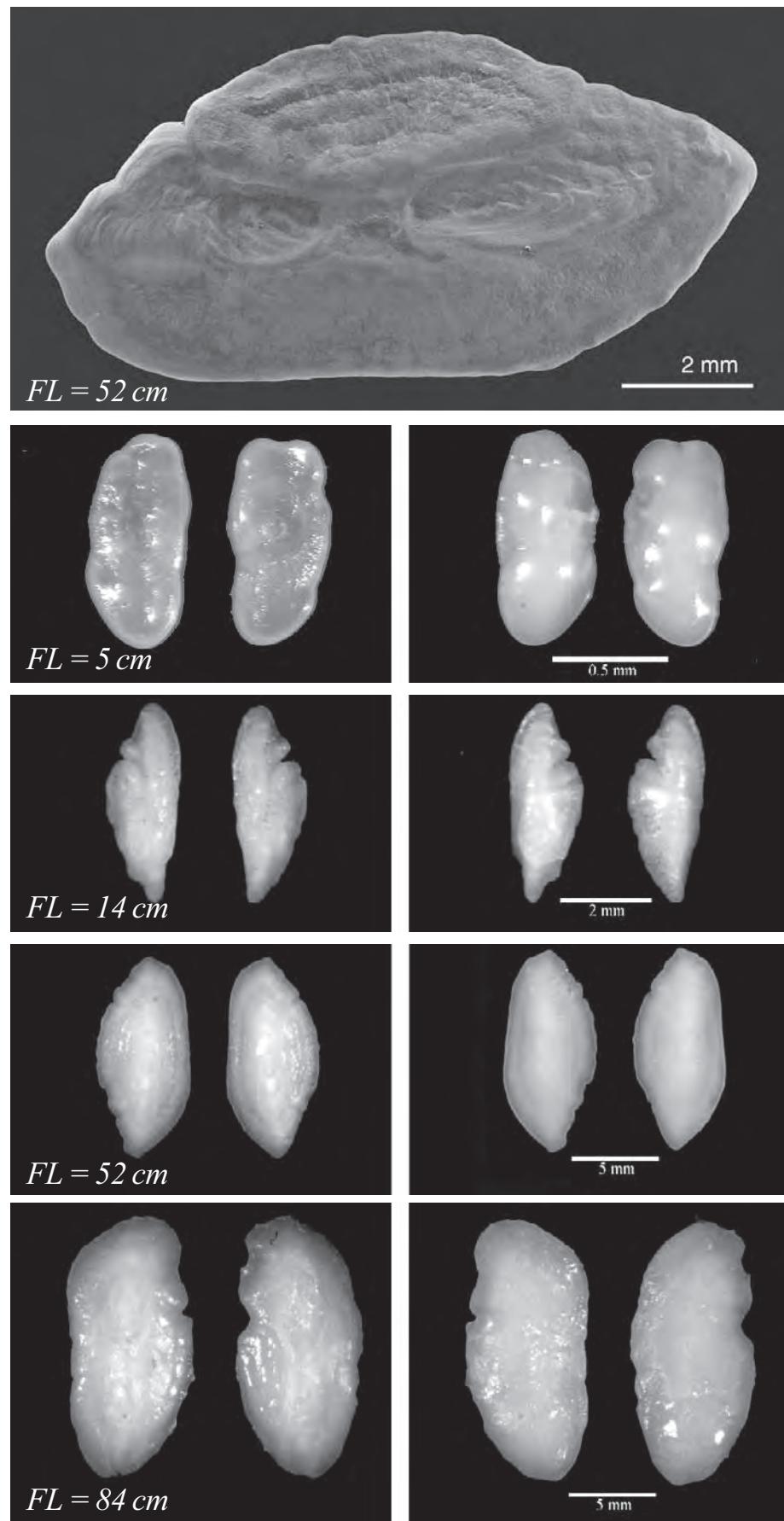
51

Gadidae: *Boreogadus saida*



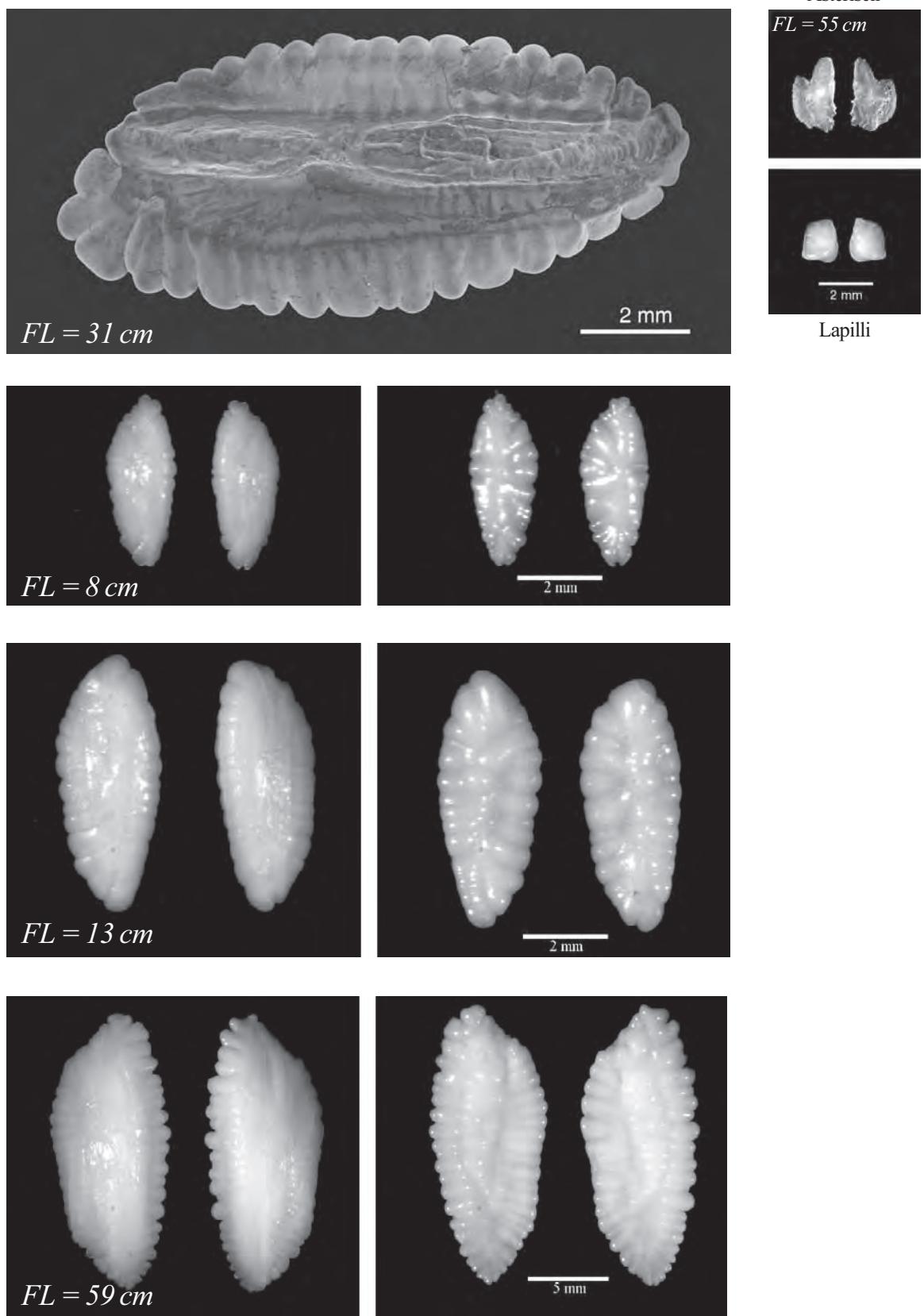
Gadiformes

Gadidae: *Brosme brosme*



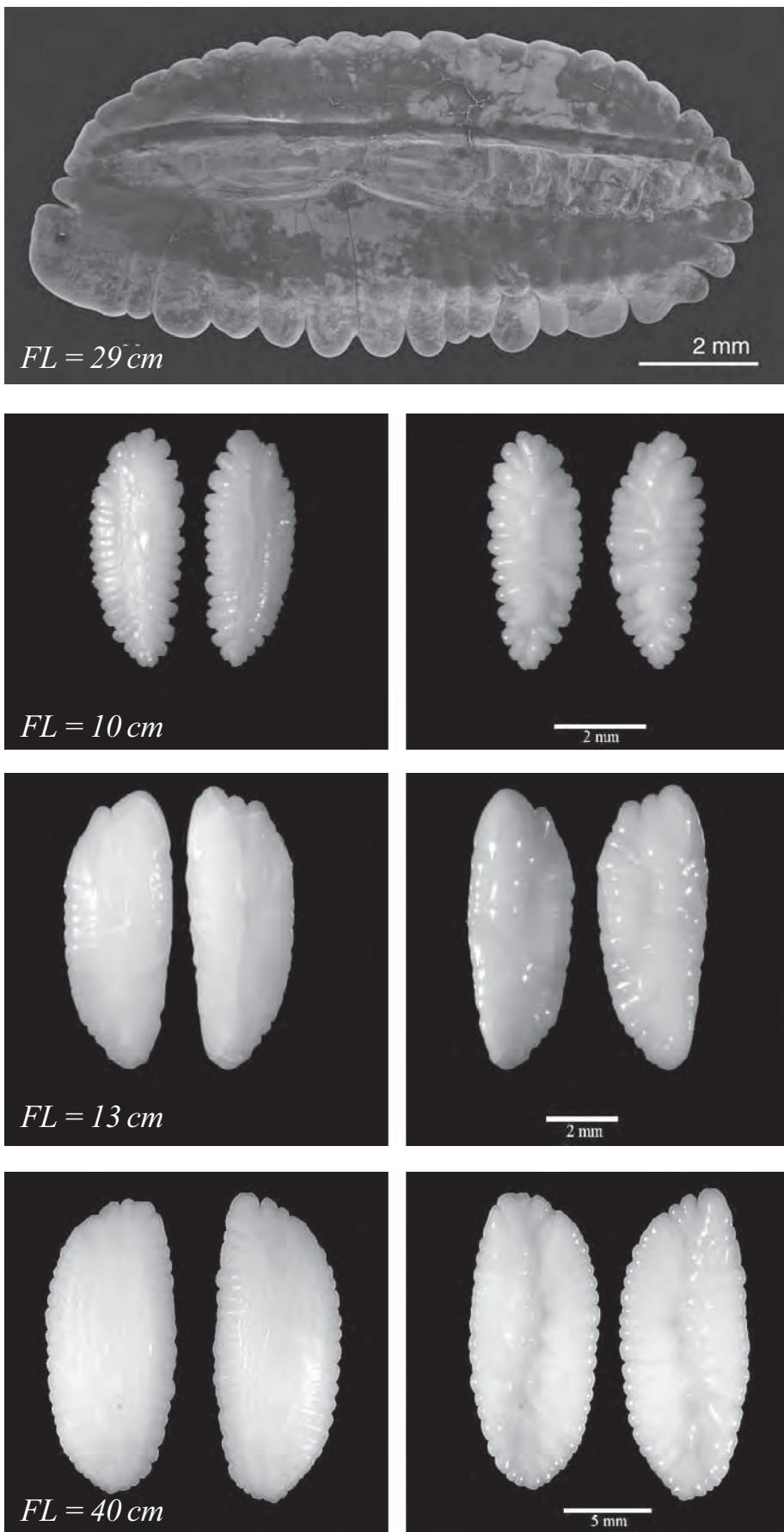
Gadiformes

Gadidae: *Gadus morhua*



Gadiformes

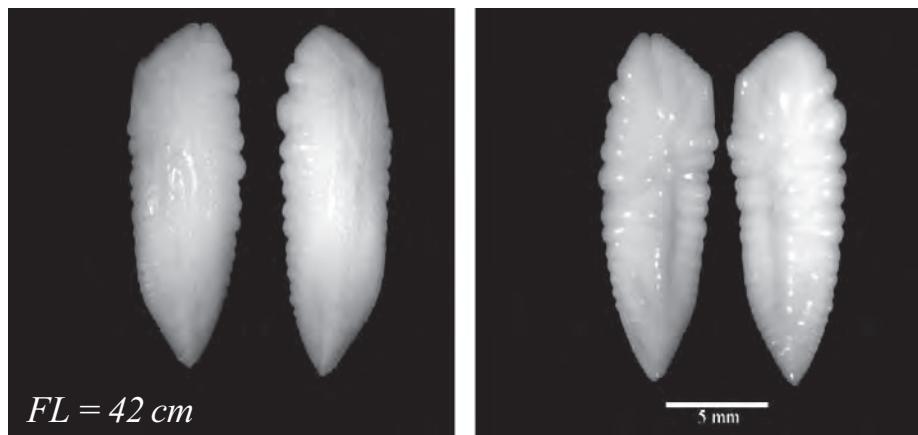
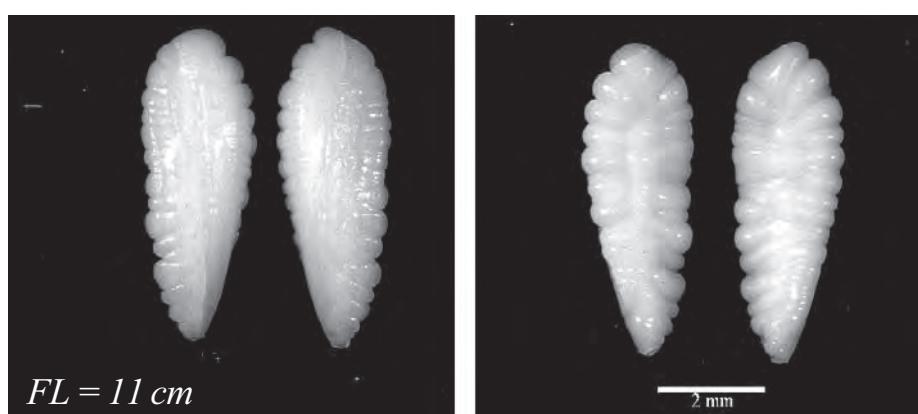
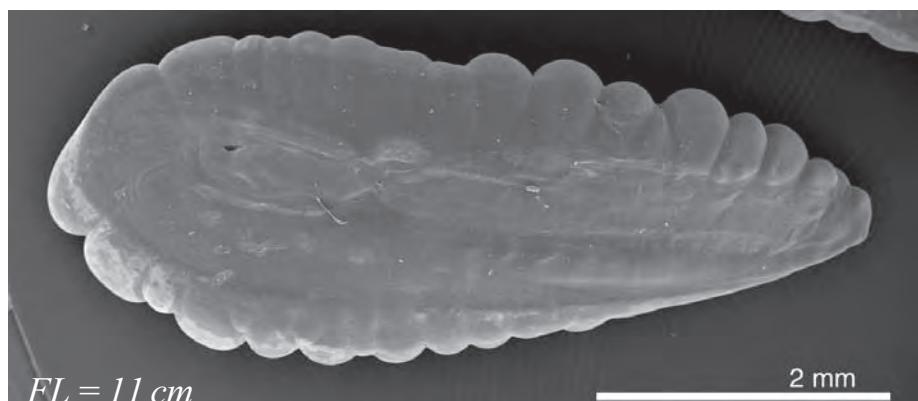
Gadidae: *Gadus ogac*



Gadiformes

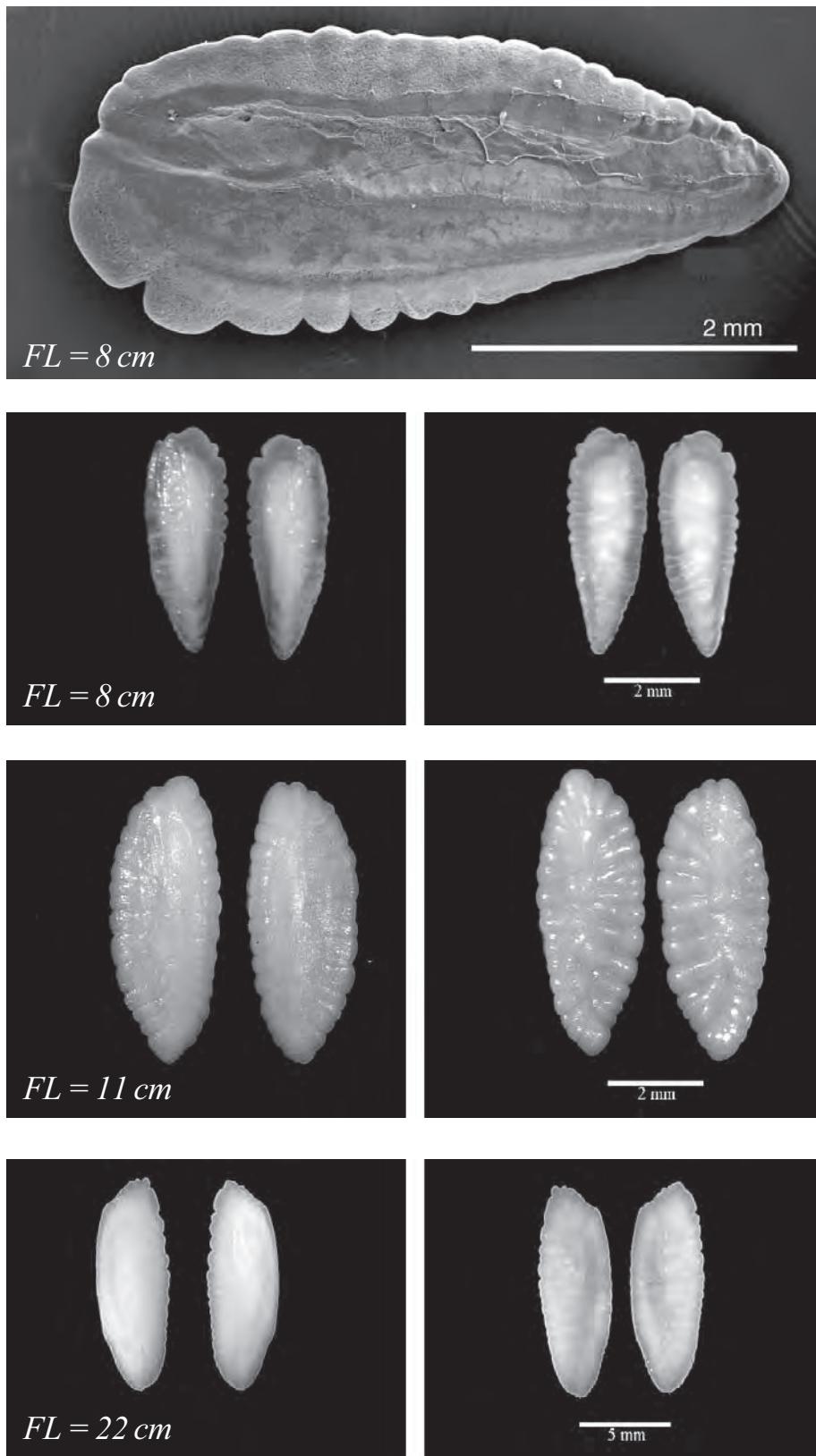
55

Gadidae: *Melanogrammus aeglefinus*



Gadiformes

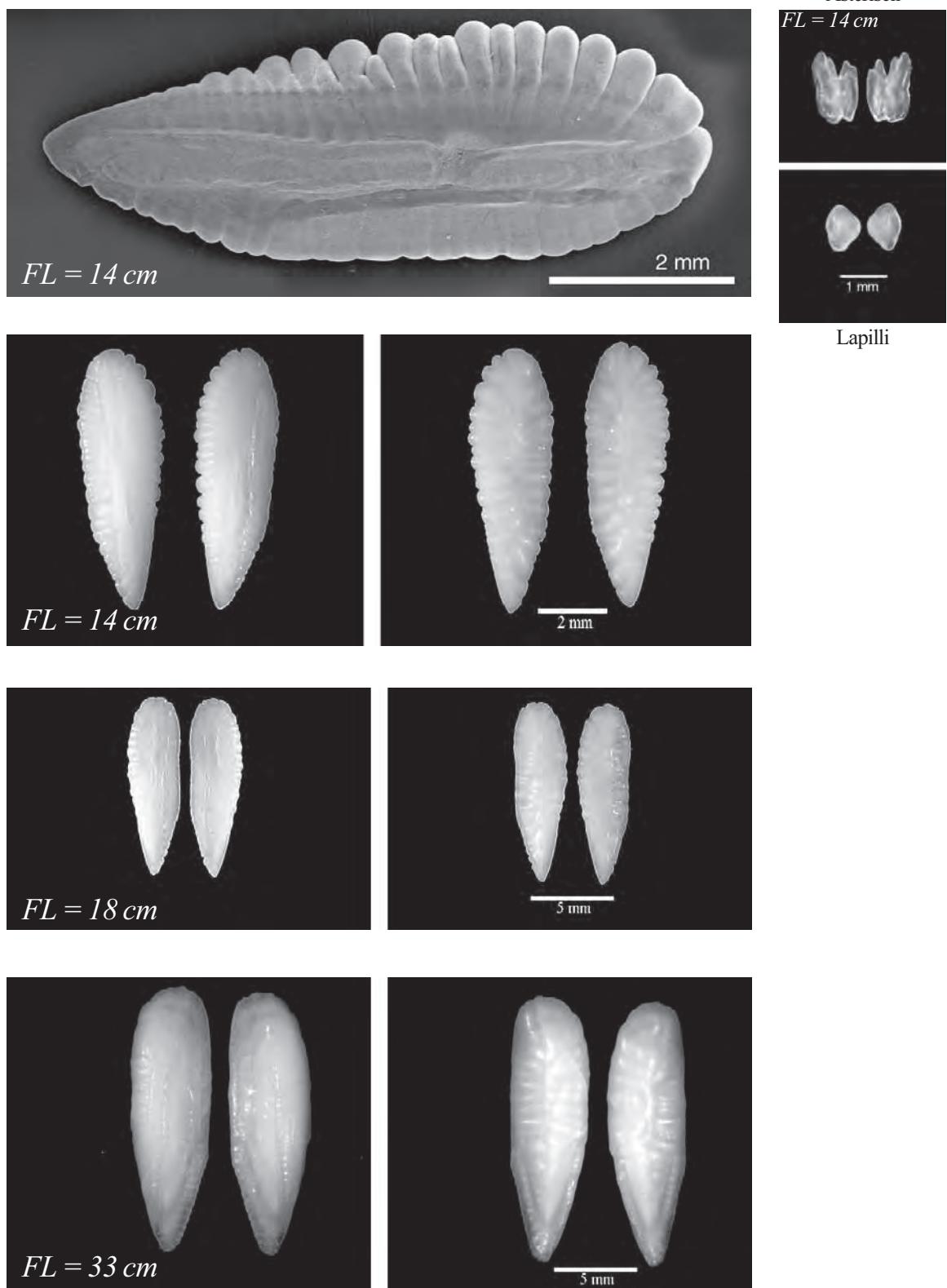
Gadidae: *Microgadus tomcod*



Gadiformes

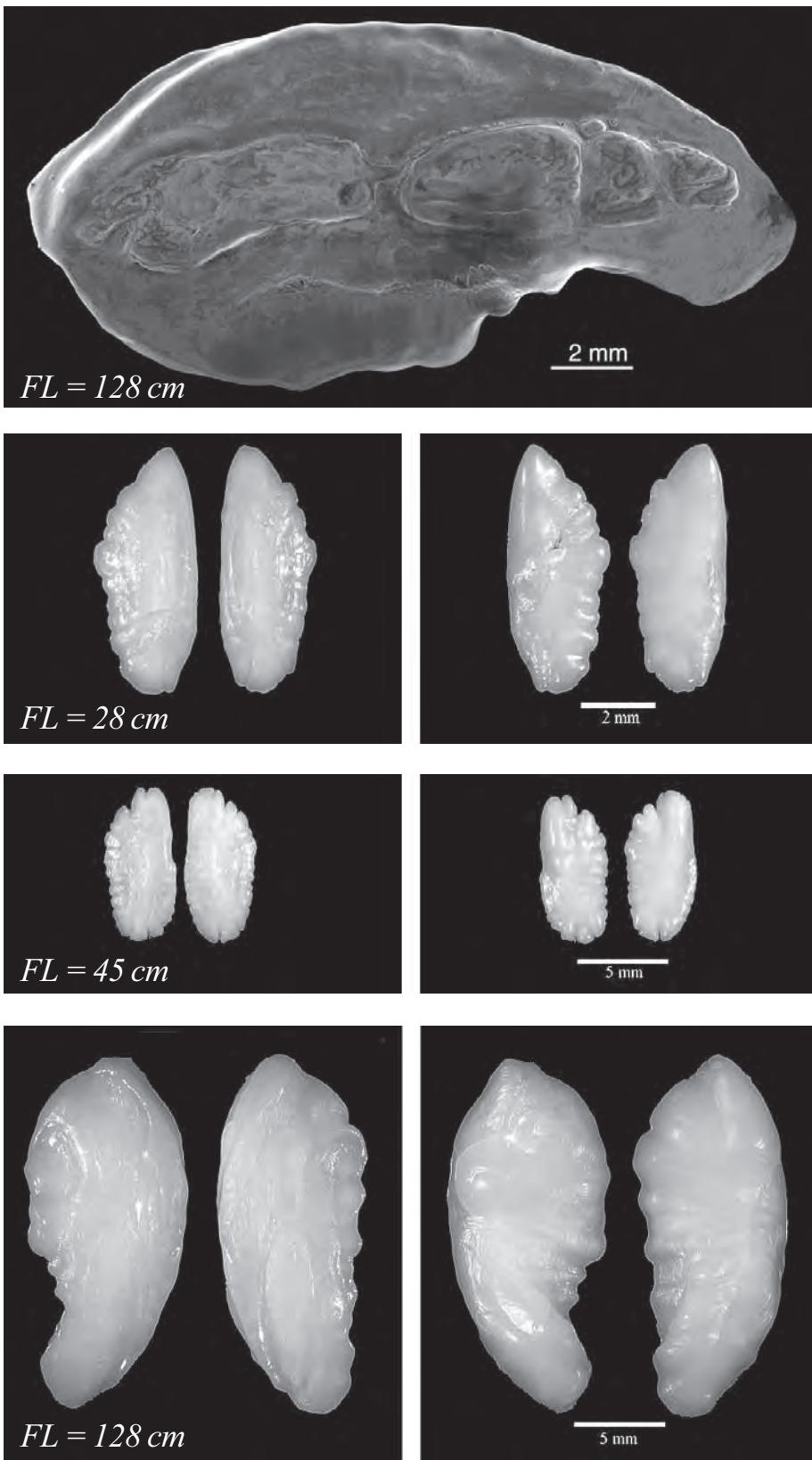
57

Gadidae: *Micromesistius poutassou*



Gadiformes

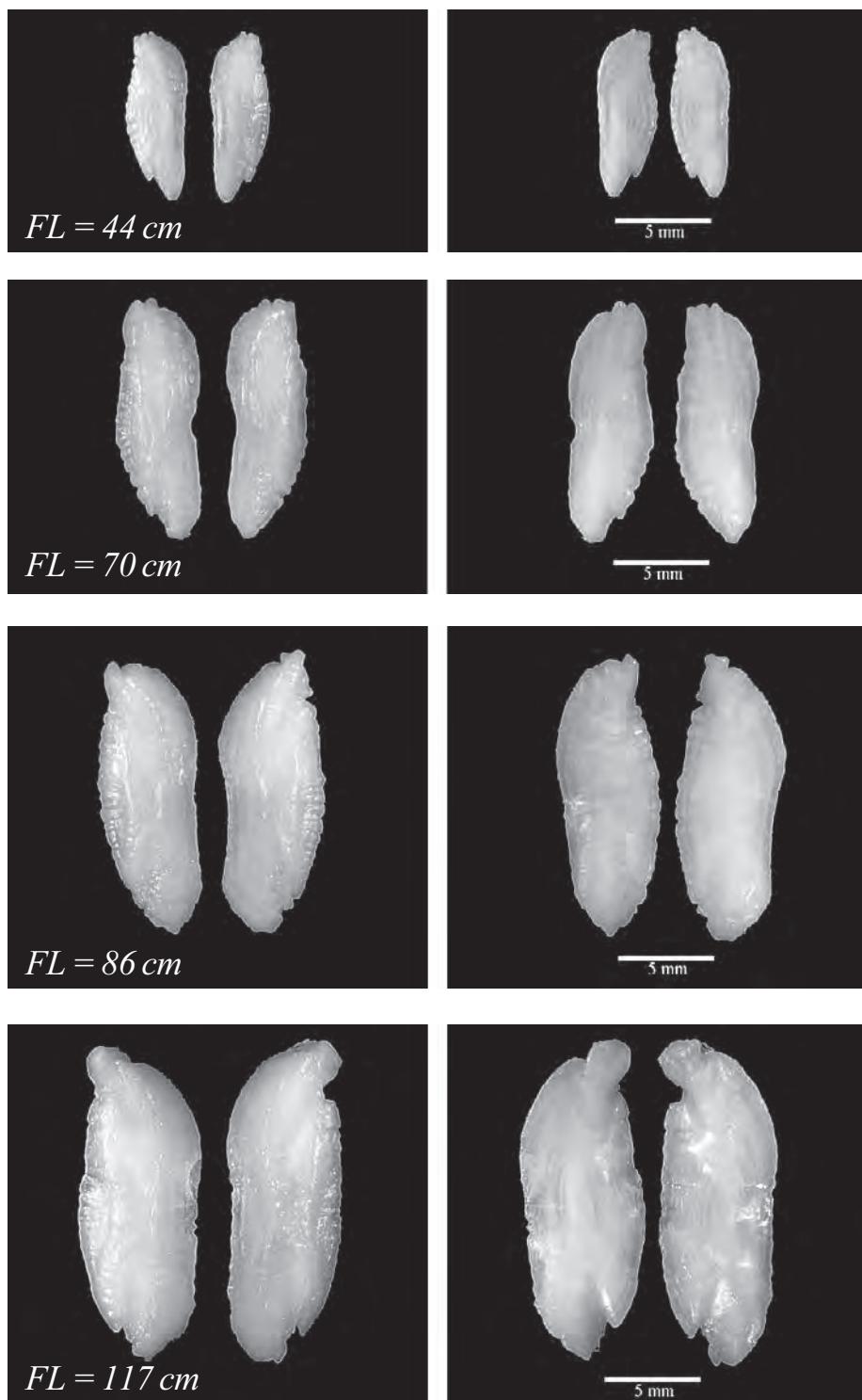
Gadidae: *Molva dypterygia*



Gadiformes

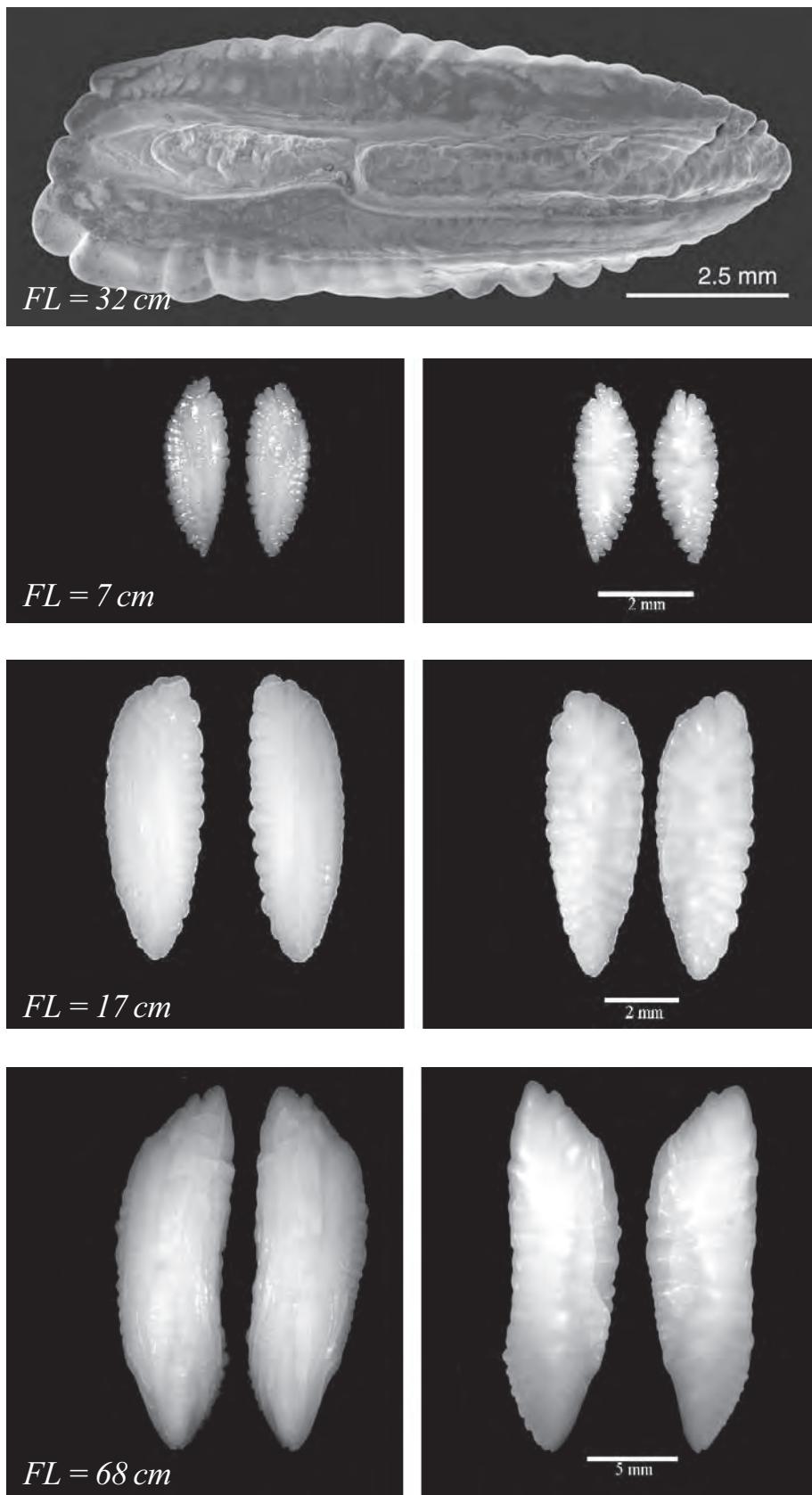
Gadidae: *Molva molva*

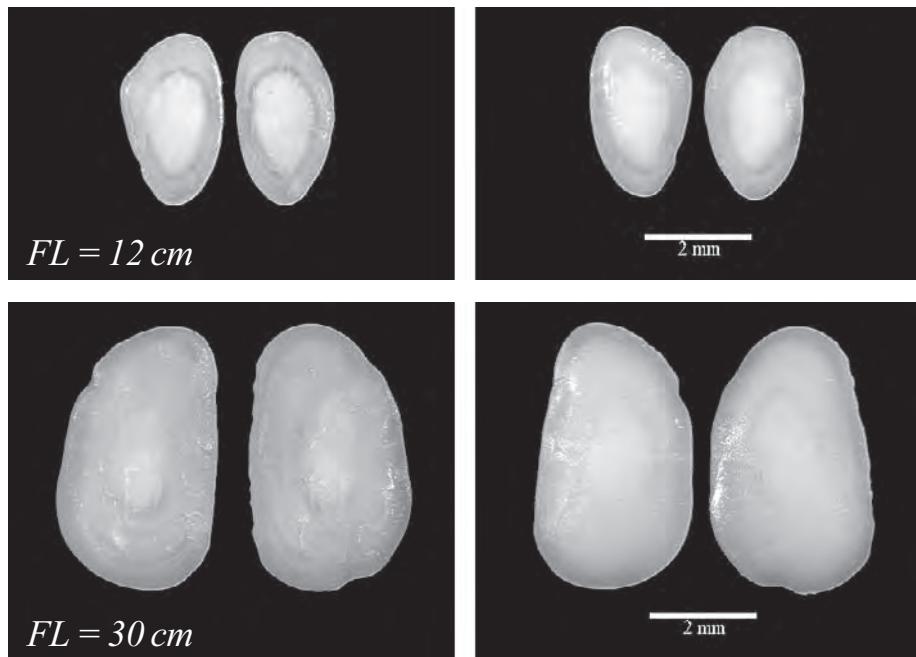
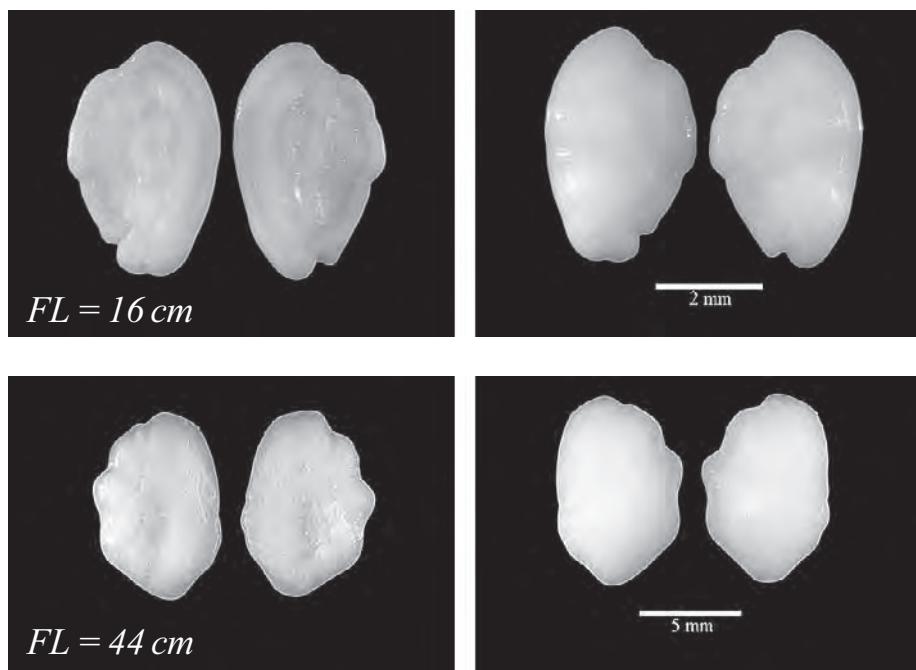
59



Gadiformes

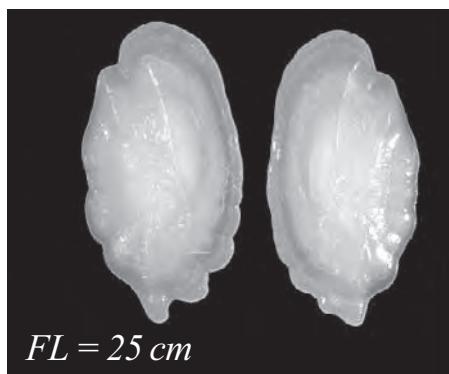
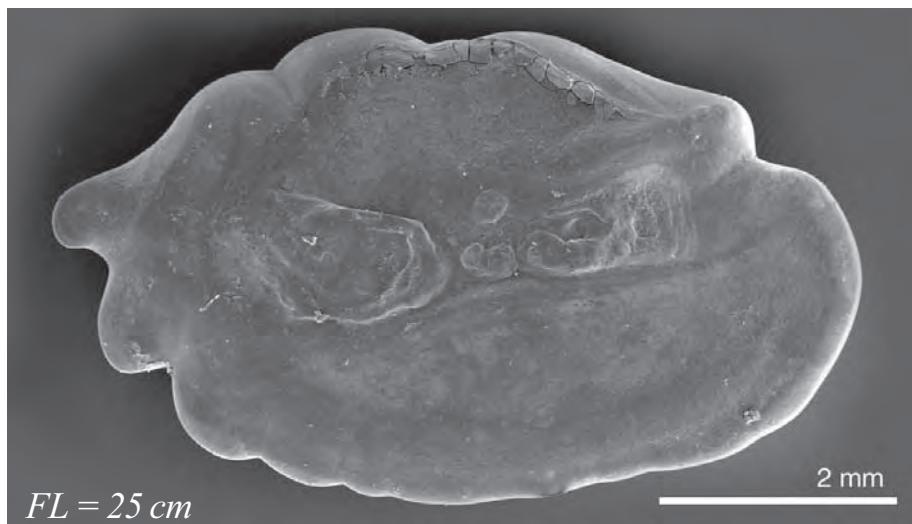
Gadidae: *Pollachius virens*



Macrouridae: *Chalinura brevibarbis*Macrouridae: *Coryphaenoides guentheri*

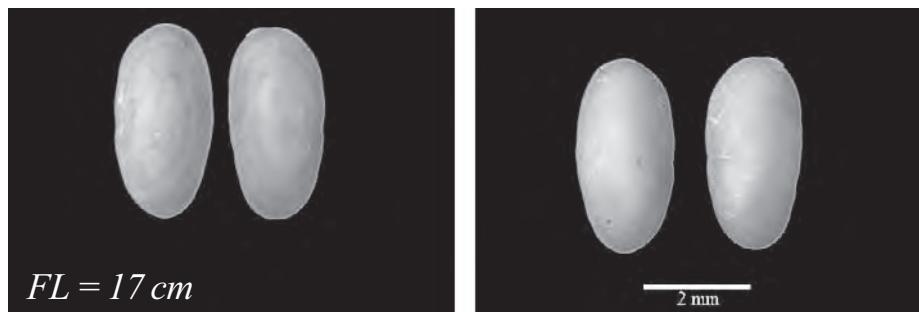
Gadiformes

Macrouridae: *Coryphaenoides rupestris*



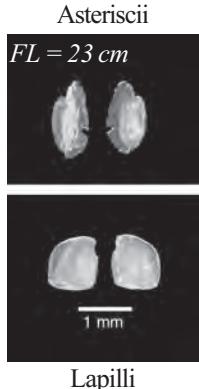
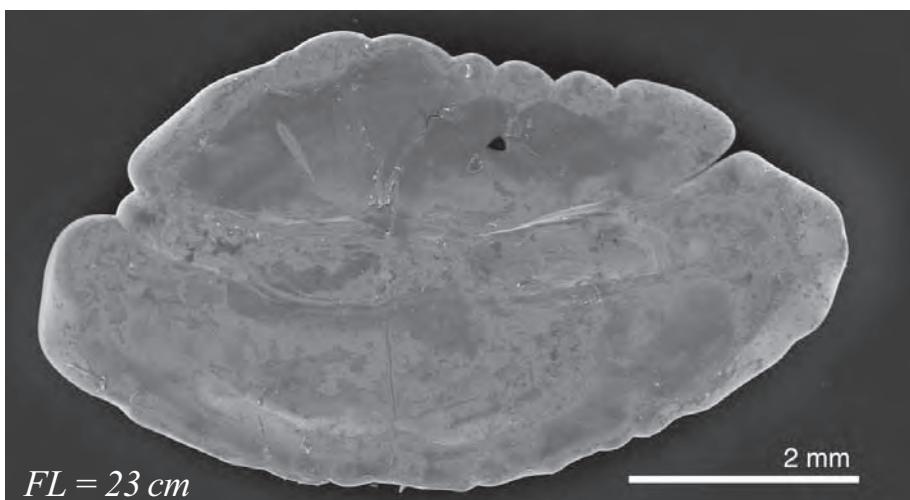
Gadiformes

Macrouridae: *Lionurus carapinus*



Gadiformes

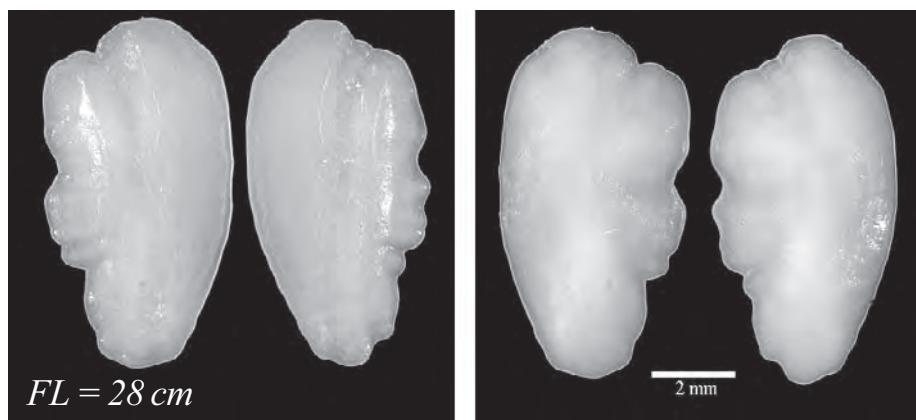
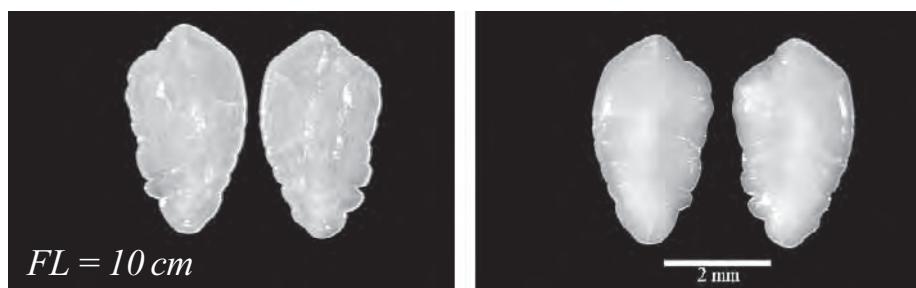
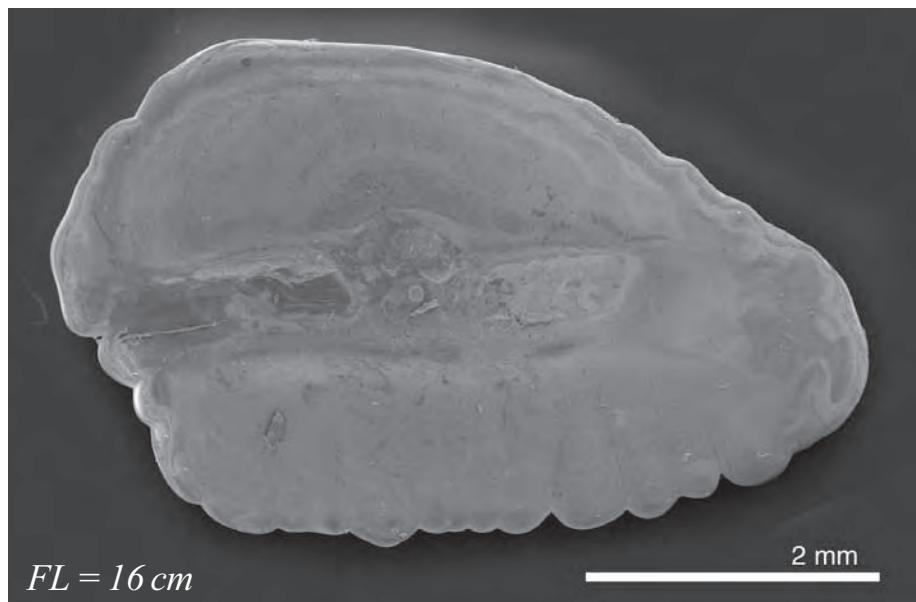
Macrouridae: *Macrourus berglax*



Gadiformes

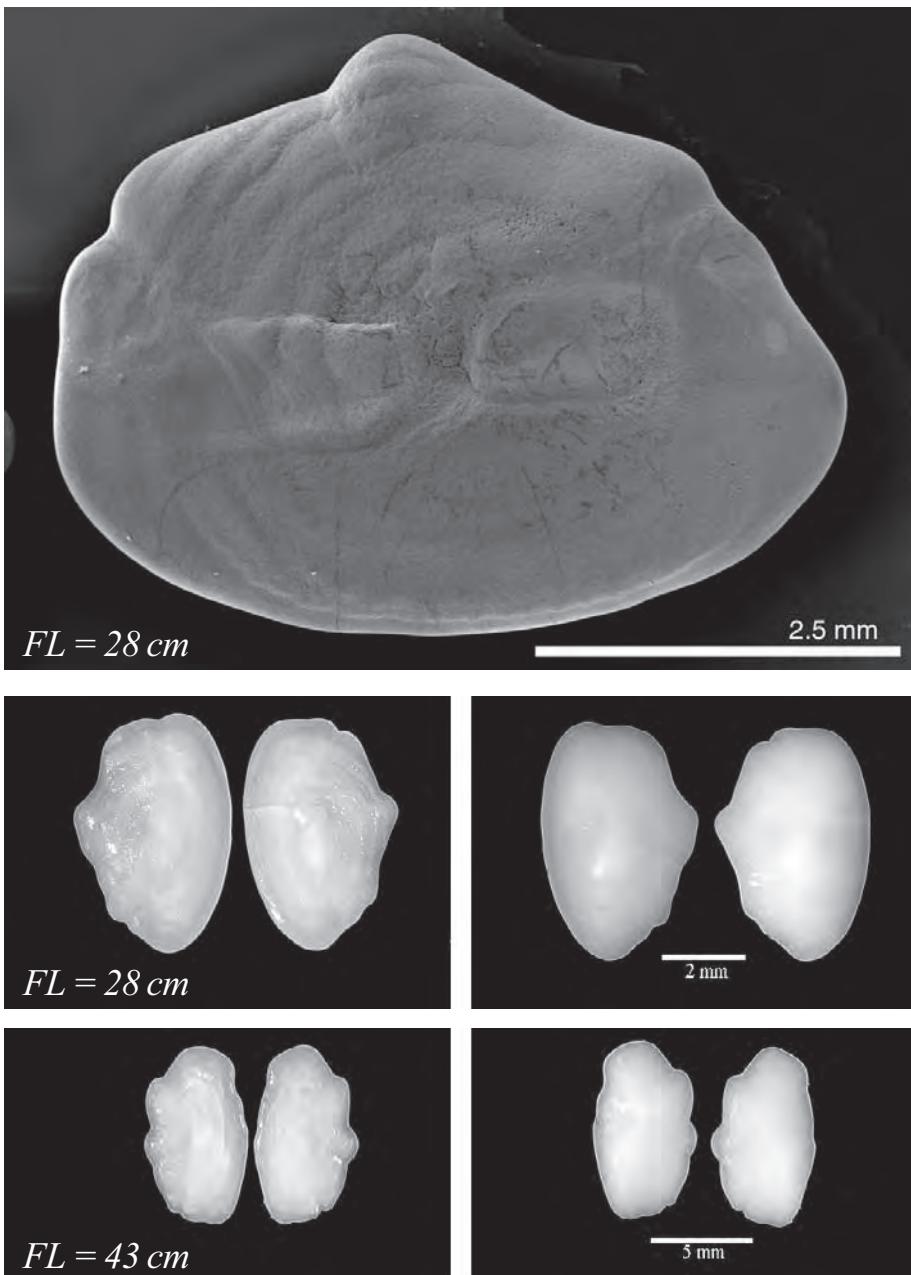
65

Macrouridae: *Nezumia bairdi*

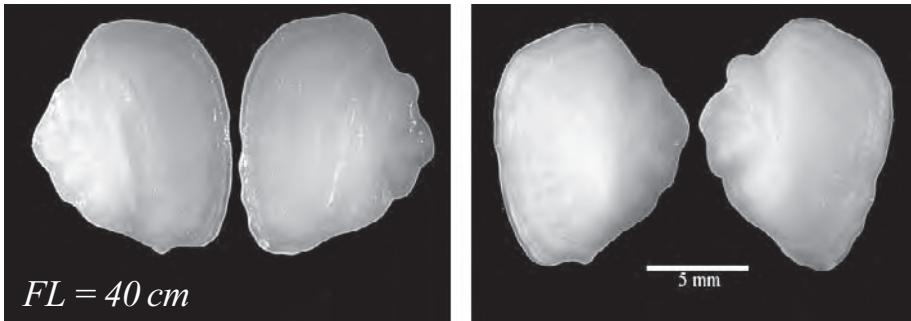


Gadiformes

Macrouridae: *Nezumia sclerorhynchus*



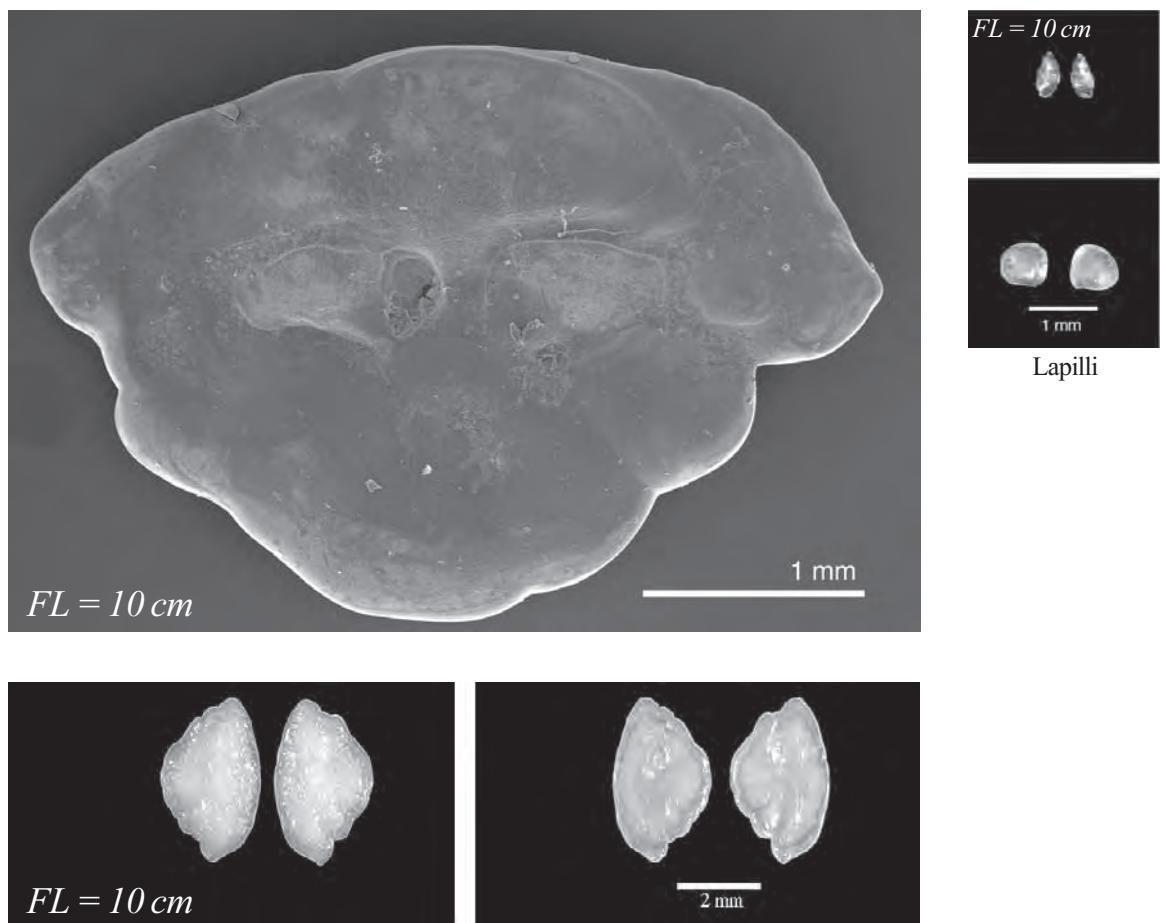
Macrouridae: *Trachyrhynchus murrayi*



Gadiformes

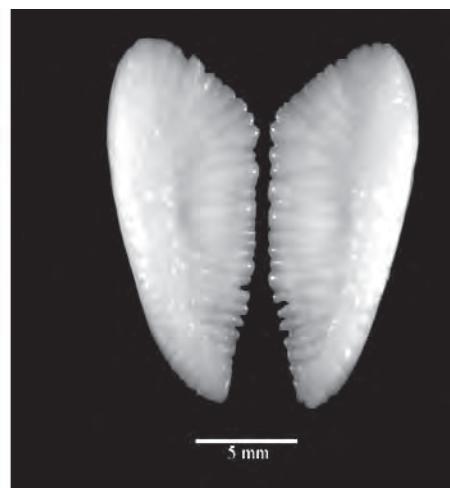
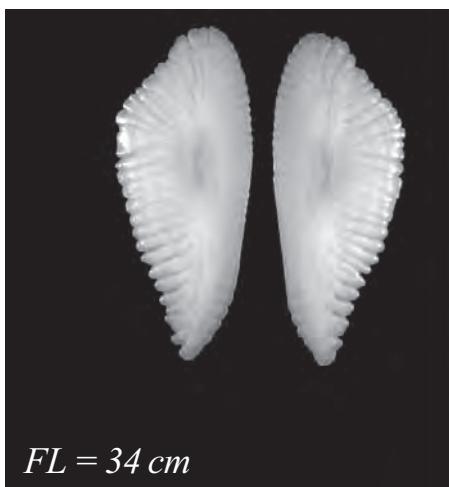
67

Melanonidae: *Melanonus zugmayeri*



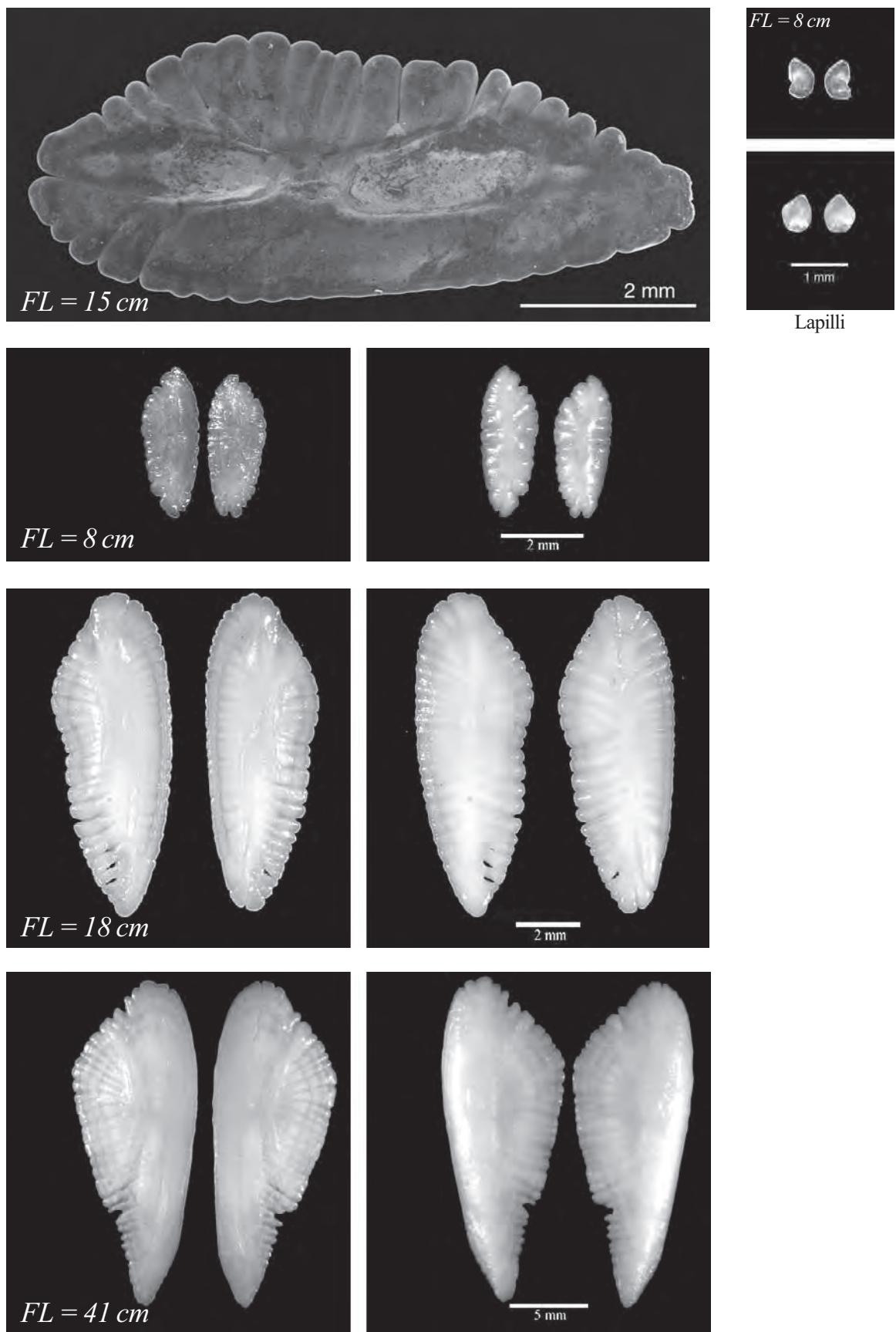
Gadiformes

Merlucciidae: *Merluccius albidus*



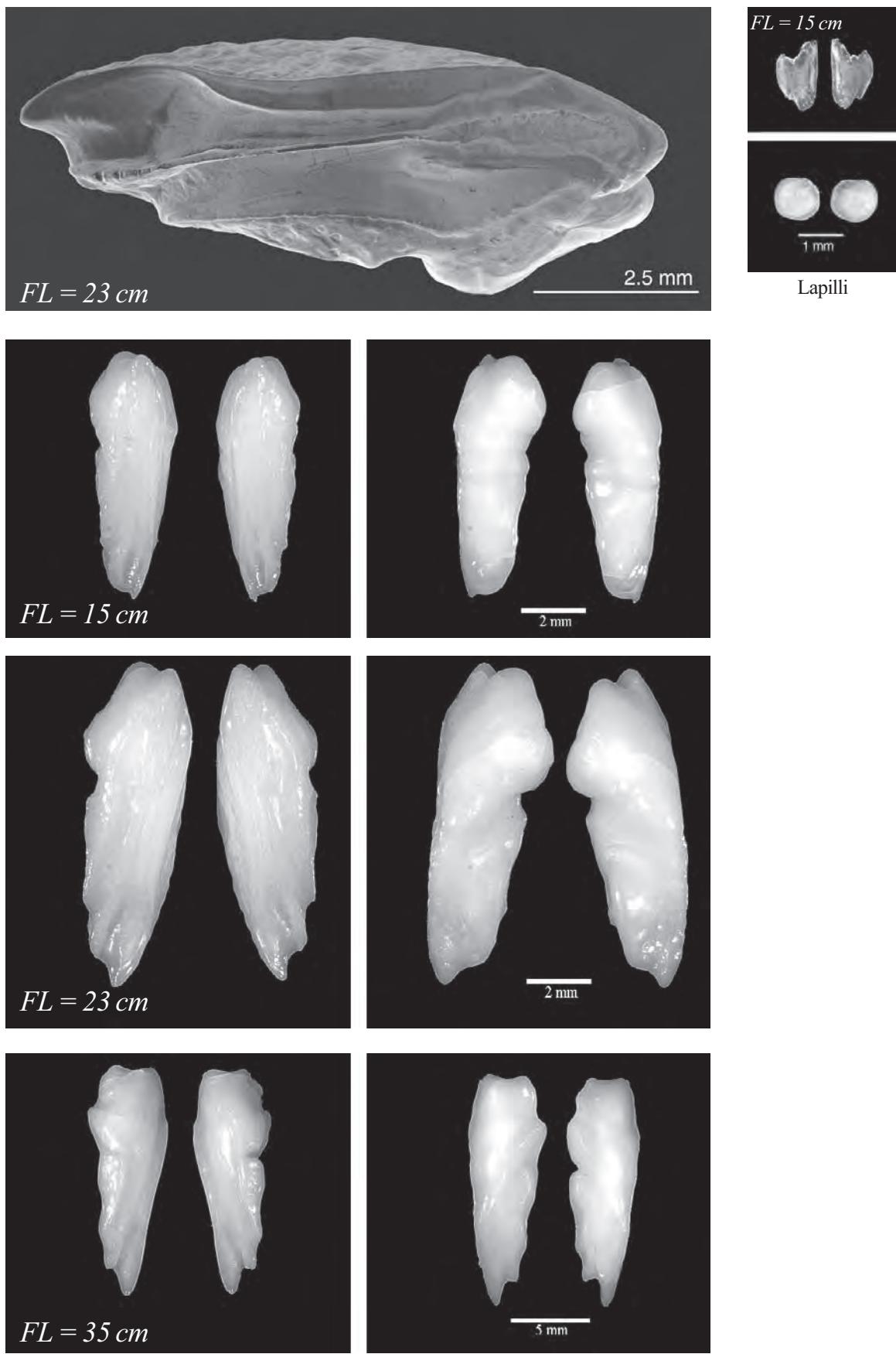
Gadiformes

Merlucciidae: *Merluccius bilinearis*



Gadiformes

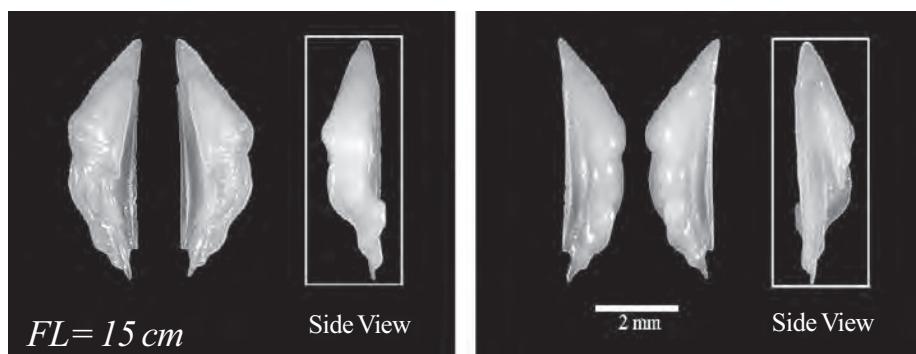
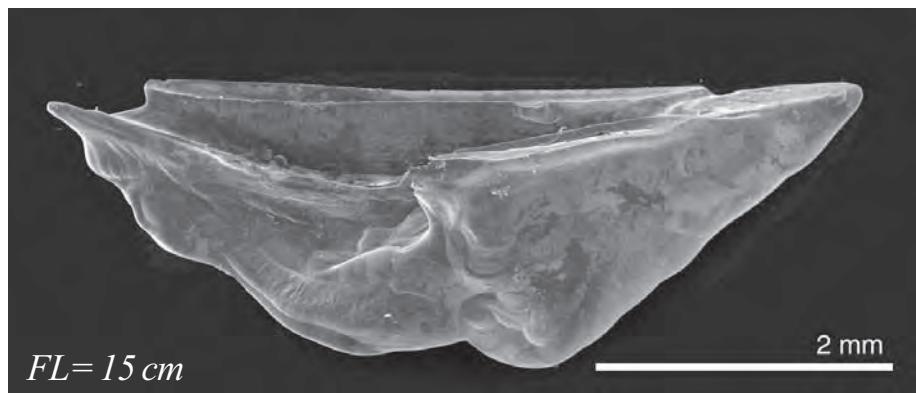
Moridae: *Antimora rostrata*



Gadiformes

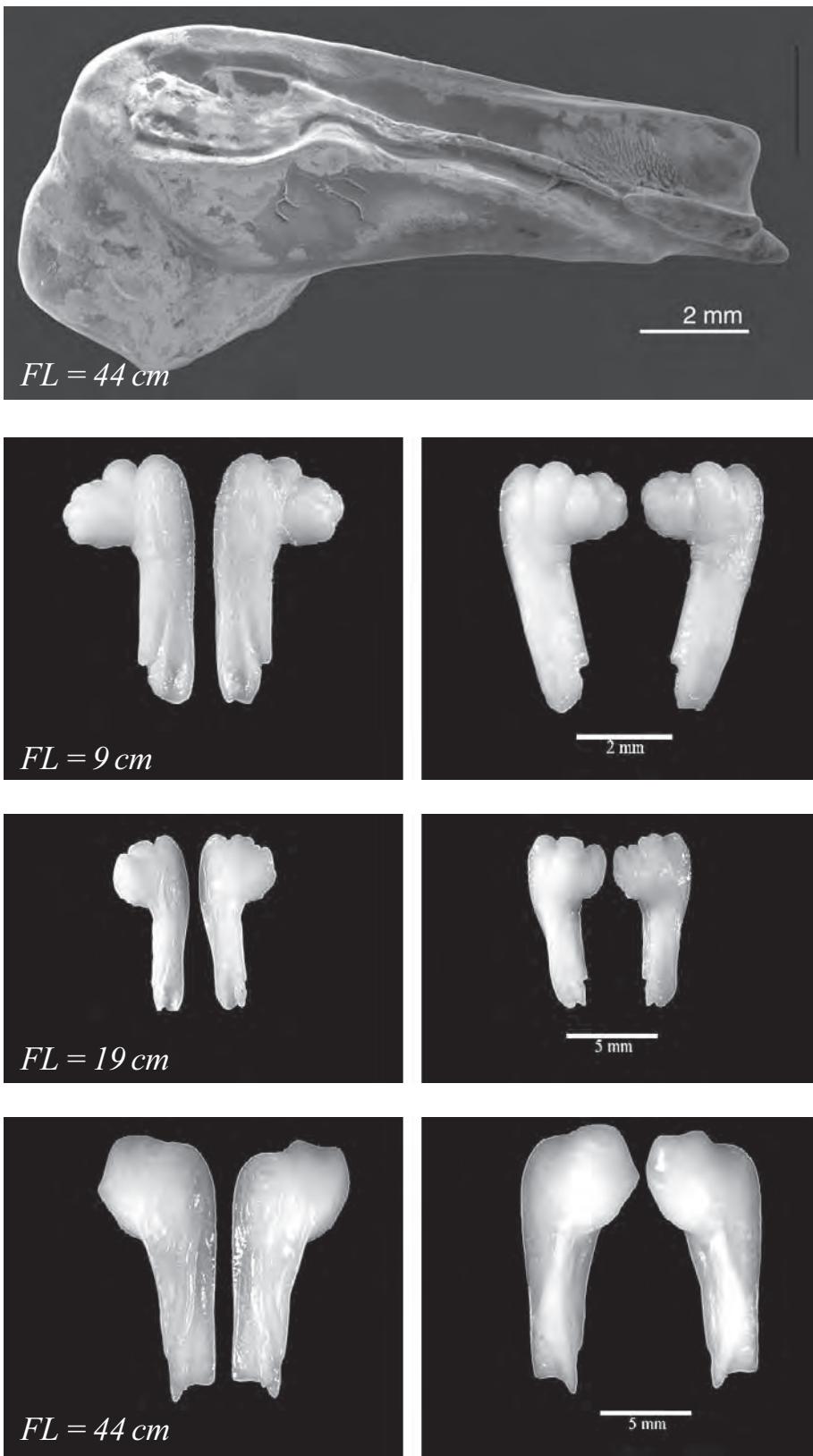
71

Moridae: *Brosmiculus imberbis*



Gadiformes

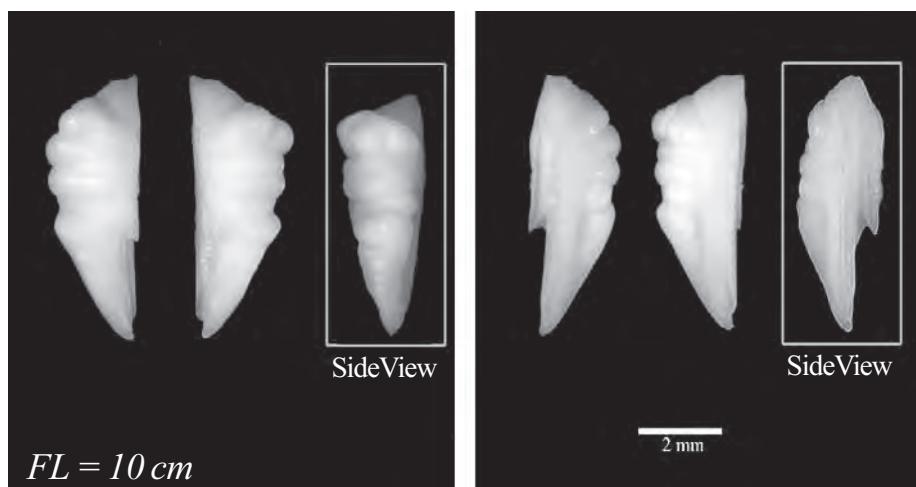
Moridae: *Halargyreus johnsoni*



Gadiformes

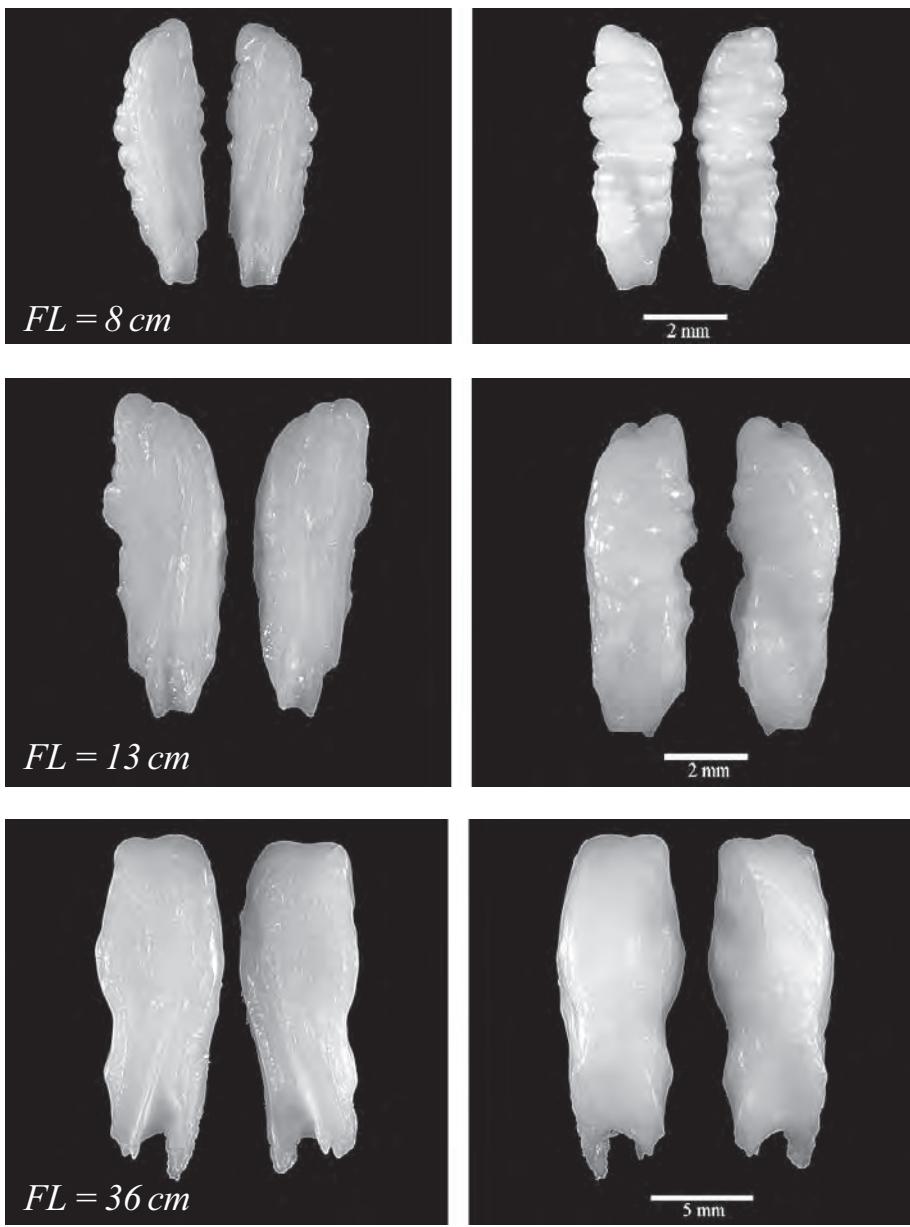
73

Moridae: *Laemonema barbatula*



Gadiformes

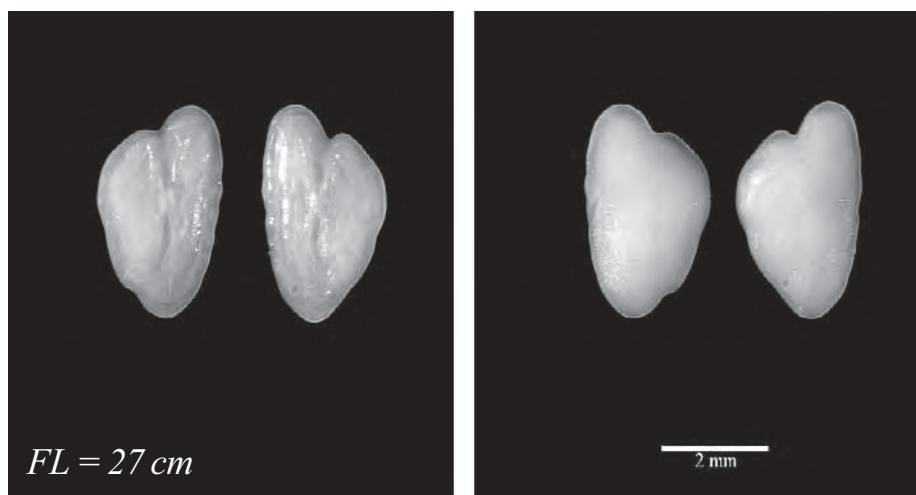
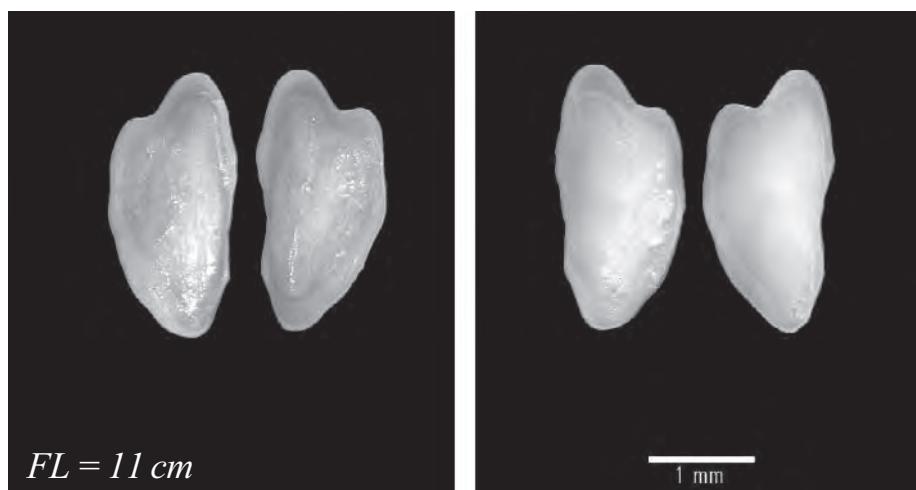
Moridae: *Lepidion eques*



Gadiformes

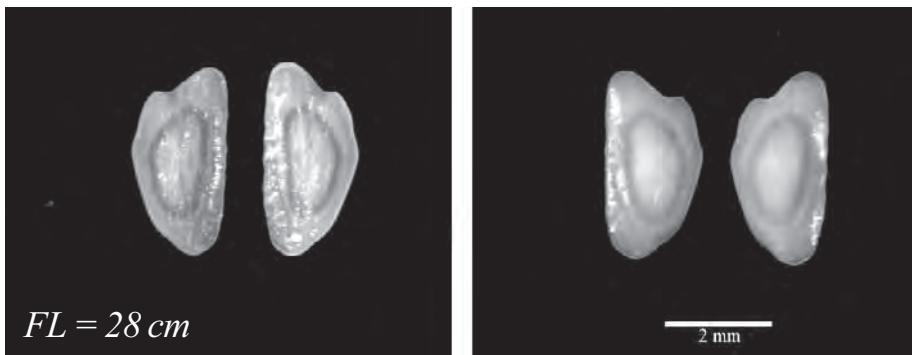
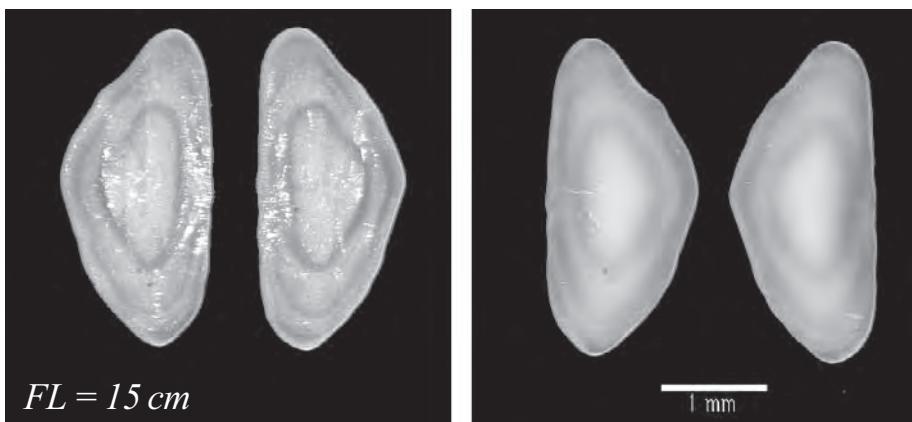
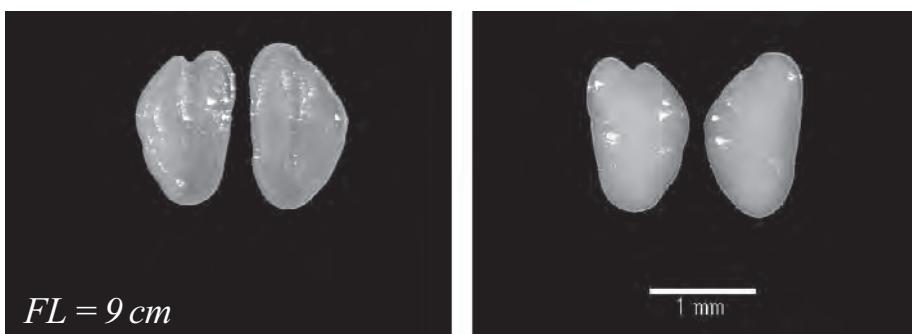
75

Phycidae: *Enchelyopus cimbrius*



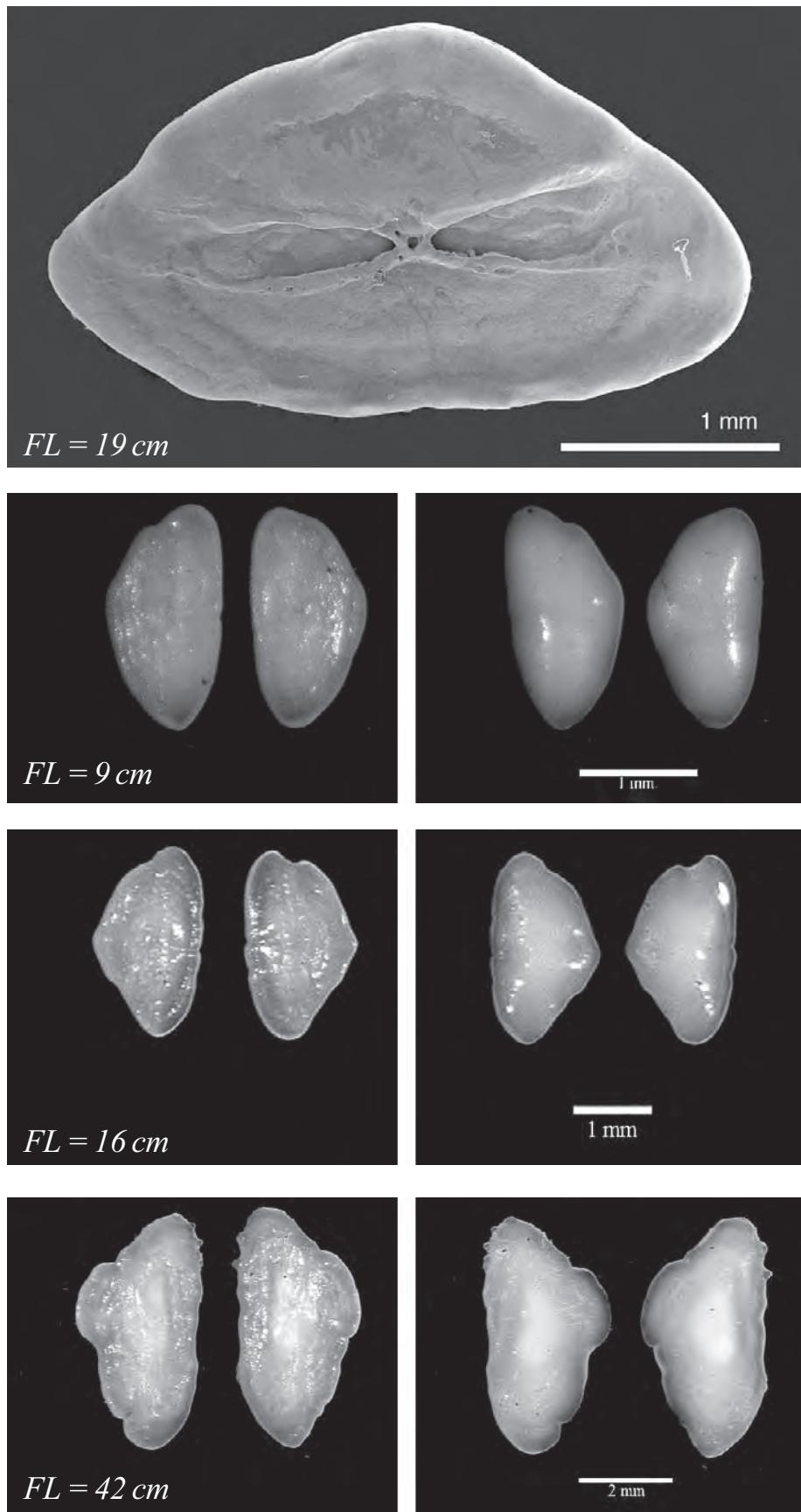
Gadiformes

Phycidae: *Gaidropsarus argentatus*



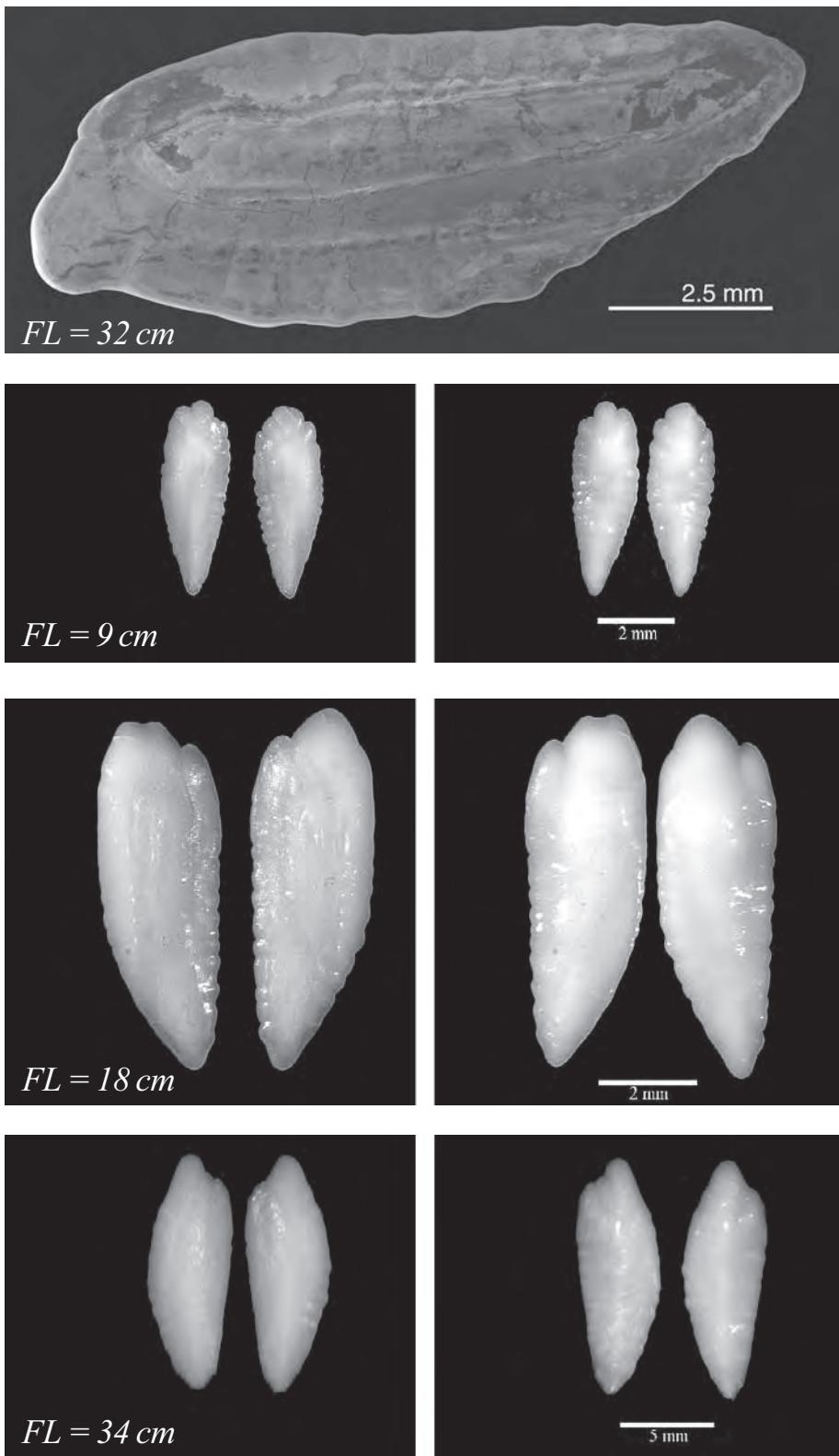
Gadiformes

Phycidae: *Gaidropsarus ensis*



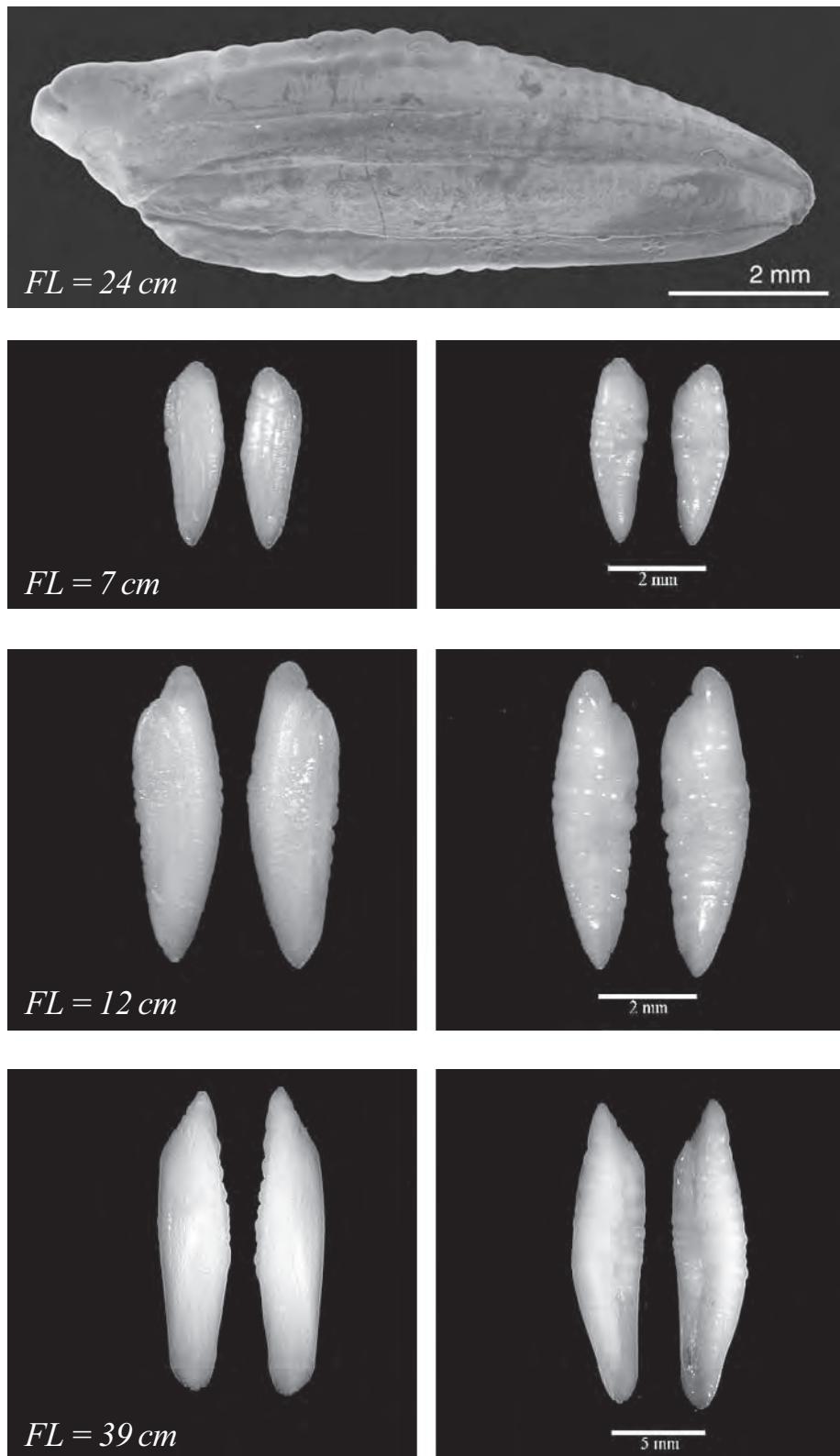
Gadiformes

Phycidae: *Urophycis chesteri*



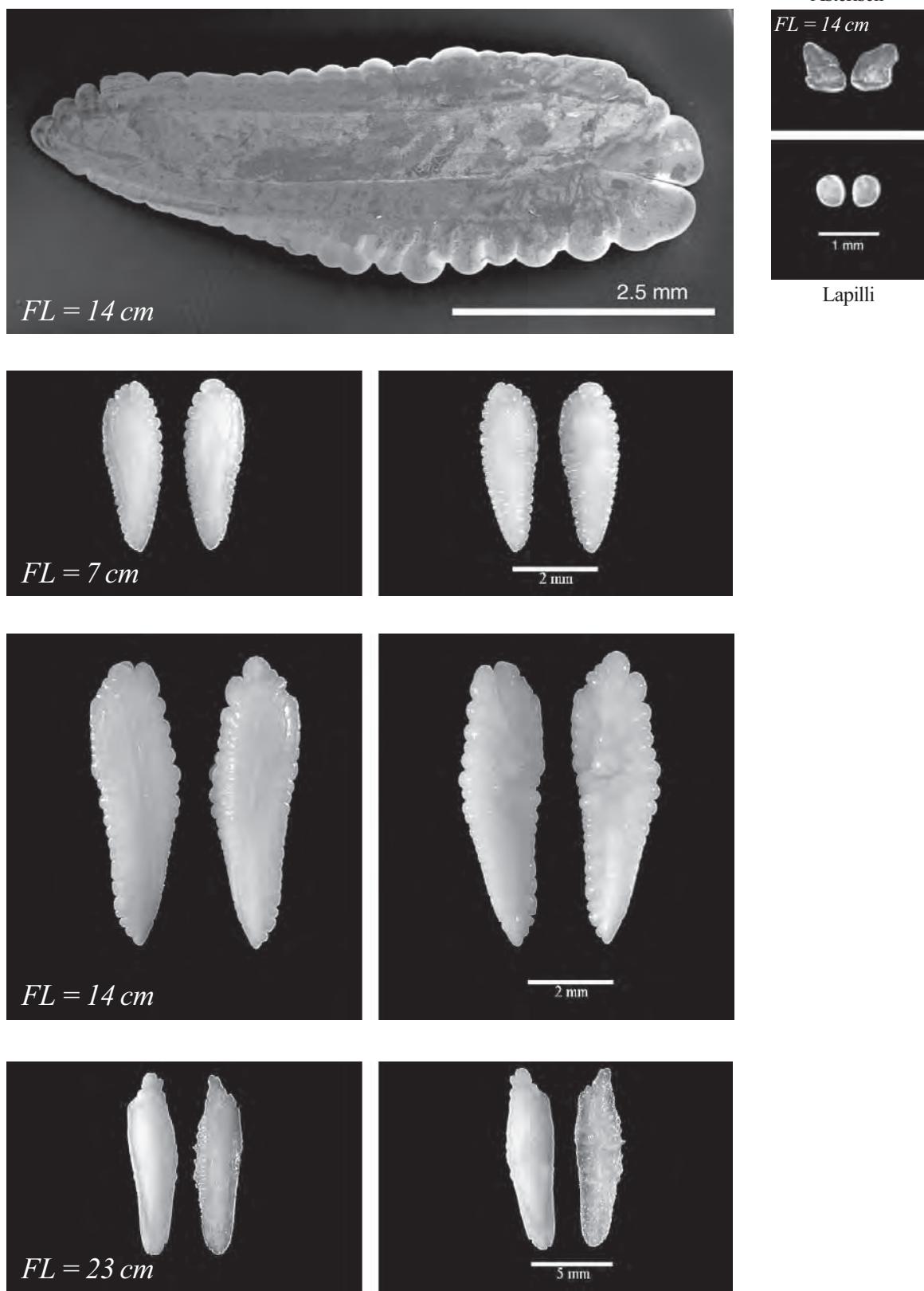
Gadiformes

Phycidae: *Urophycis chuss*



Gadiformes

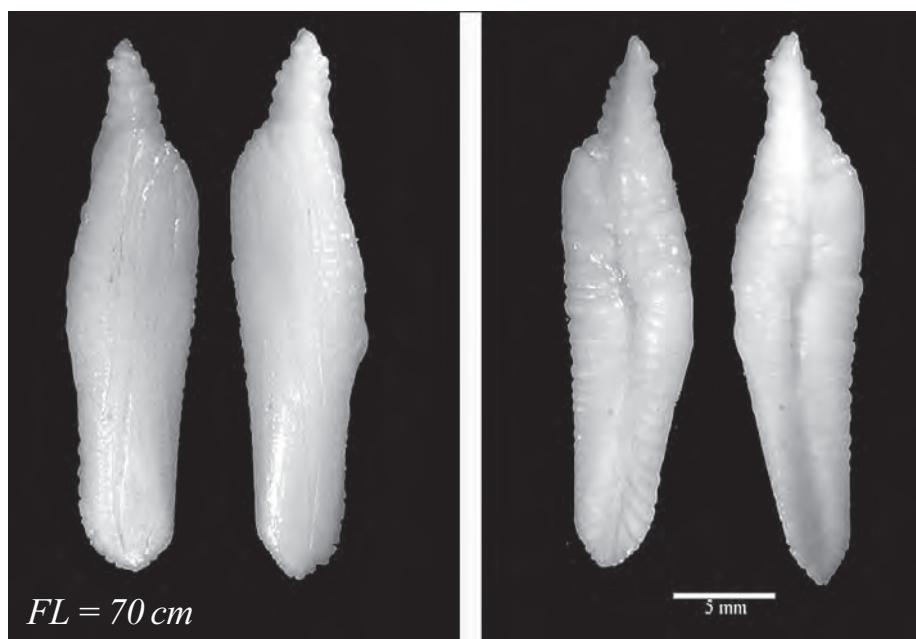
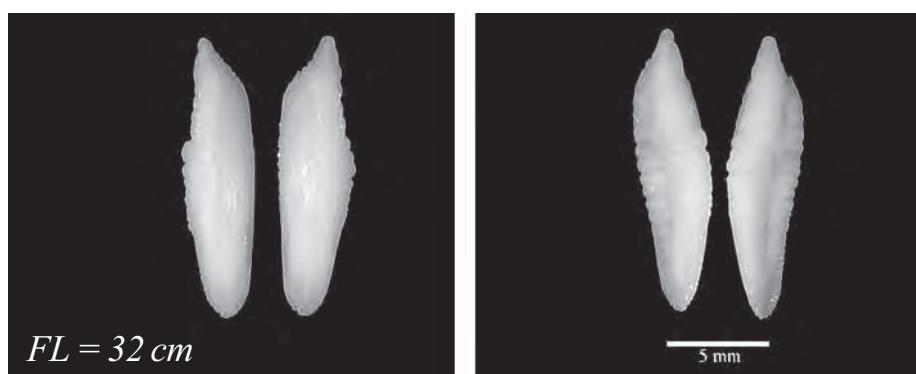
Phycidae: *Urophycis regia*



Gadiformes

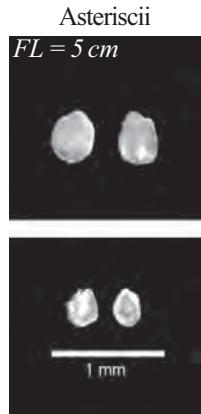
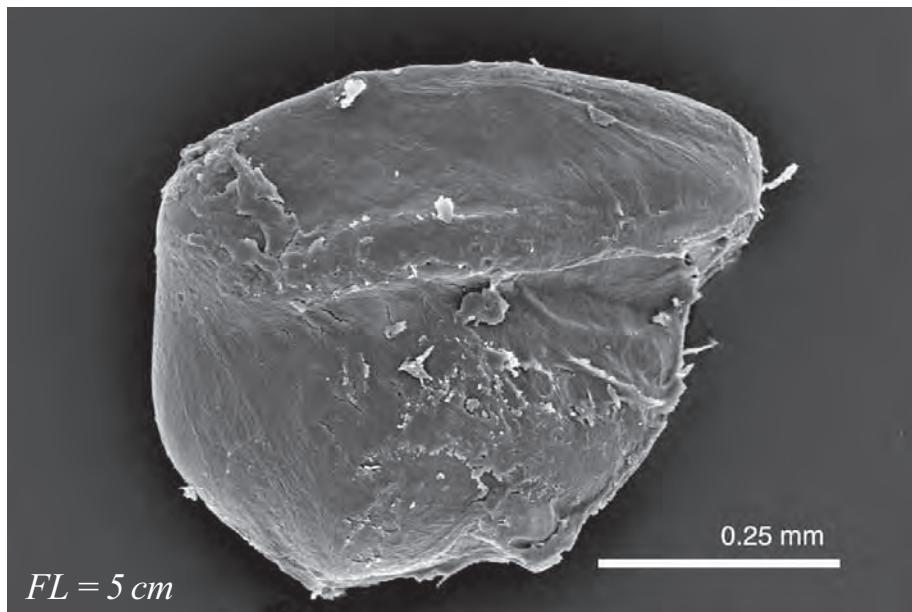
81

Phycidae: *Urophycis tenuis*



Gasterosteiformes

Gasterosteidae: *Apeltes quadratus*



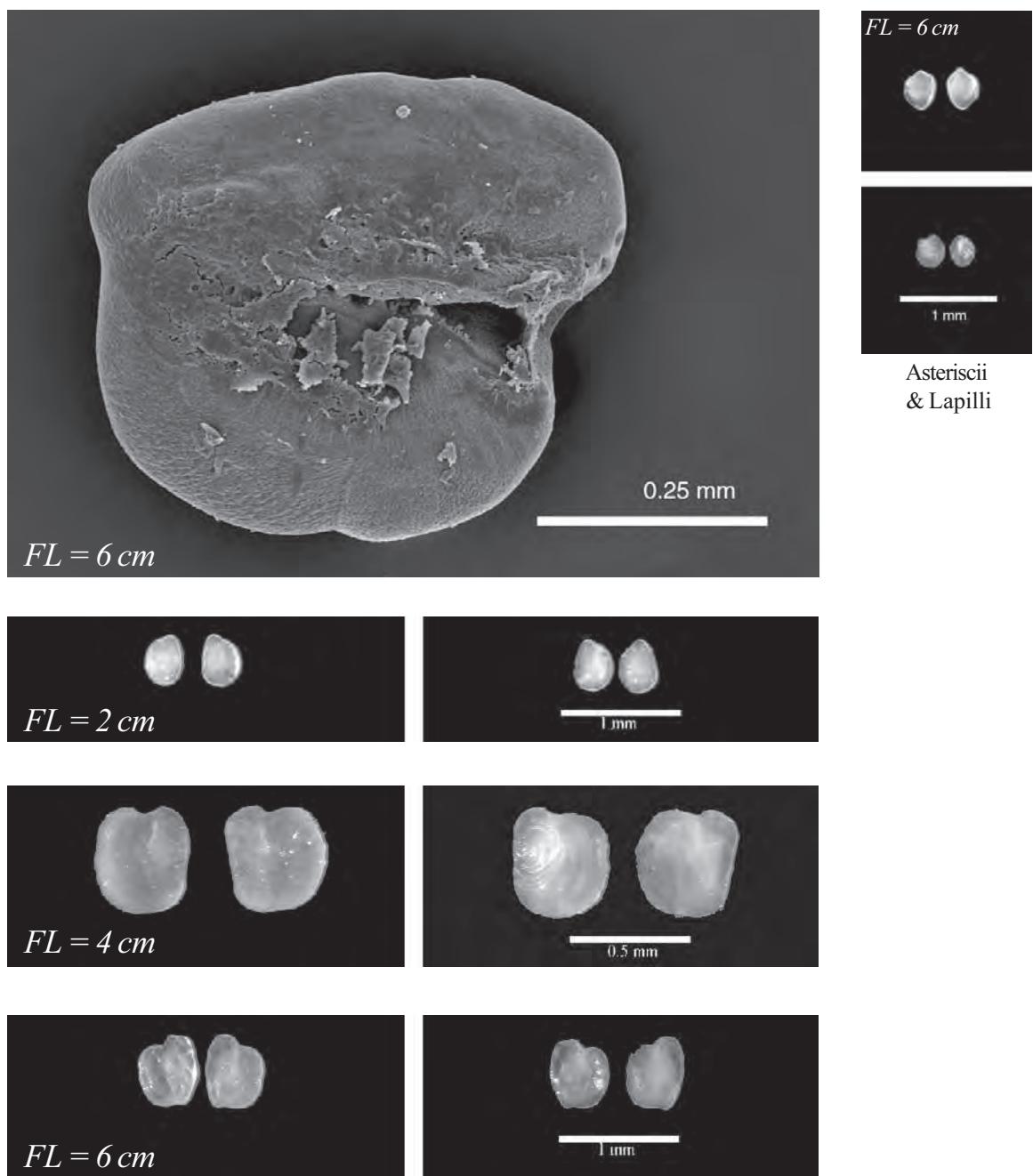
Lapilli



Gasterosteiformes

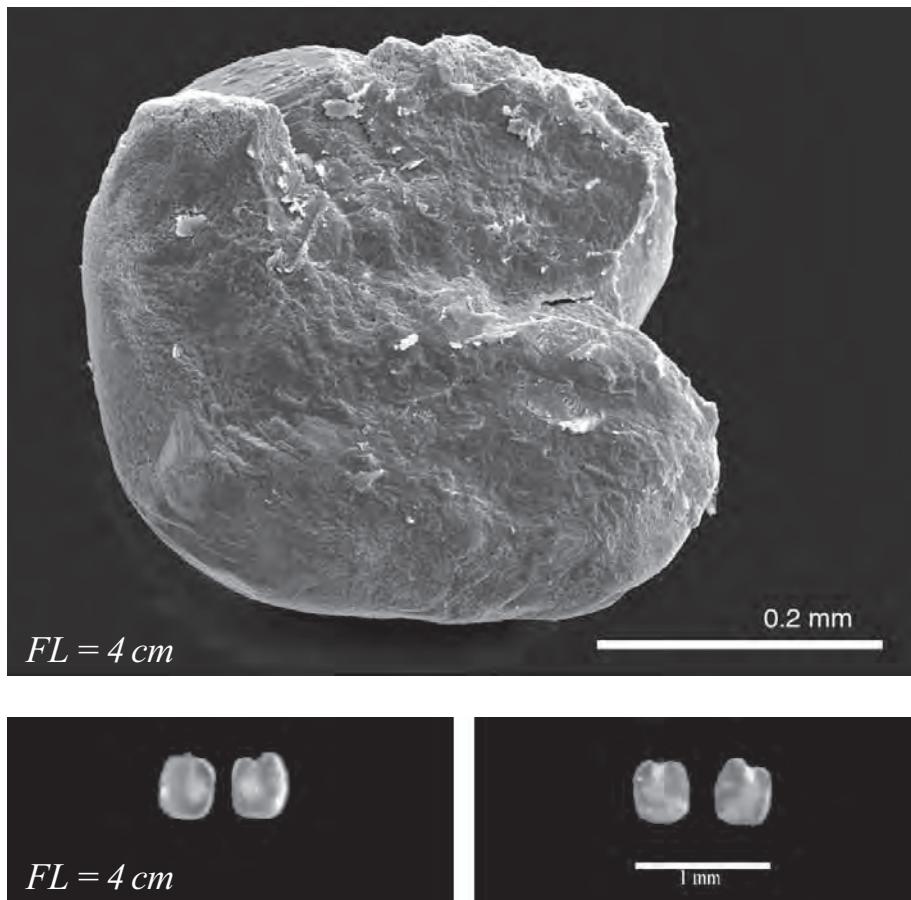
83

Gasterosteidae: *Gasterosteus aculeatus*



Gasterosteiformes

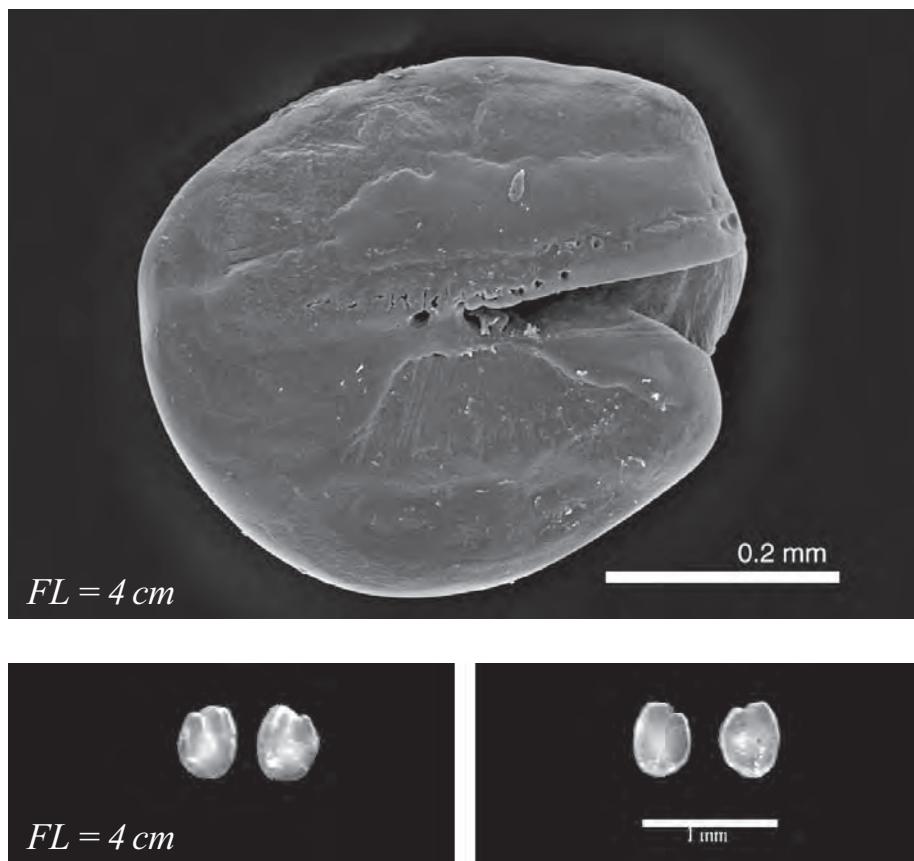
Gasterosteidae: *Gasterosteus wheatlandi*



Gasterosteiformes

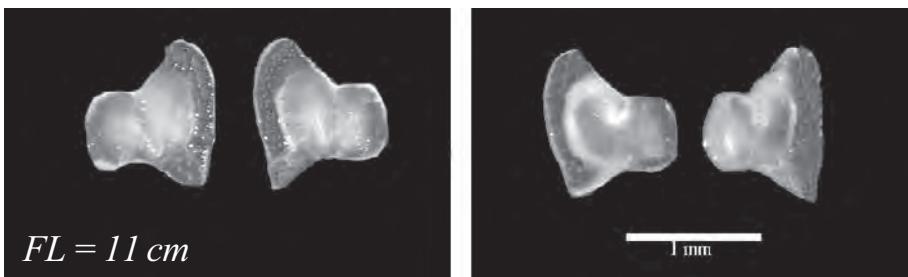
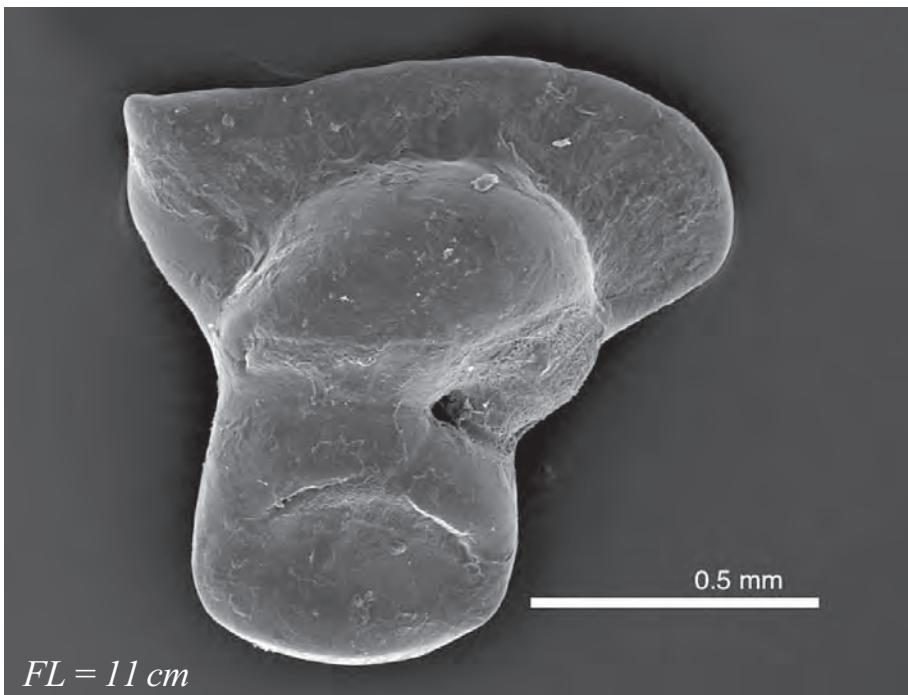
85

Gasterosteidae: *Pungitius pungitius*



Gasterosteiformes

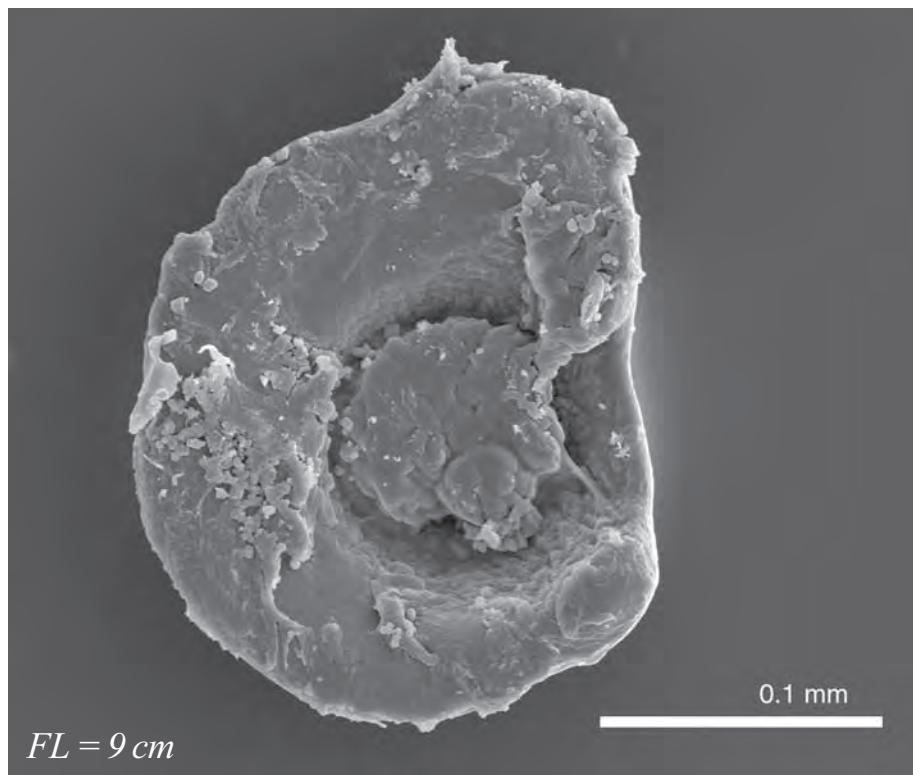
Macrorhamphosidae: *Macrorhamphosus scolopax*



Gasterosteiformes

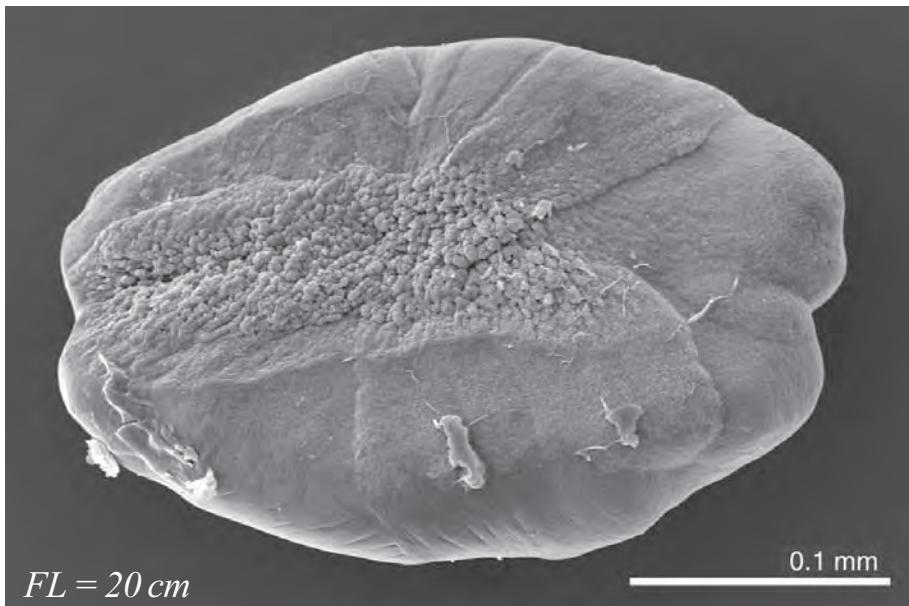
87

Syngnathidae: *Hippocampus erectus*



Gasterosteiformes

Syngnathidae: *Syngnathus fuscus*

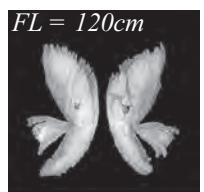


Lampridiformes

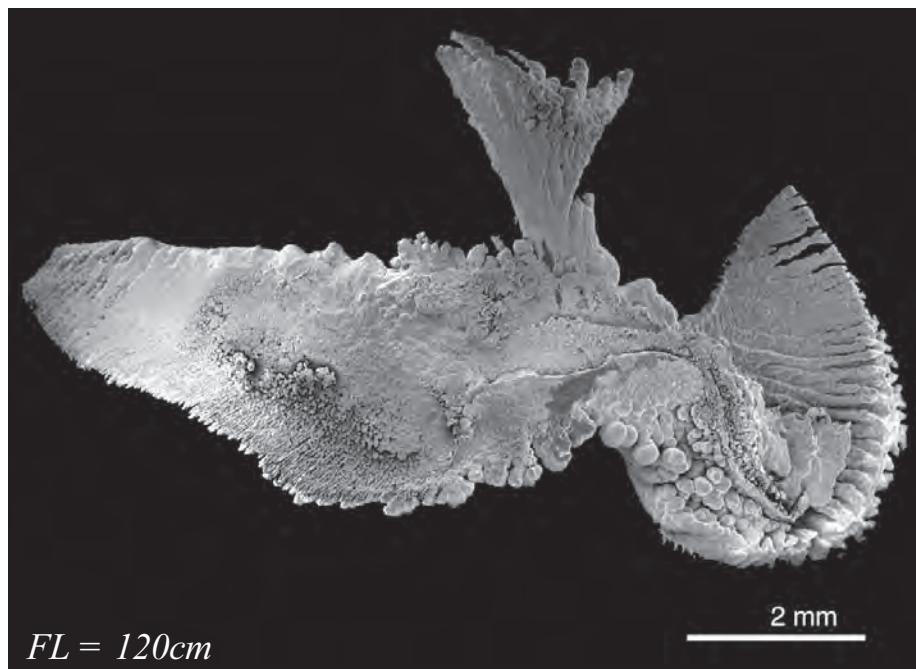
Lampridae: *Lampris guttatus*

89

Asterisci

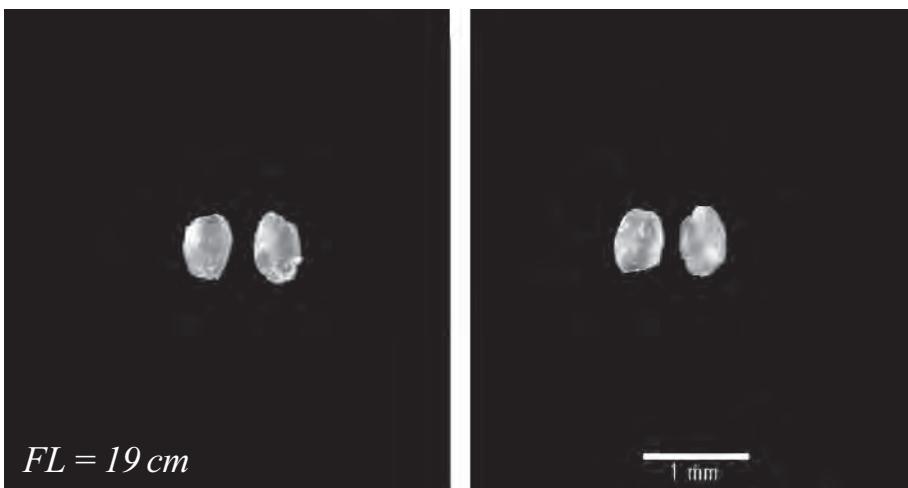
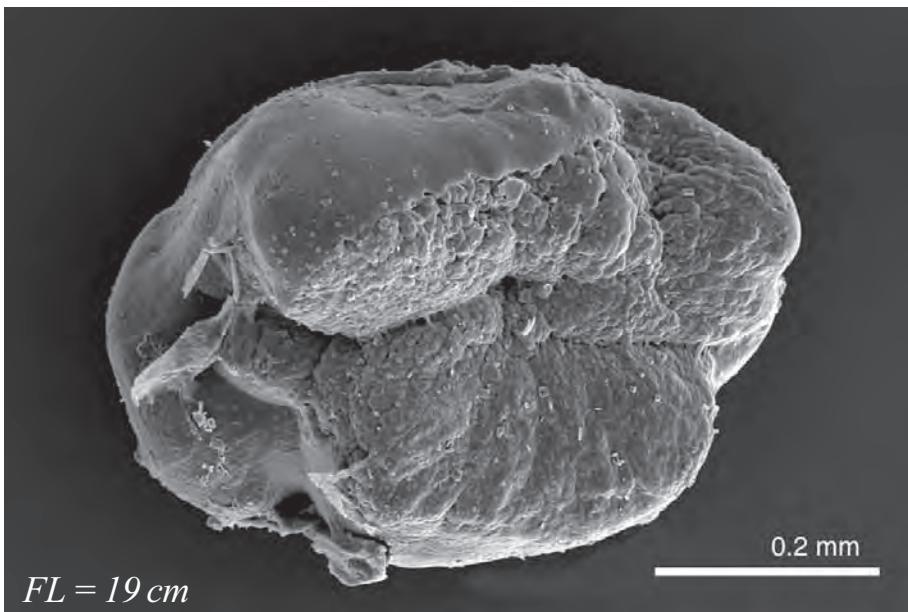


Lapilli



Lophiiformes

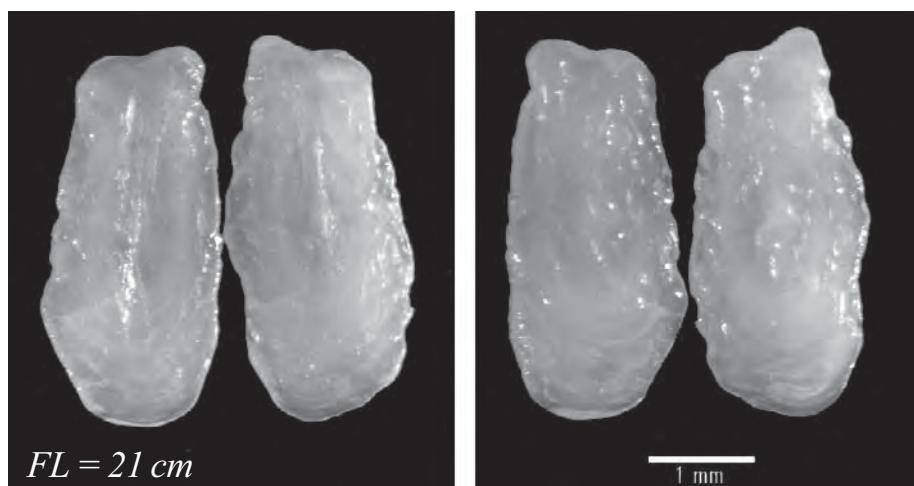
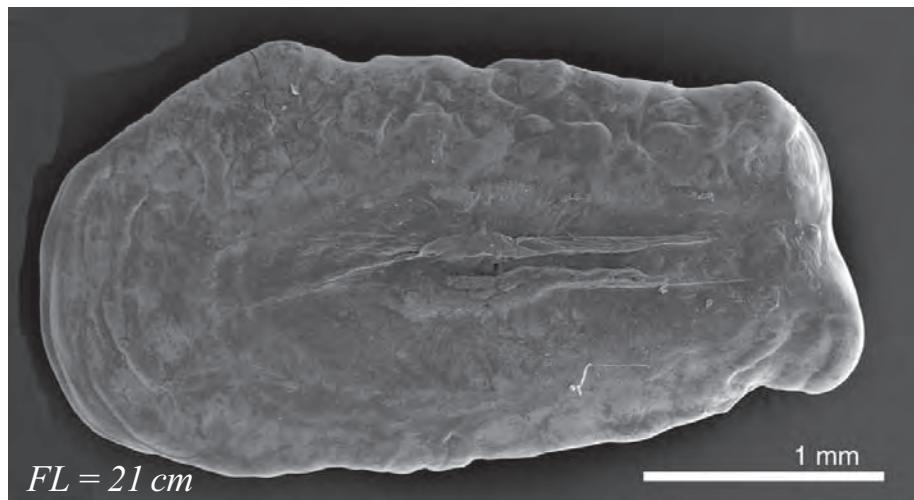
Ceratiidae: *Ceratias holboelli*



Lophiiformes

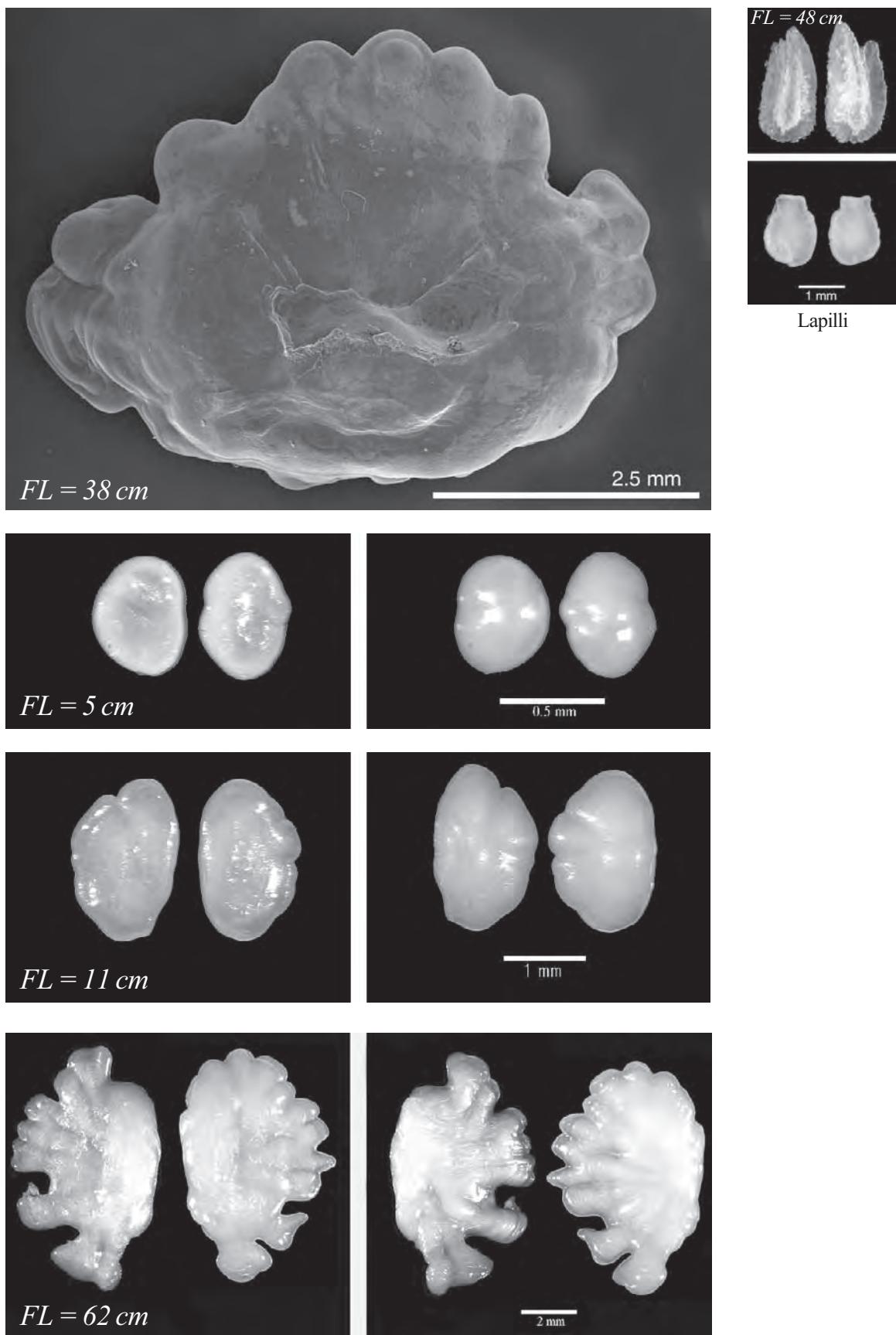
Ceratiidae: *Cryptopsaras couesi*

91



Lophiiformes

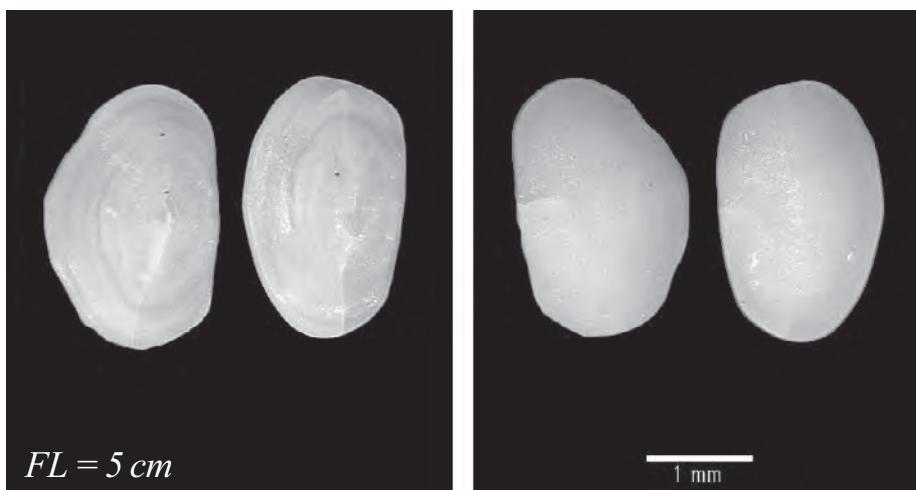
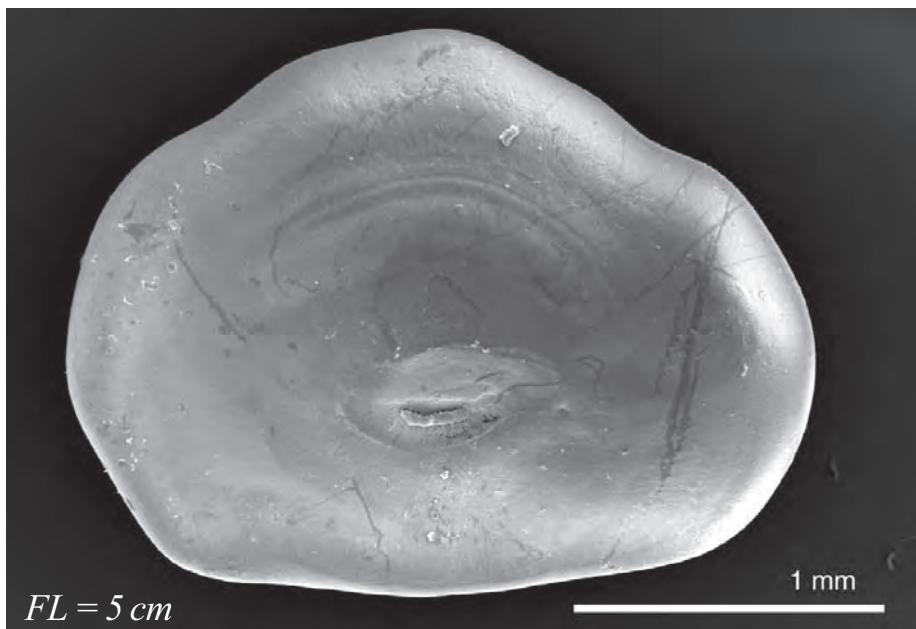
Lophiidae: *Lophius americanus*



Lophiiformes

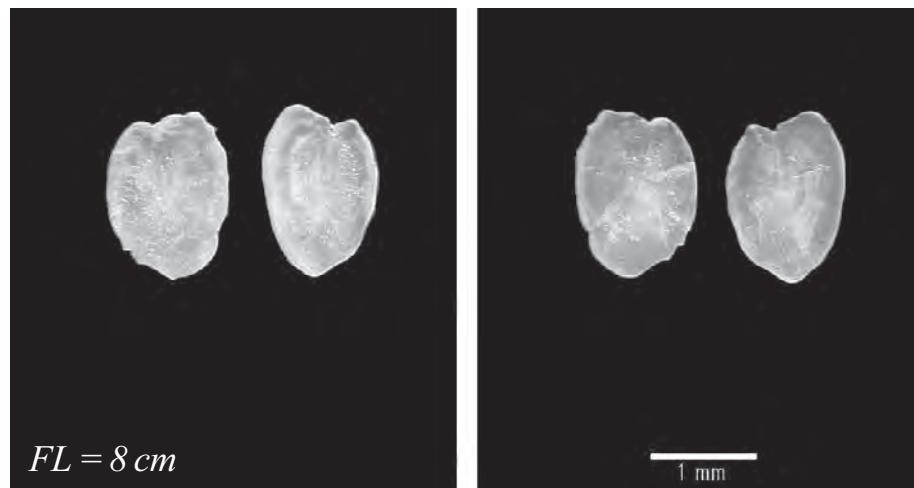
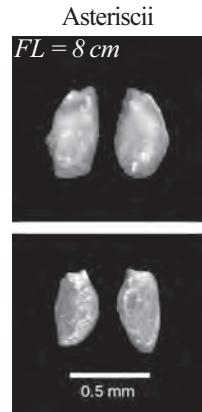
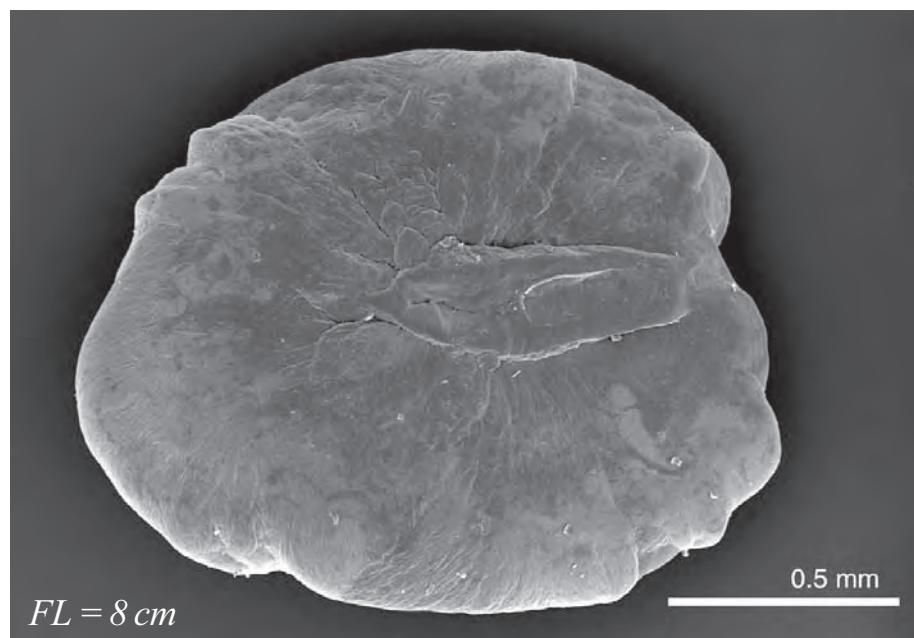
93

Ogcocephalidae: *Dibranchus atlanticus*



Lophiiformes

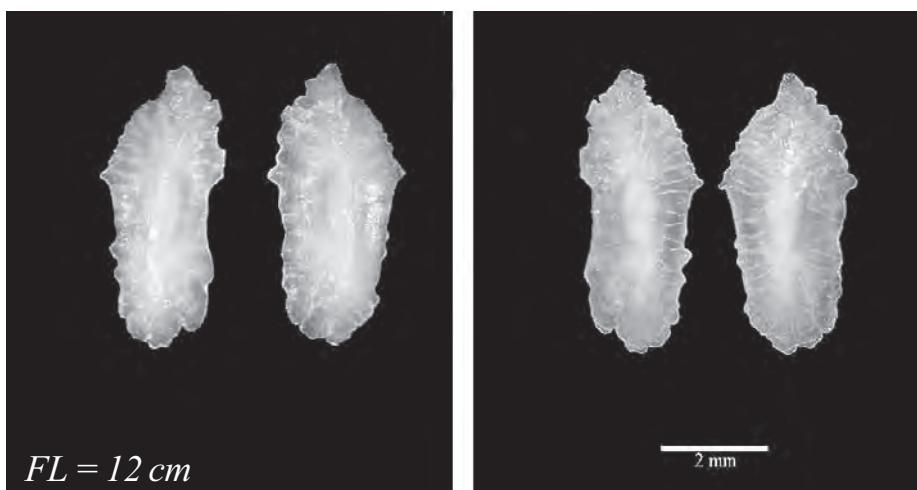
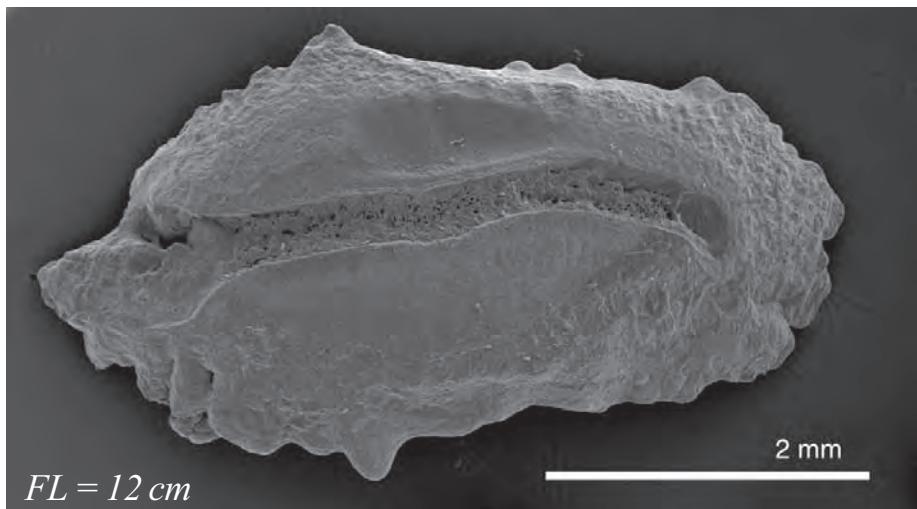
Oneirodidae: *Oneirodes* sp.



Mugiliformes

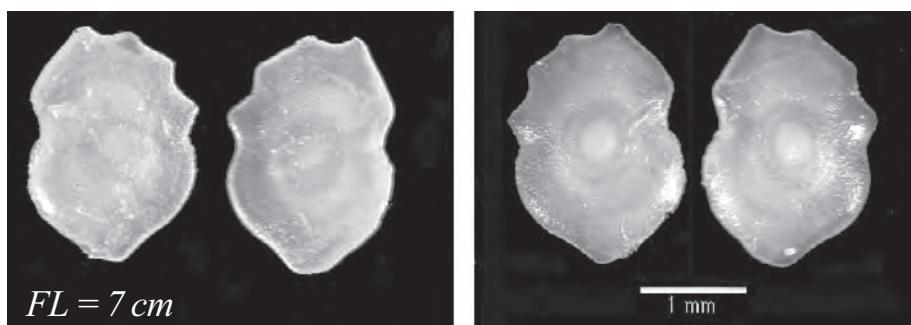
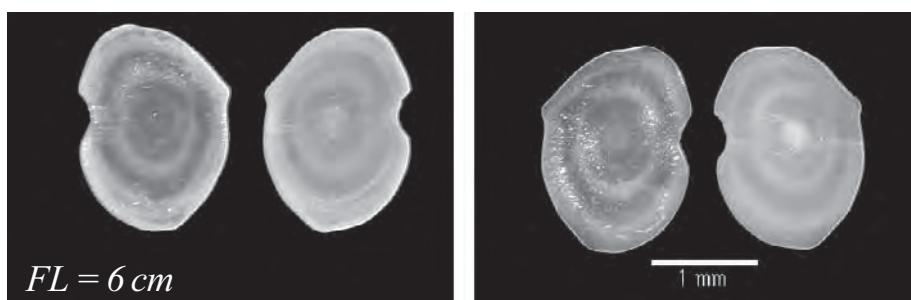
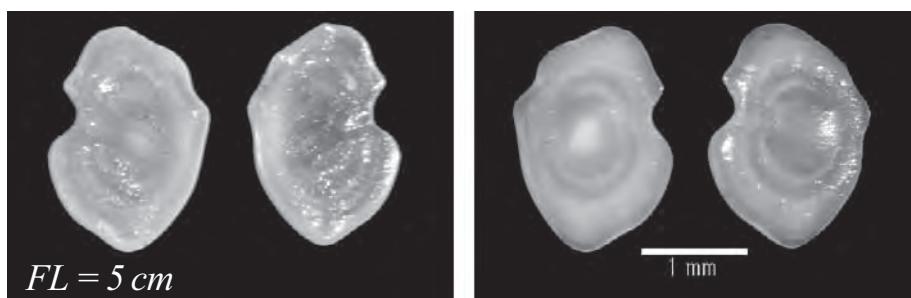
95

Mugilidae: *Mugil curema*



Myctophiformes

Myctophidae: *Benthosema glaciale*

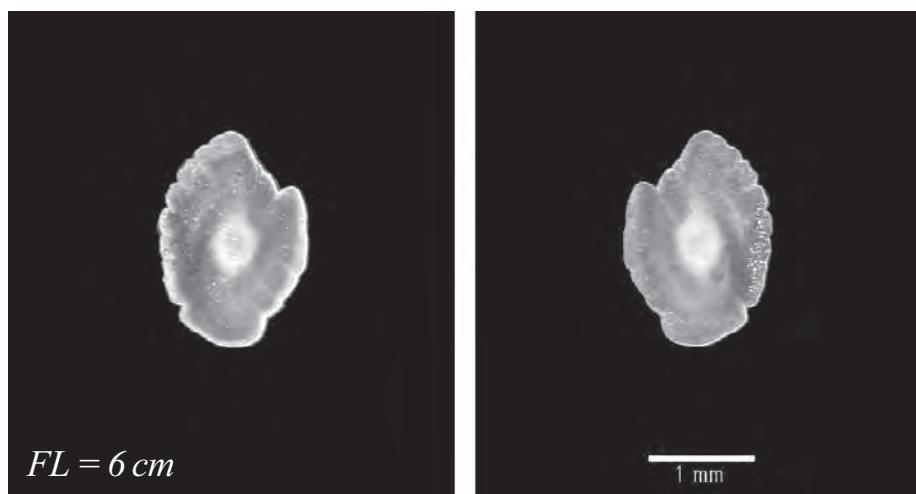
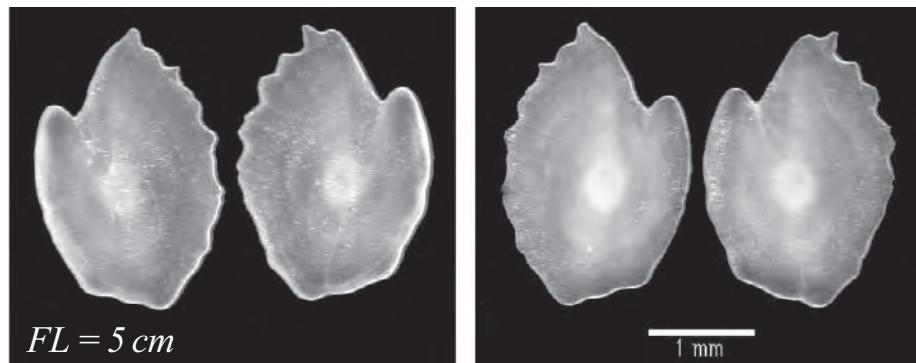


Myctophiformes

Myctophidae: *Benthosema suborbitale*

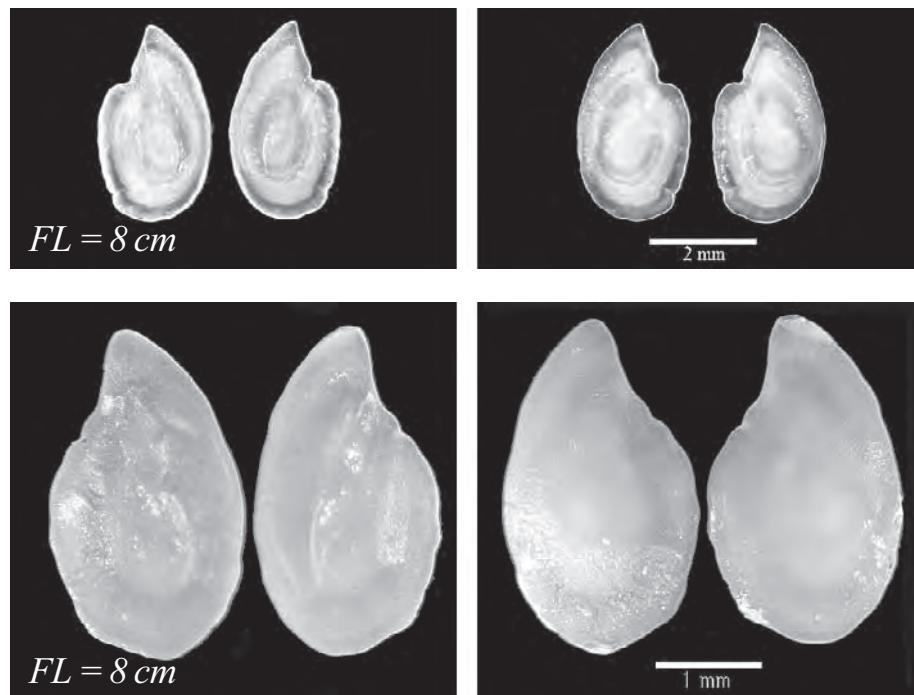


Myctophidae: *Bolinichthys photothorax*

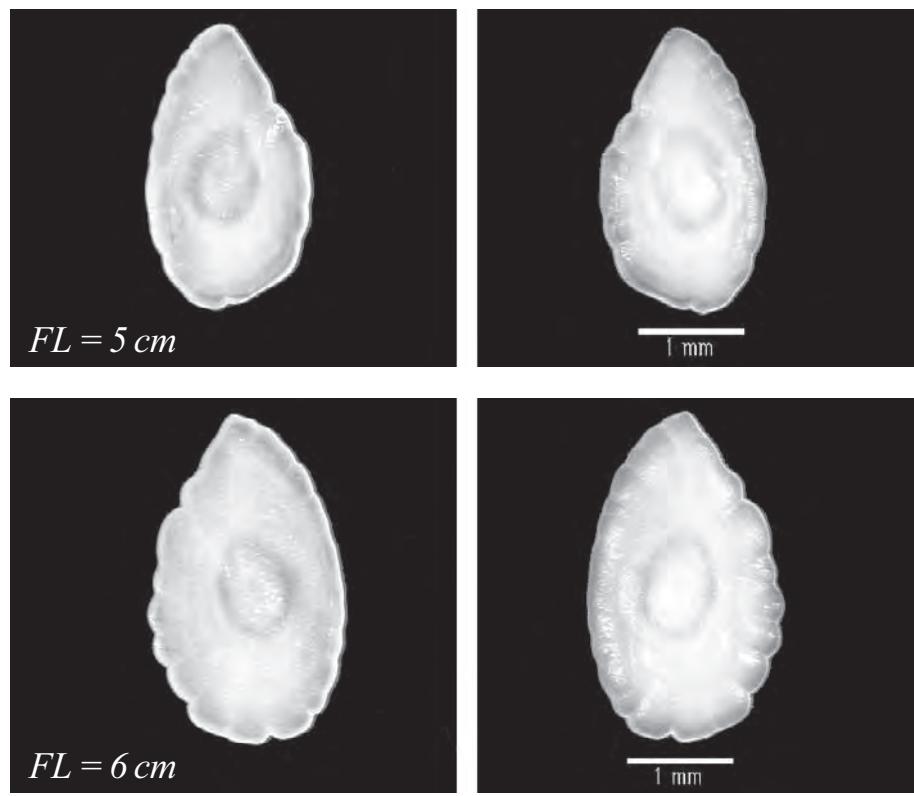


Myctophiformes

Myctophidae: *Ceratoscopelus maderensis*

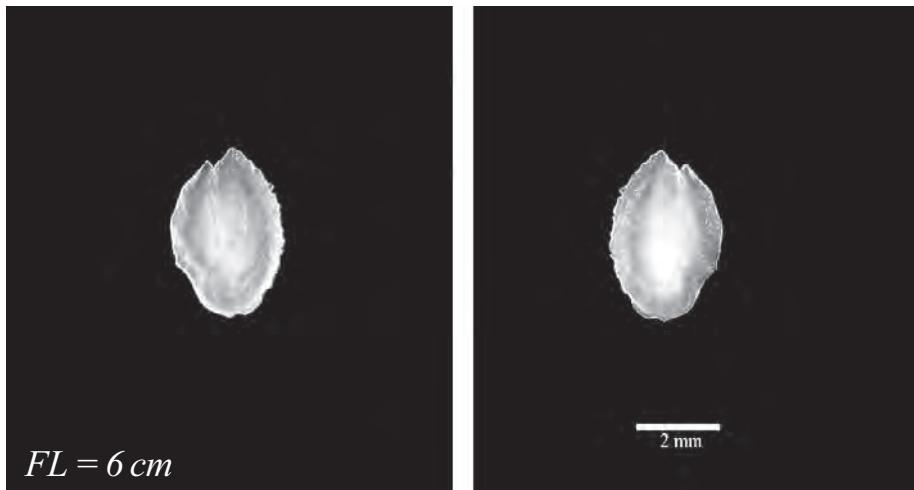
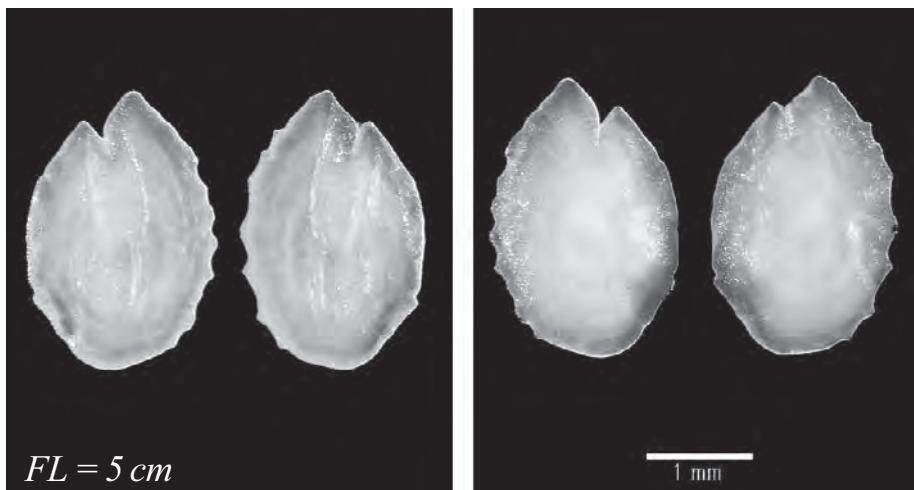
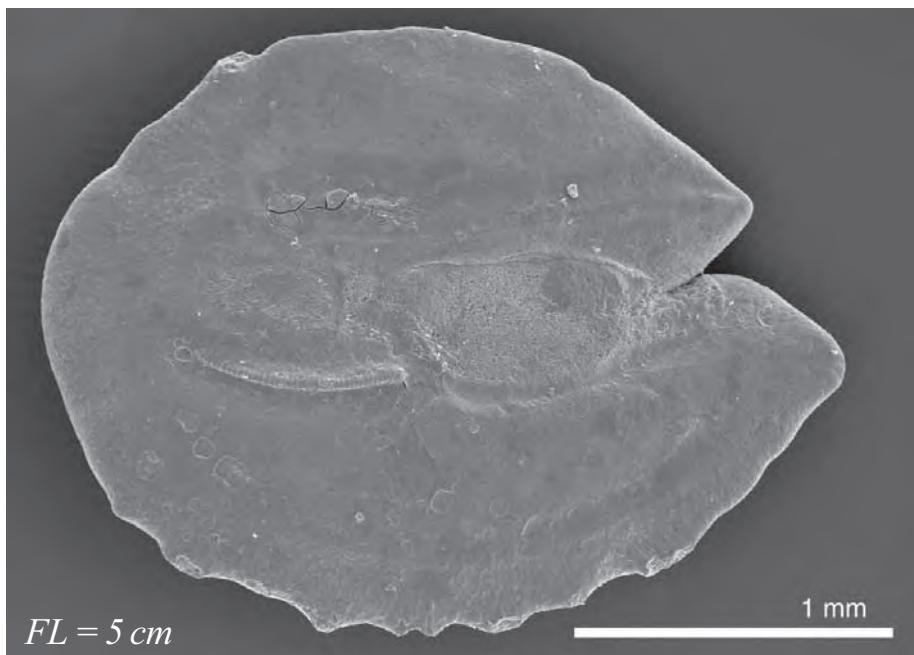


Myctophidae: *Ceratoscopelus warmingii*



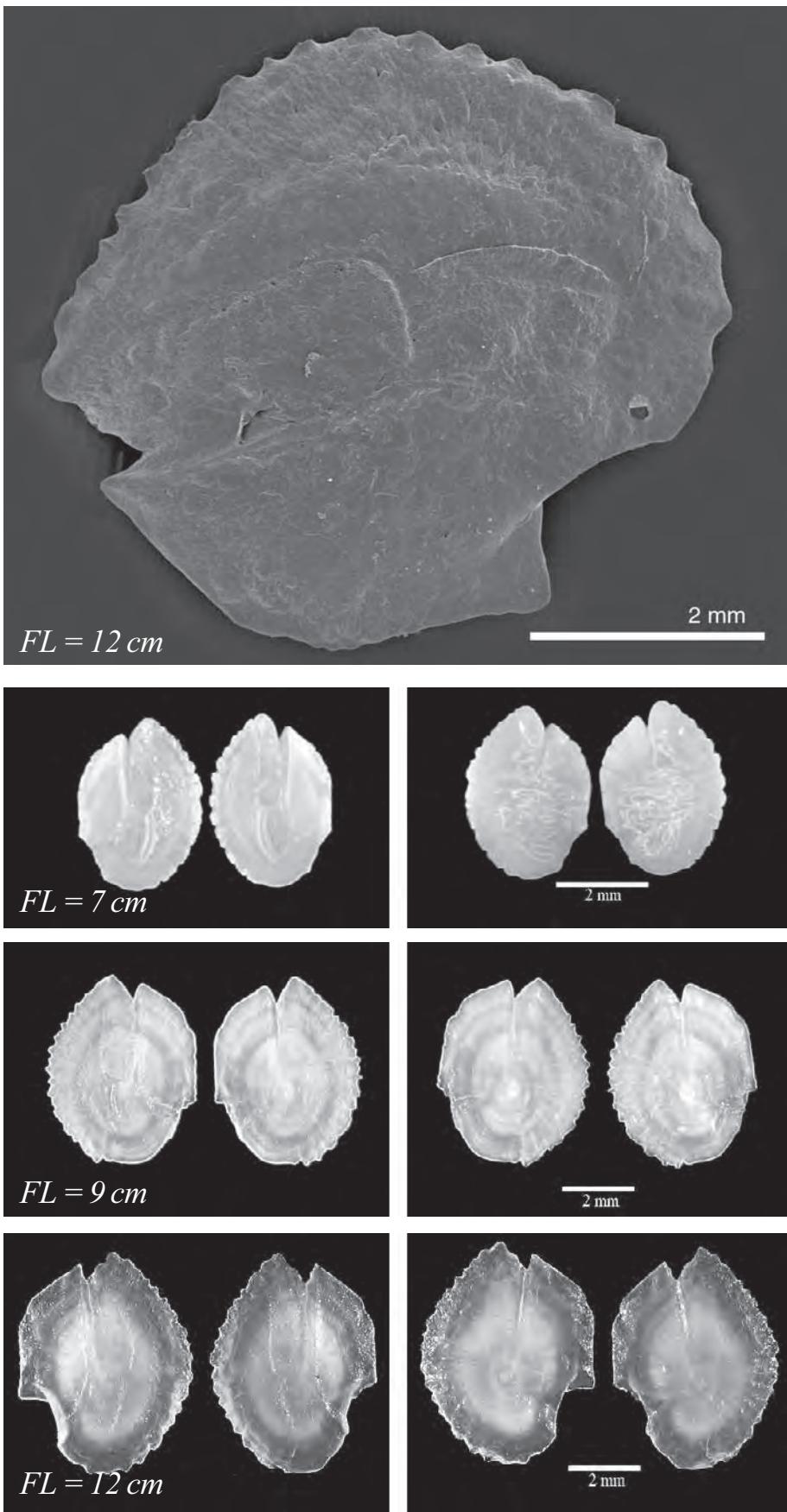
Myctophiformes

Myctophidae: *Diaphus dumerilii*



Myctophiformes

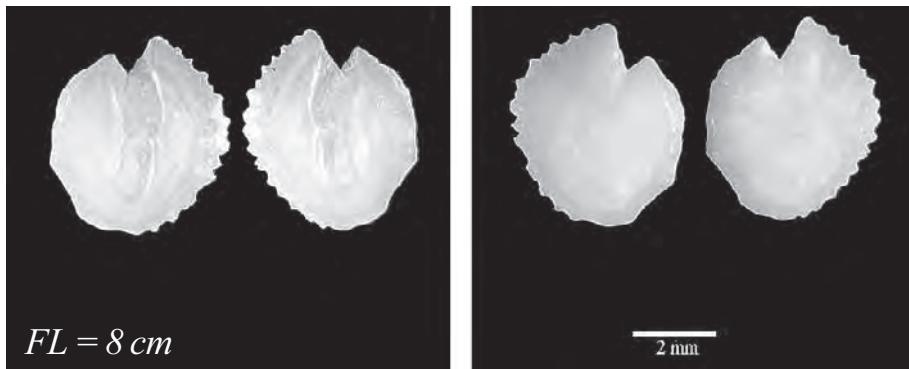
Myctophidae: *Diaphus effulgens*



Myctophiformes

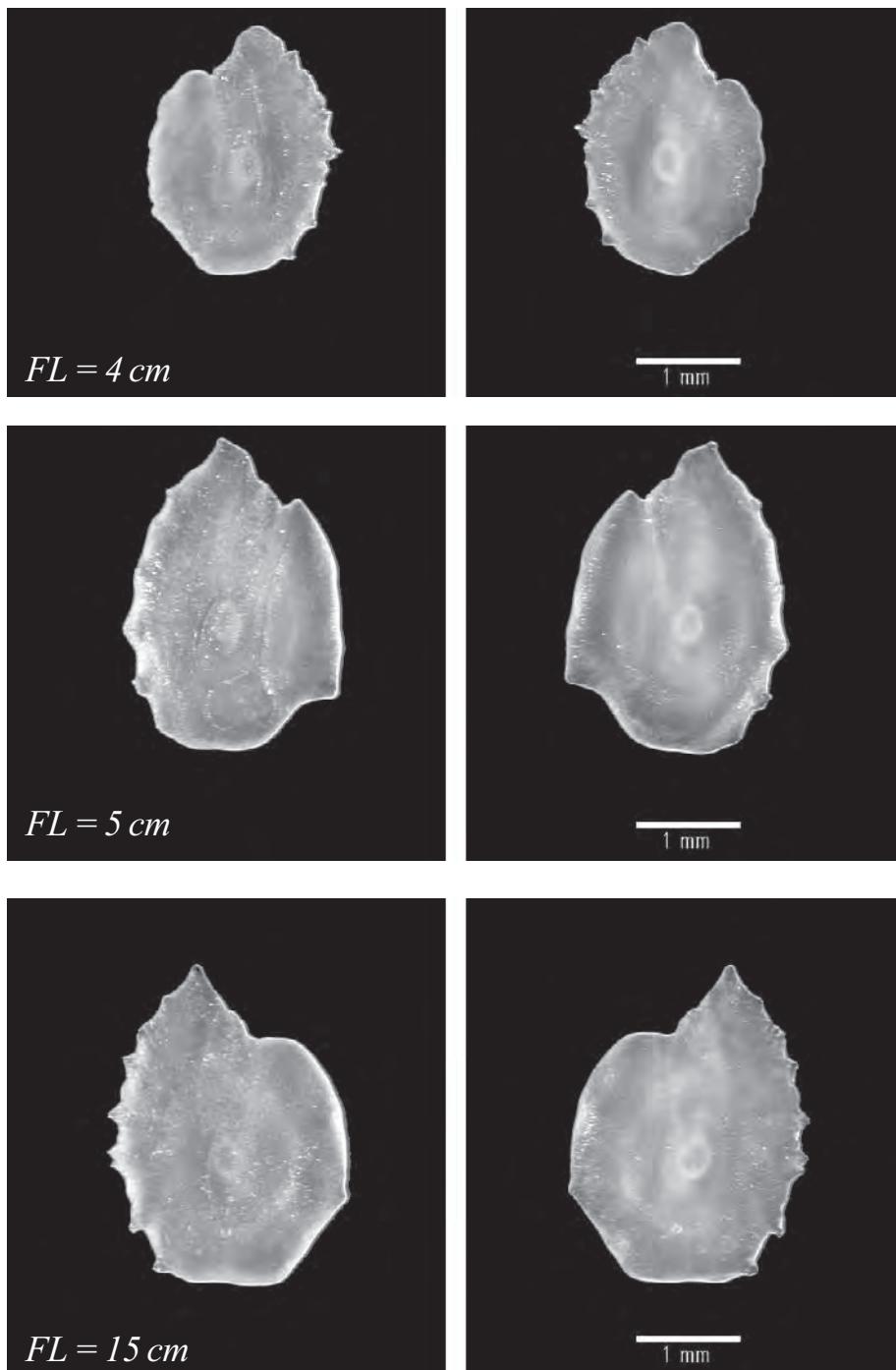
101

Myctophidae: *Diaphus metopoclampus*



Myctophiformes

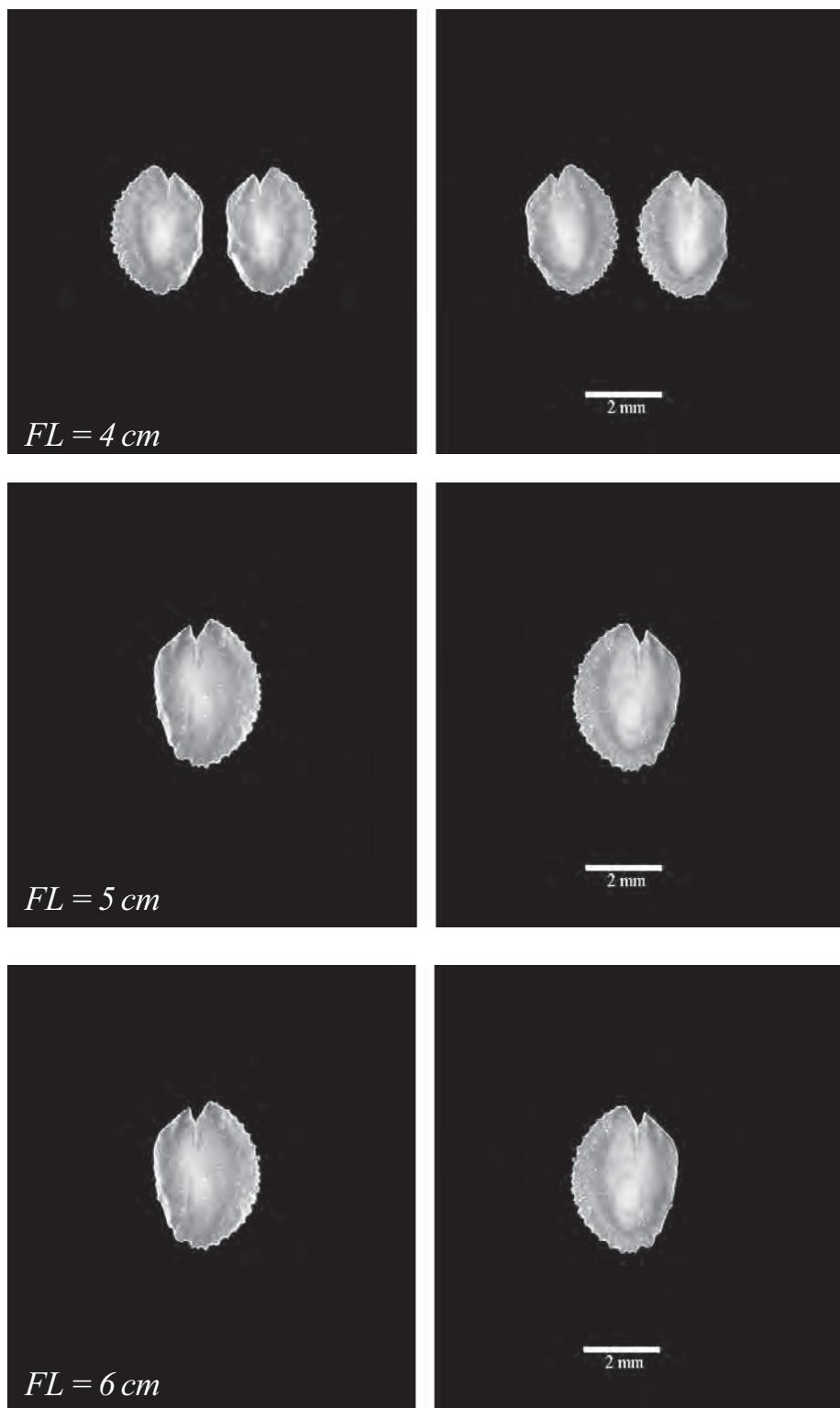
Myctophidae: *Diaphus mollis*



Myctophiformes

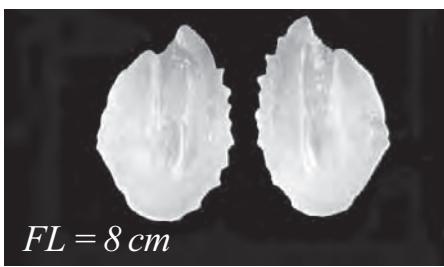
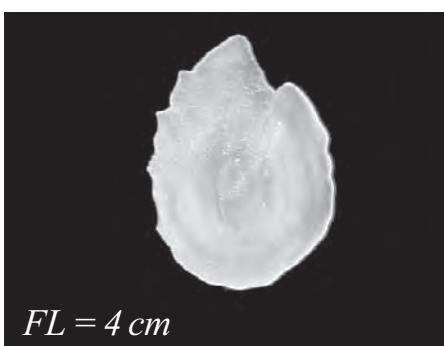
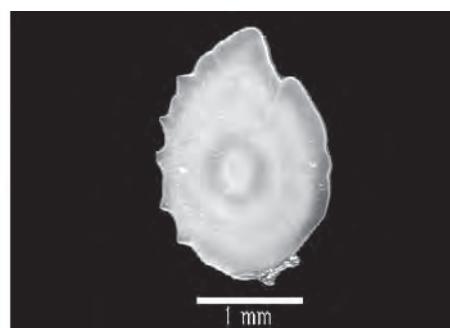
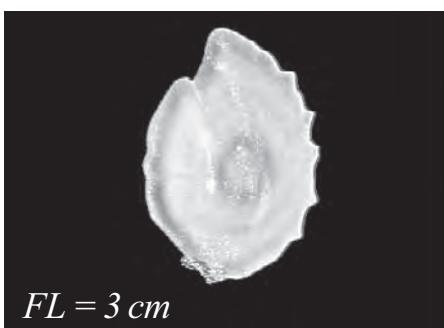
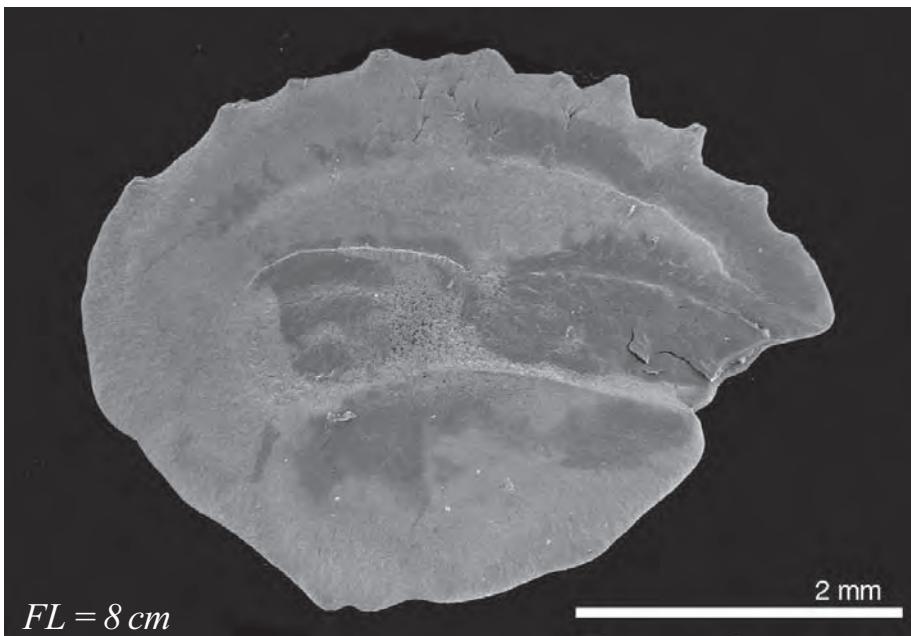
103

Myctophidae: *Diaphus perspicillatus*



Myctophiformes

Myctophidae: *Diaphus rafinesquii*



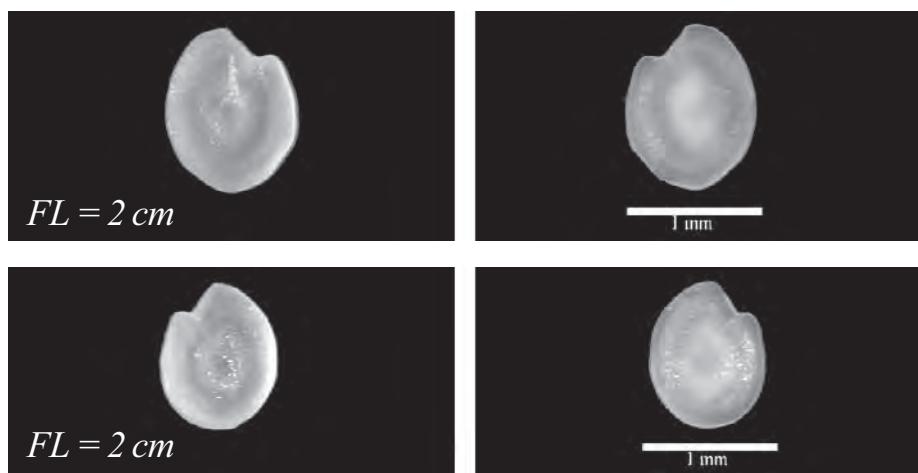
Myctophiformes

105

Myctophidae: *Diaphus termophilus*

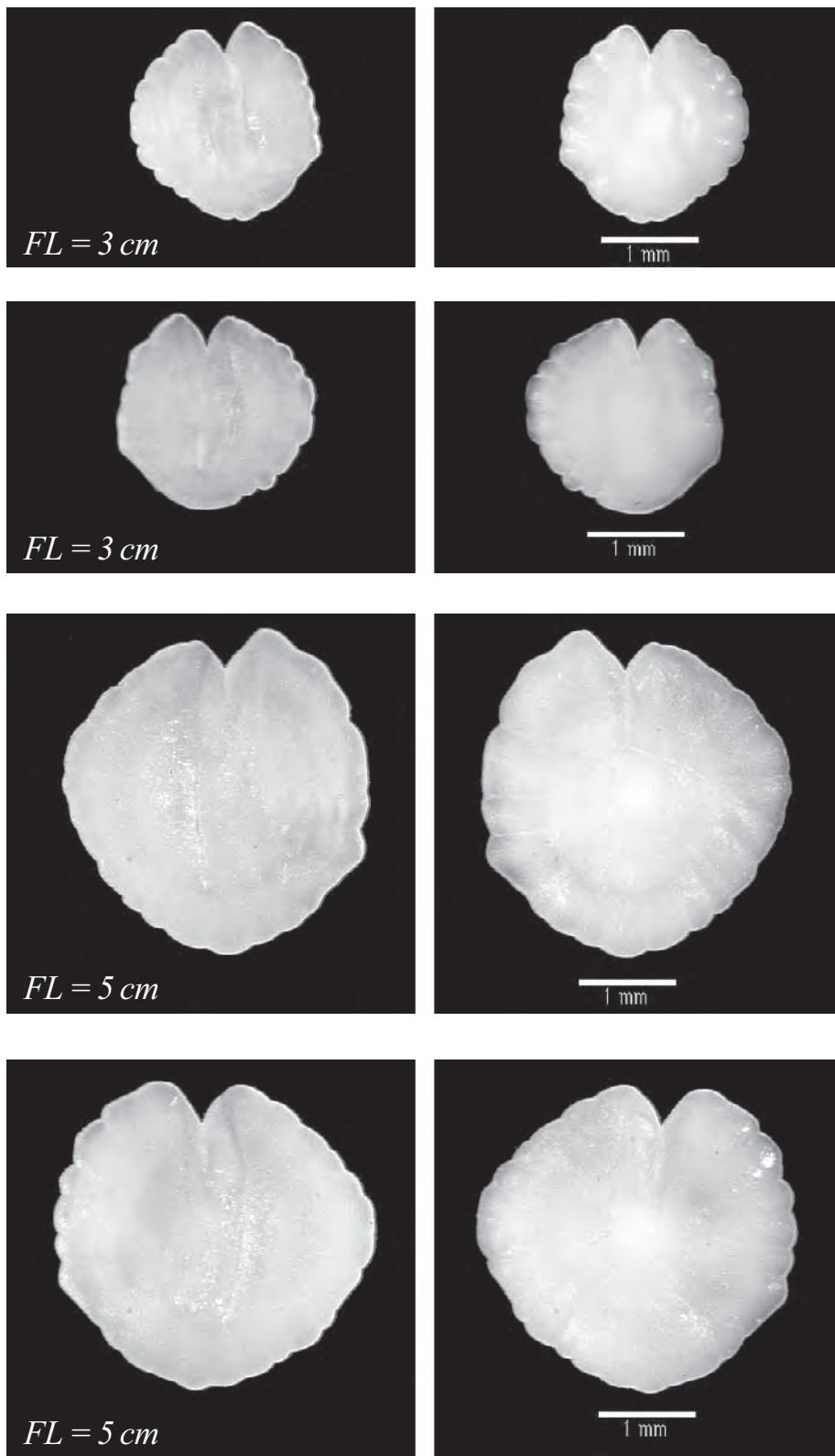


Myctophidae: *Diogenichthys atlanticus*



Myctophiformes

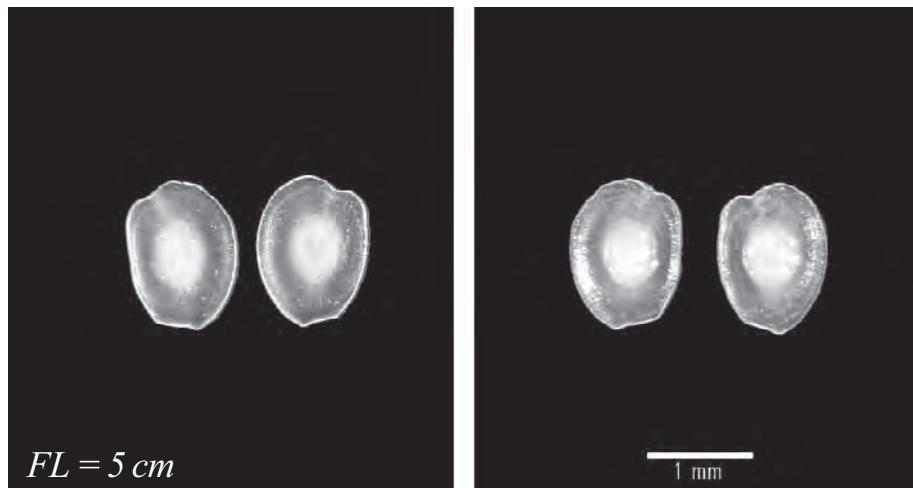
Myctophidae: *Electrona risso*



Myctophiformes

107

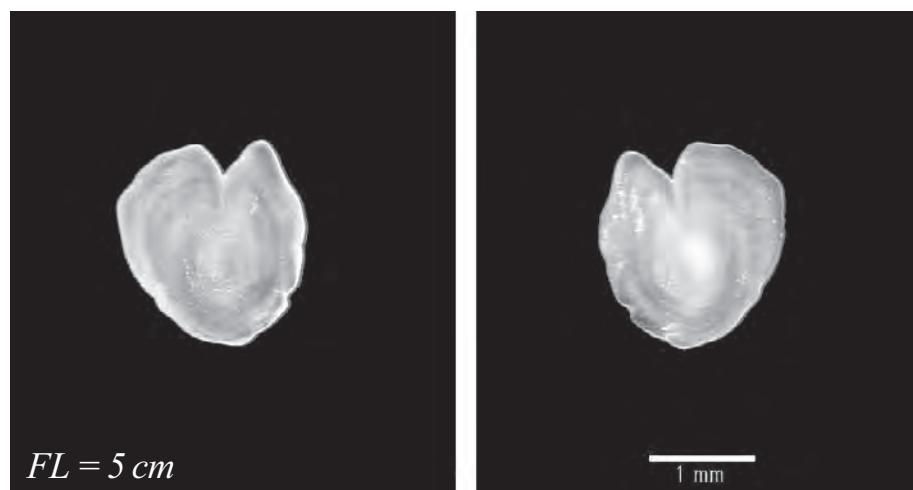
Myctophidae: *Gonichthysocco*



FL = 5 cm

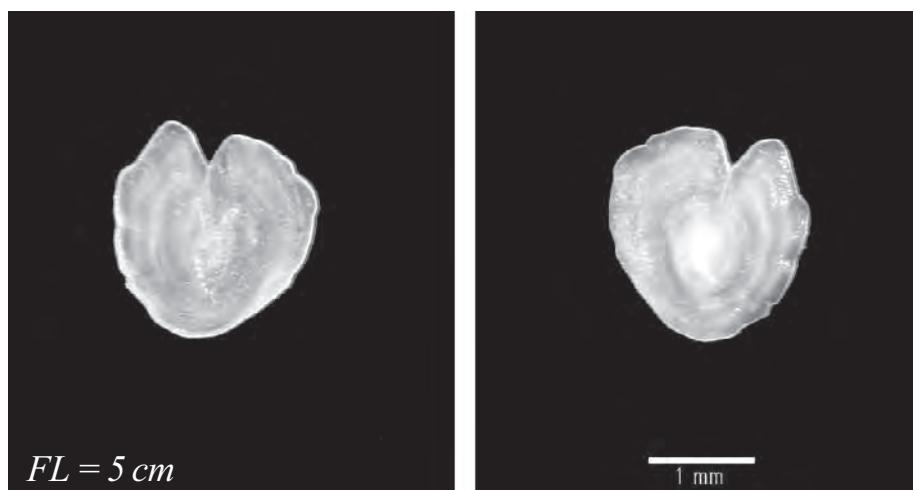
1 mm

Myctophidae: *Hygophum benoiti*



FL = 5 cm

1 mm

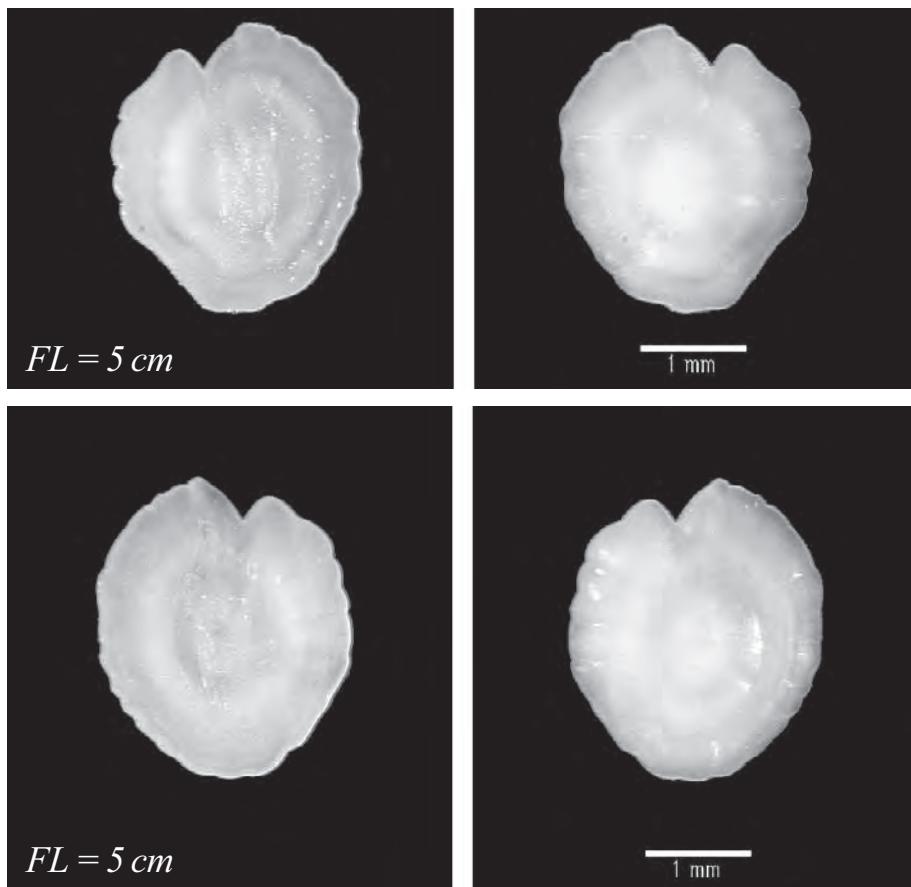


FL = 5 cm

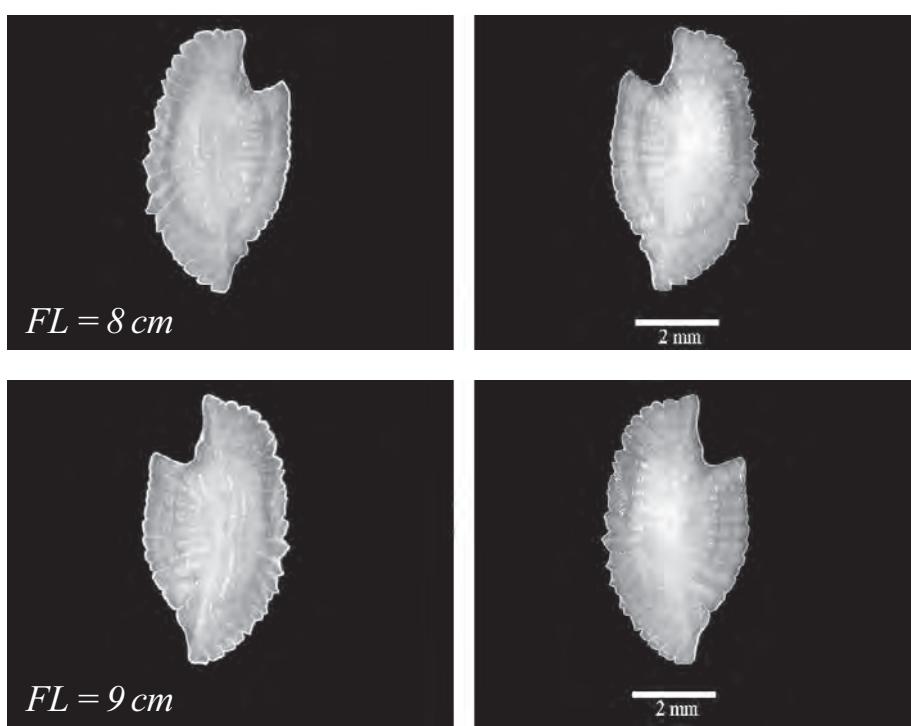
1 mm

Myctophiformes

Myctophidae: *Hygophum hygomii*



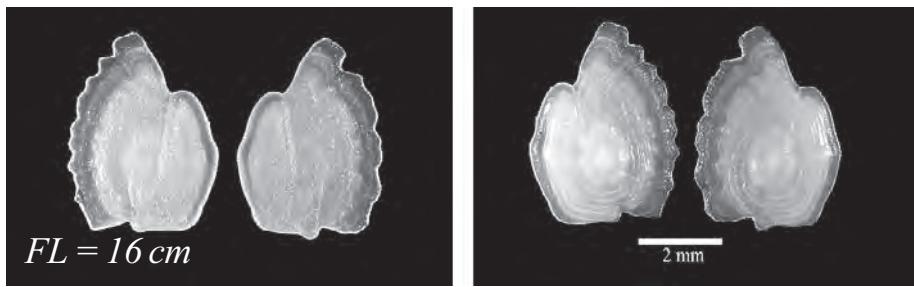
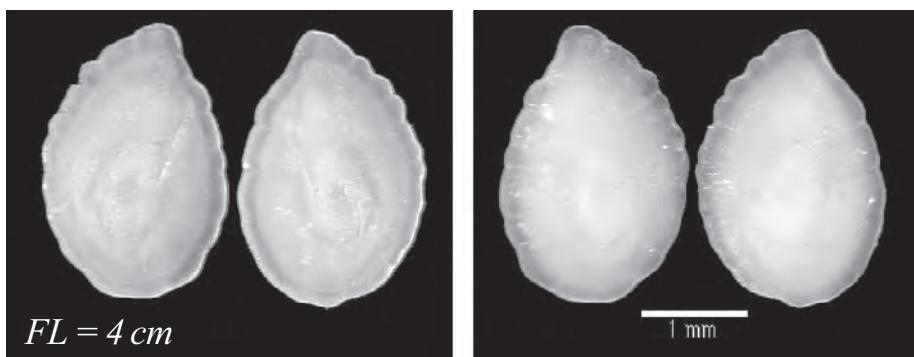
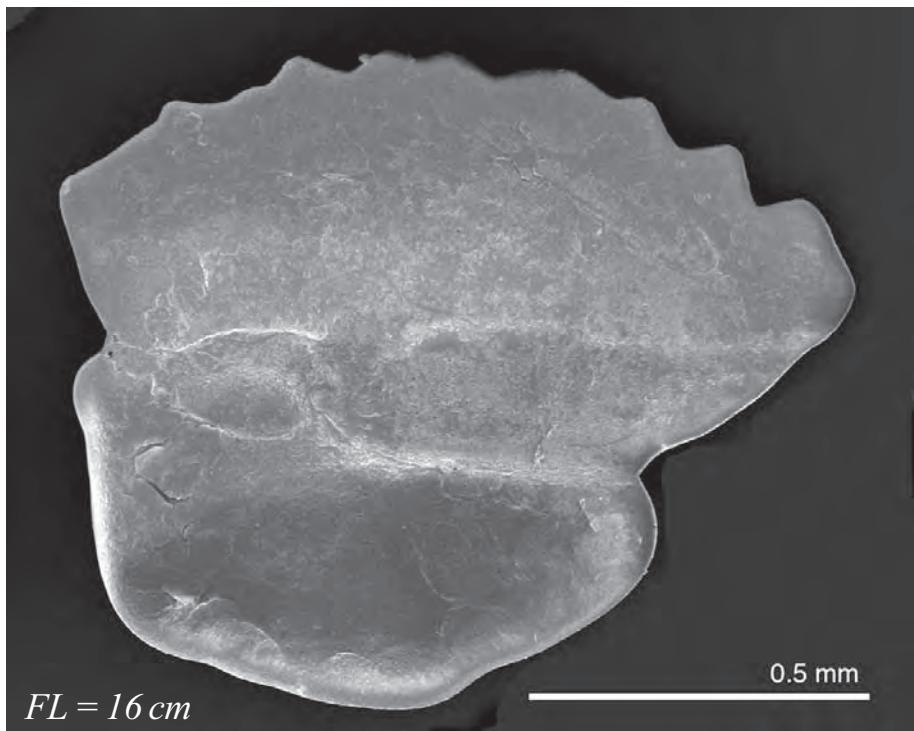
Myctophidae: *Lampadена luminosa*



Myctophiformes

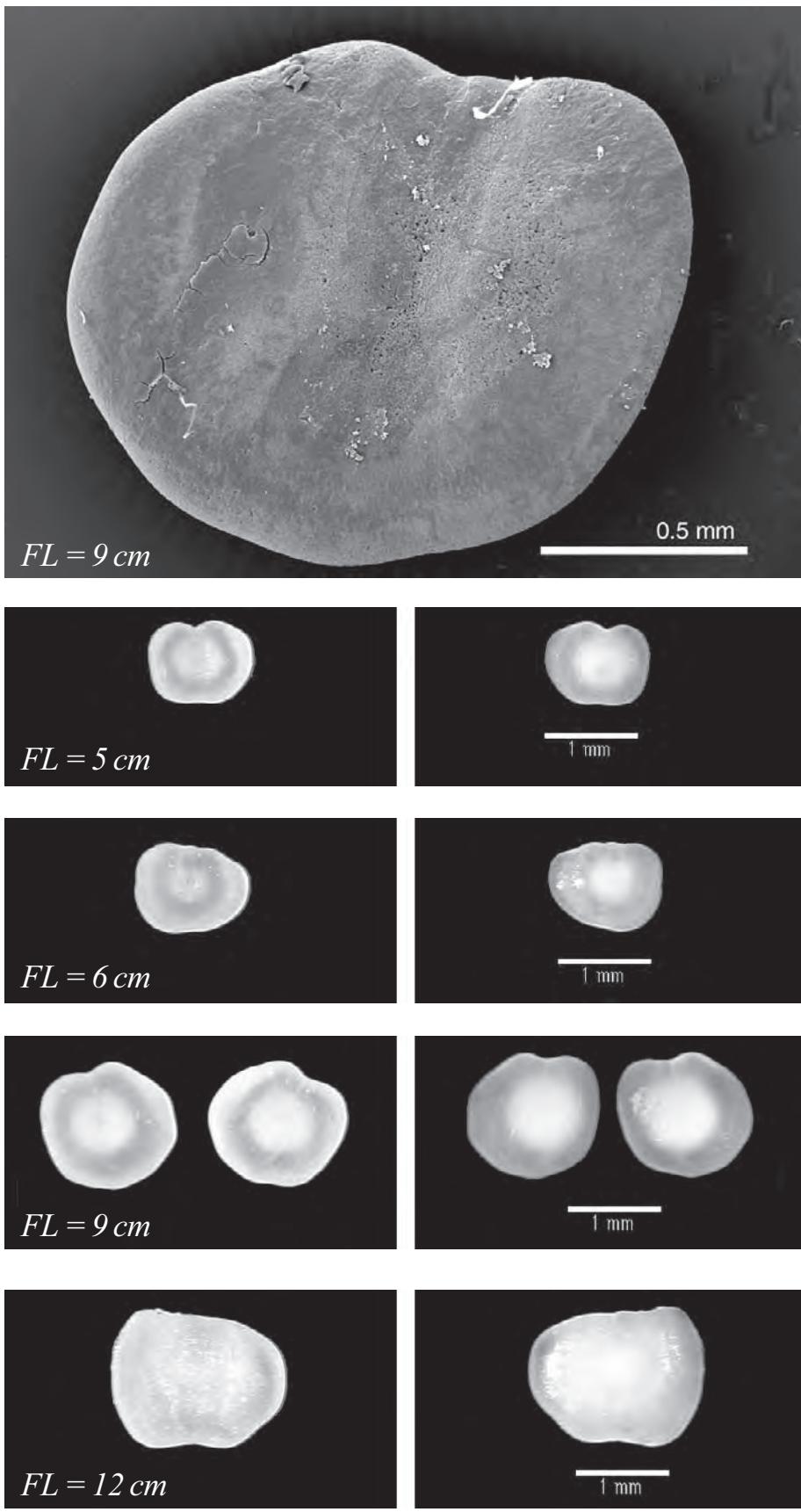
109

Myctophidae: *Lampadена speculigera*



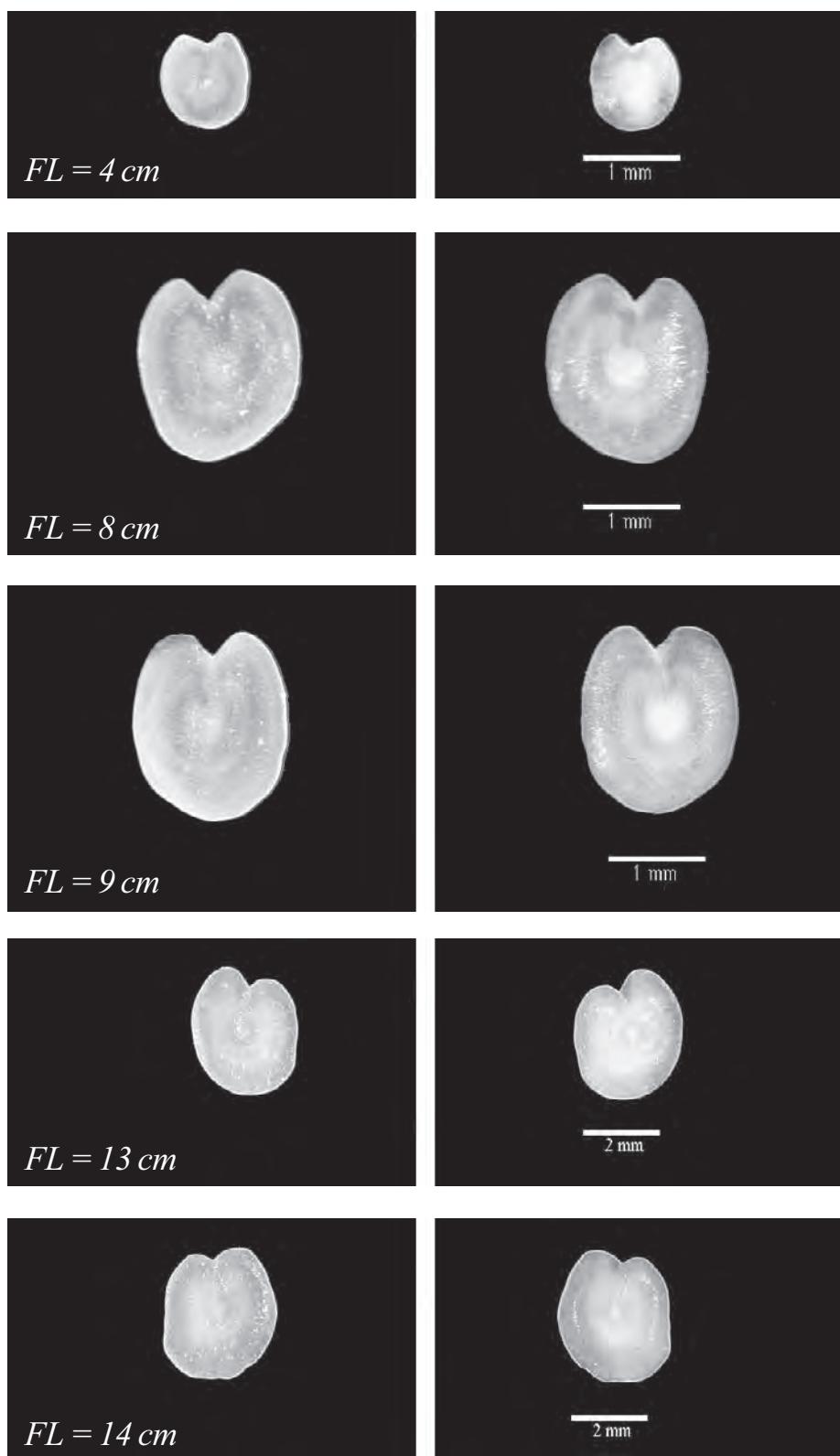
Myctophiformes

Myctophidae: *Lampanyctus ater*



Myctophiformes

Myctophidae: *Lampanyctus crocodilus*

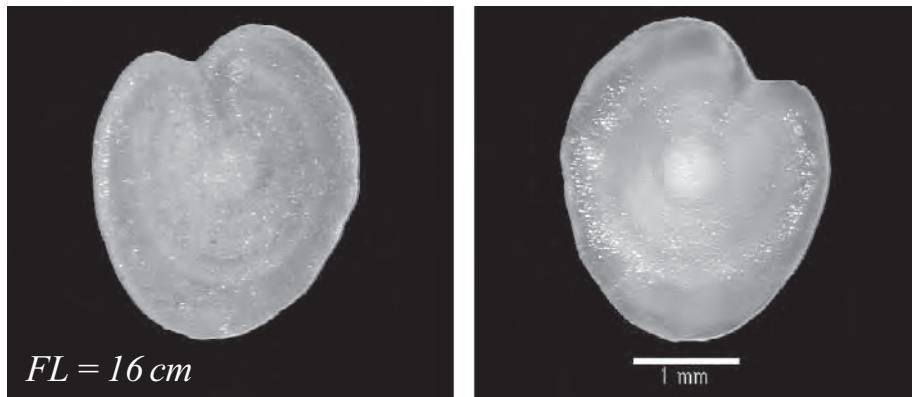
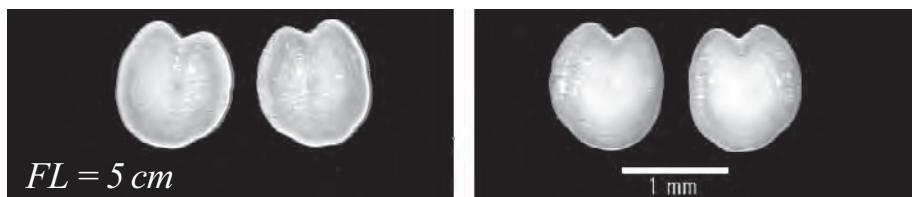


Myctophiformes

Myctophidae: *Lampanyctus festivus*



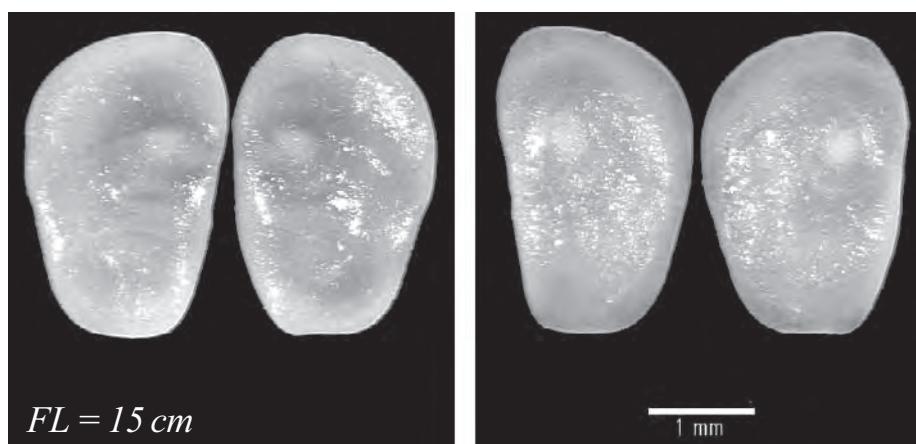
Myctophidae: *Lampanyctus intracarius*



Myctophiformes

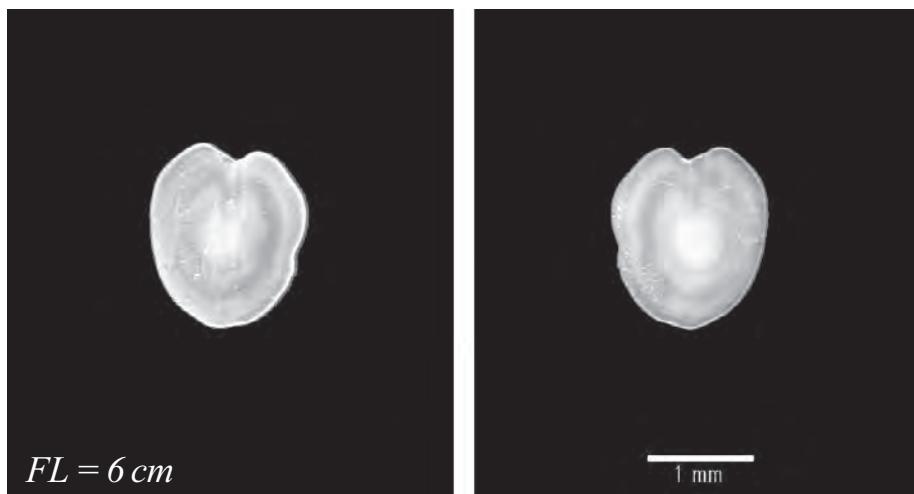
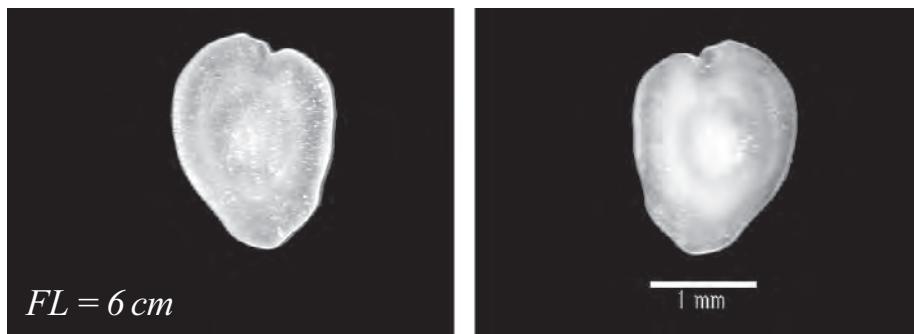
113

Myctophidae: *Lampanyctus macdonaldi*

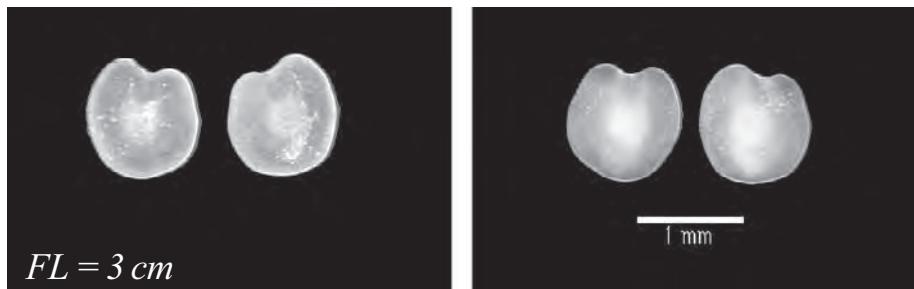


Myctophiformes

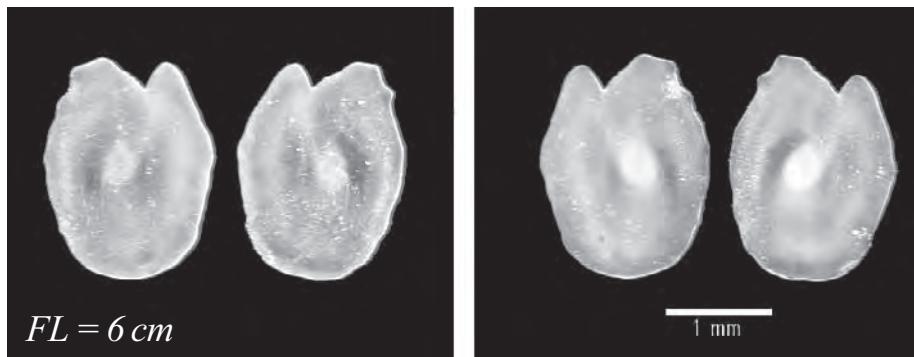
Myctophidae: *Lampanyctus photonotus*



Myctophidae: *Lampanyctus pusillus*



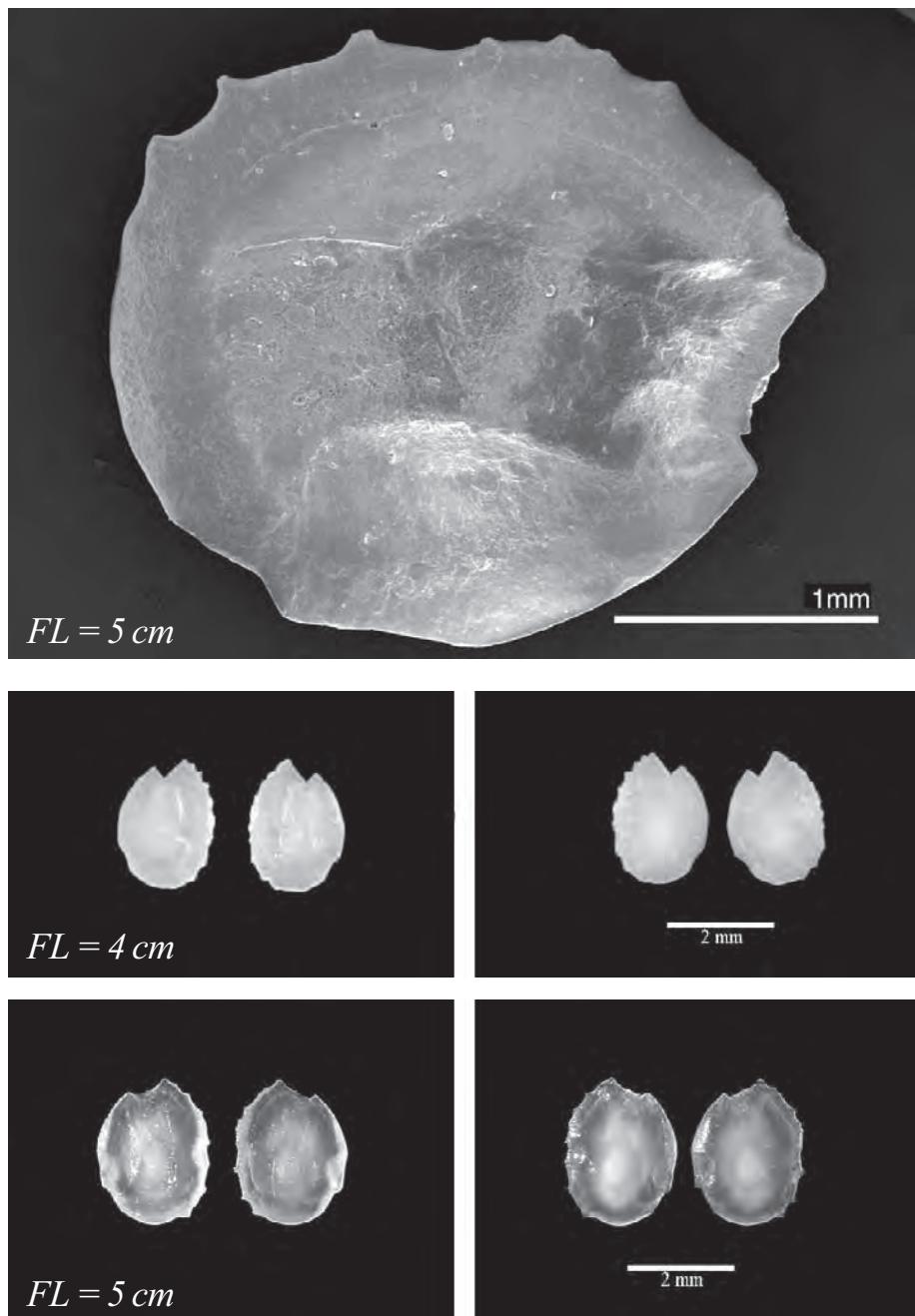
Myctophidae: *Lepidophanes guentheri*



Myctophiformes

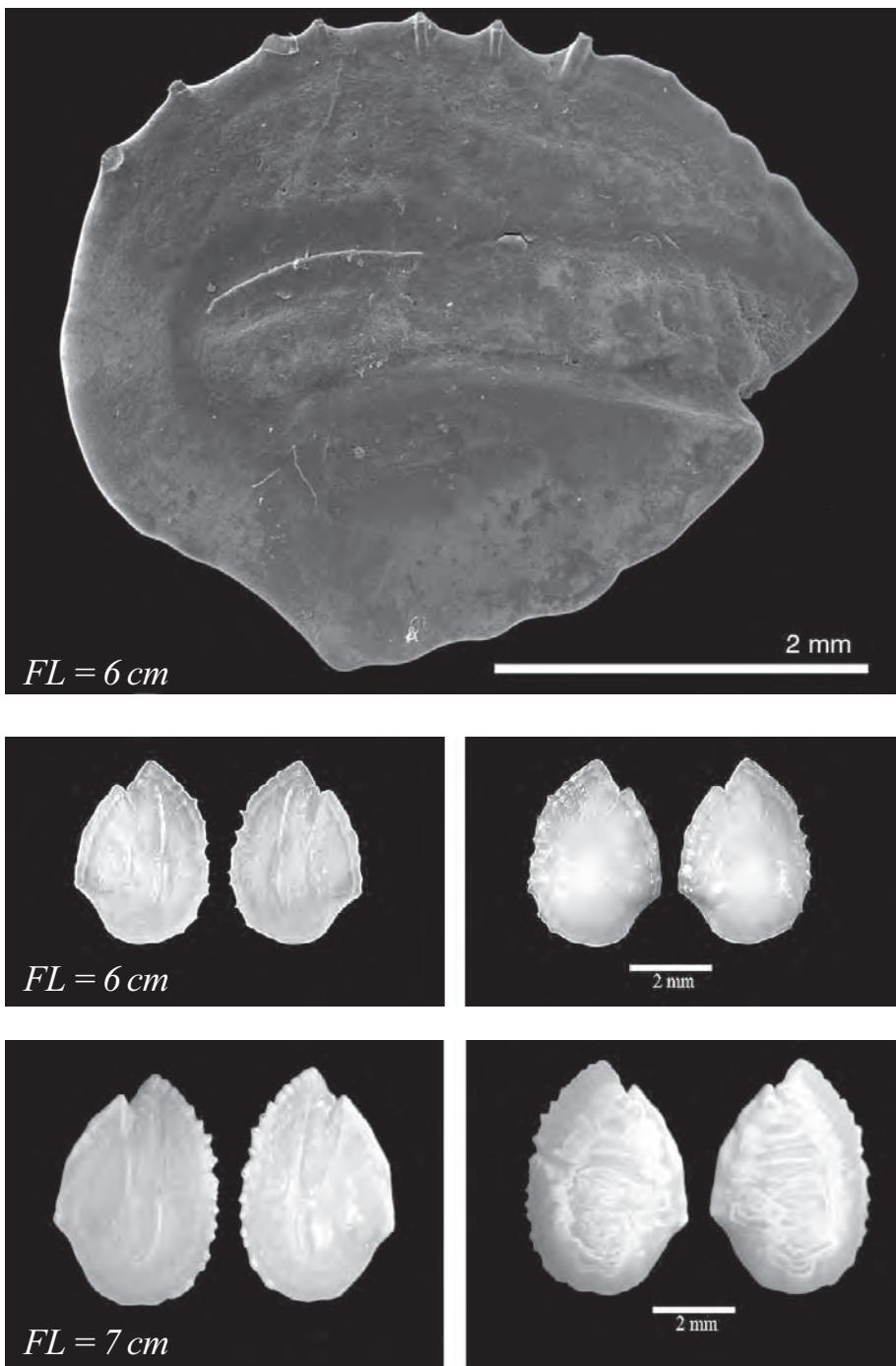
Myctophidae: *Lobianchia dofleini*

115



Myctophiformes

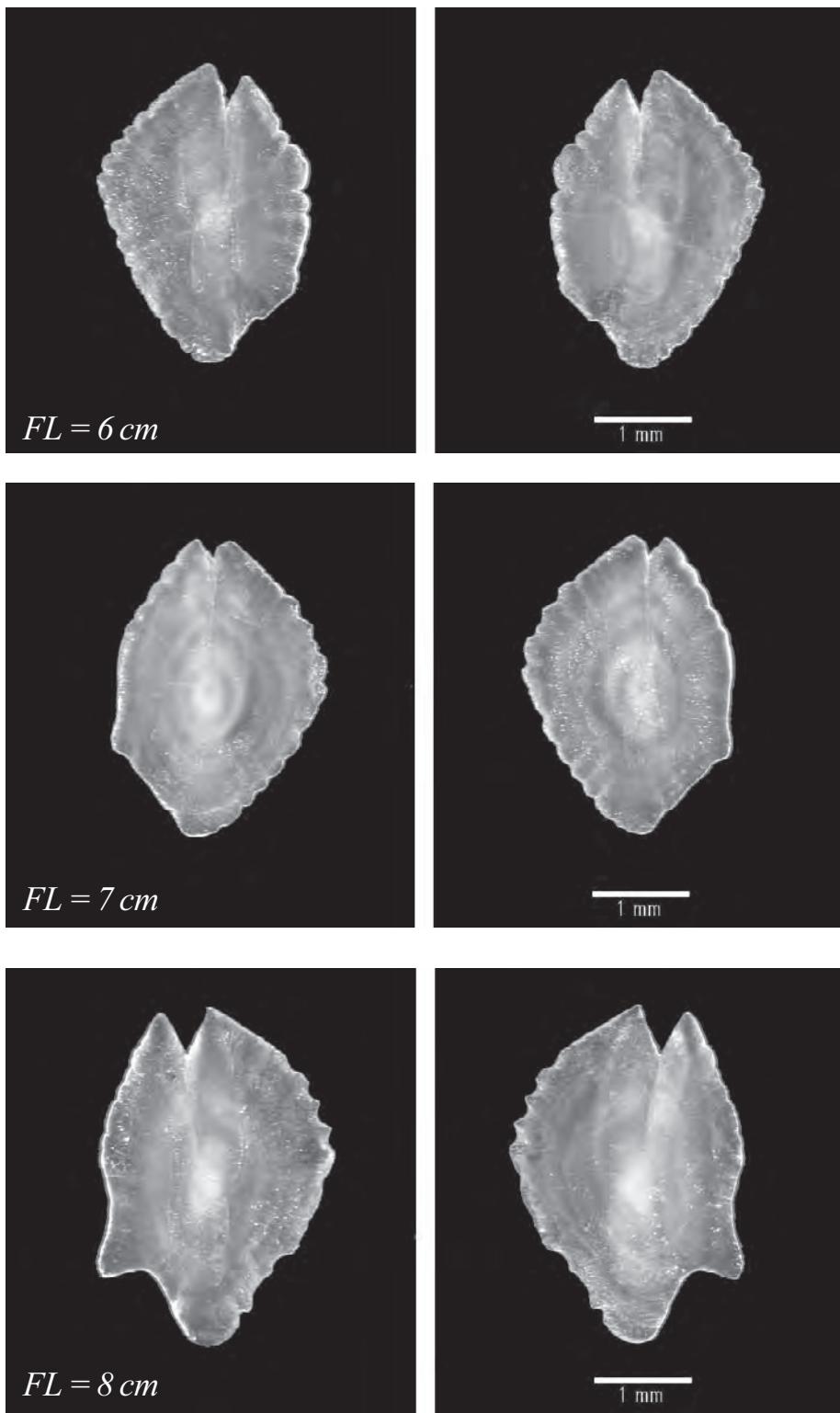
Myctophidae: *Lobianchia gemellarii*



Myctophiformes

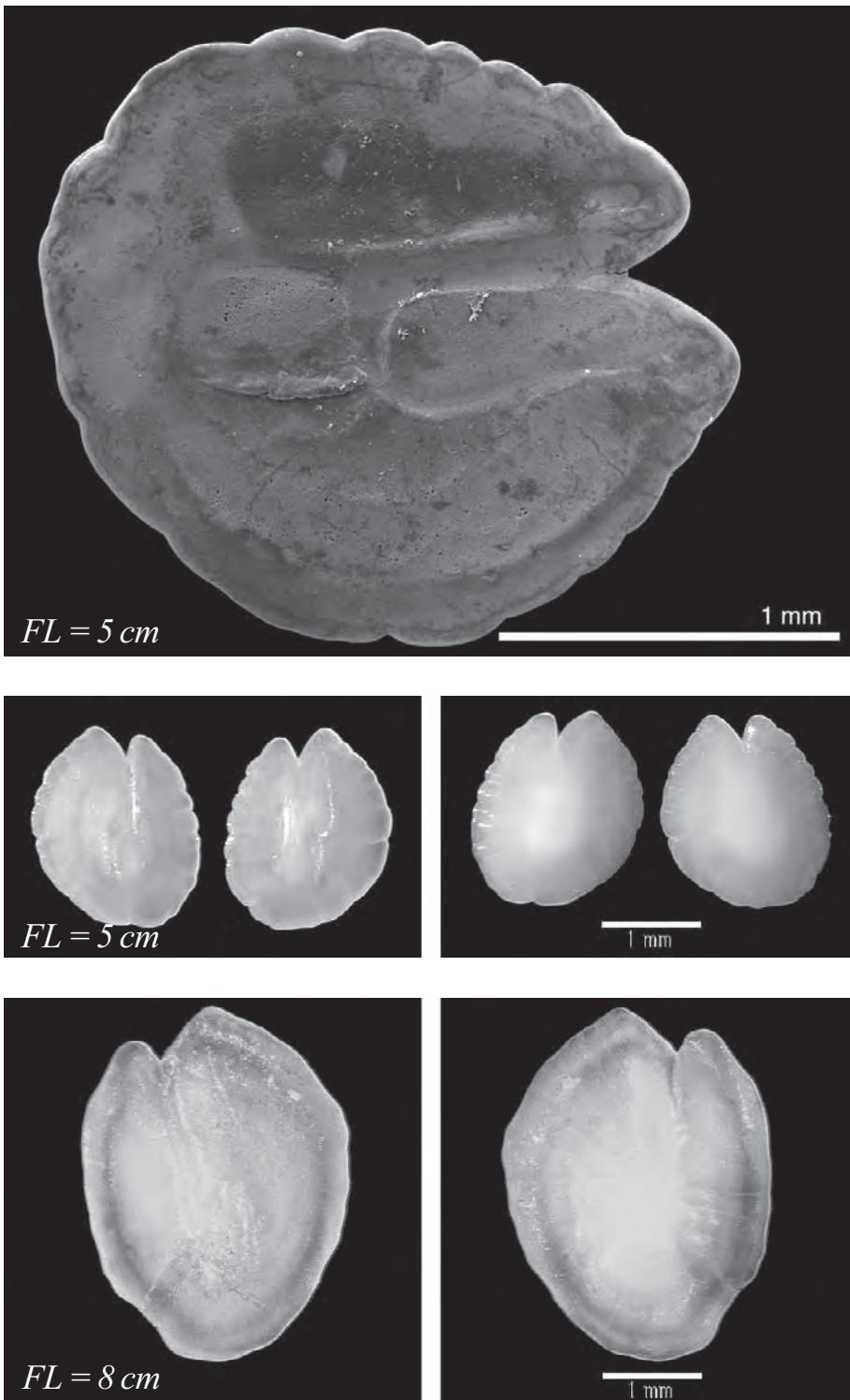
117

Myctophidae: *Myctophum asperum*



Myctophiformes

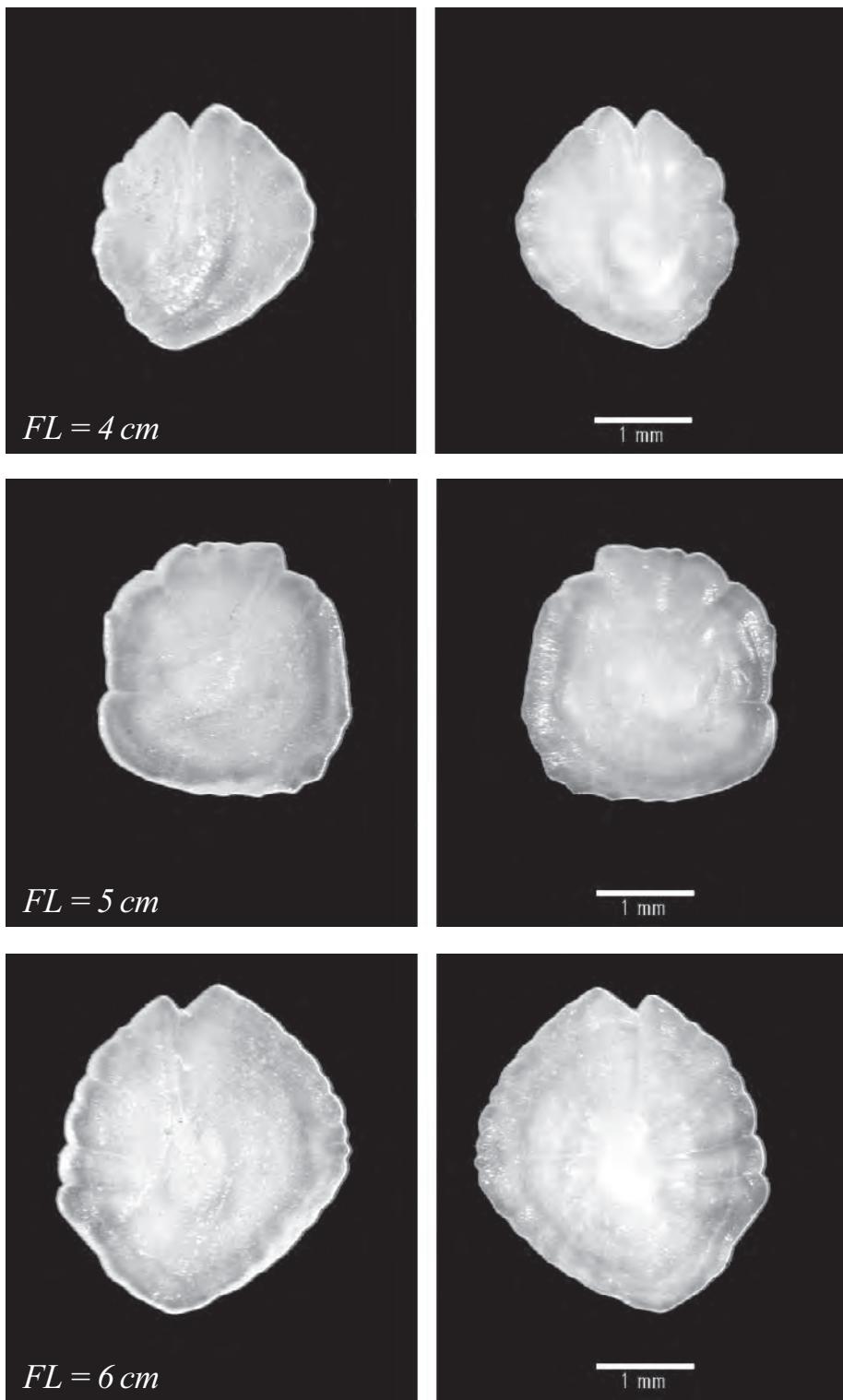
Myctophidae: *Myctophum punctatum*



Myctophiformes

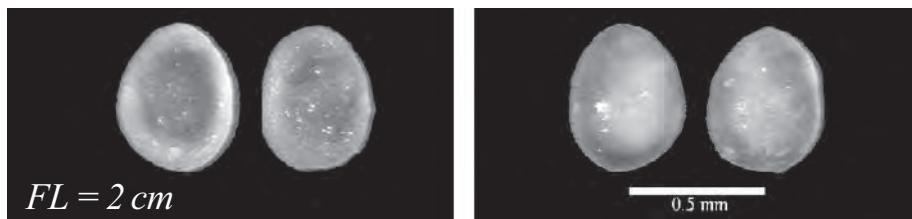
119

Myctophidae: *Myctophum selenops*

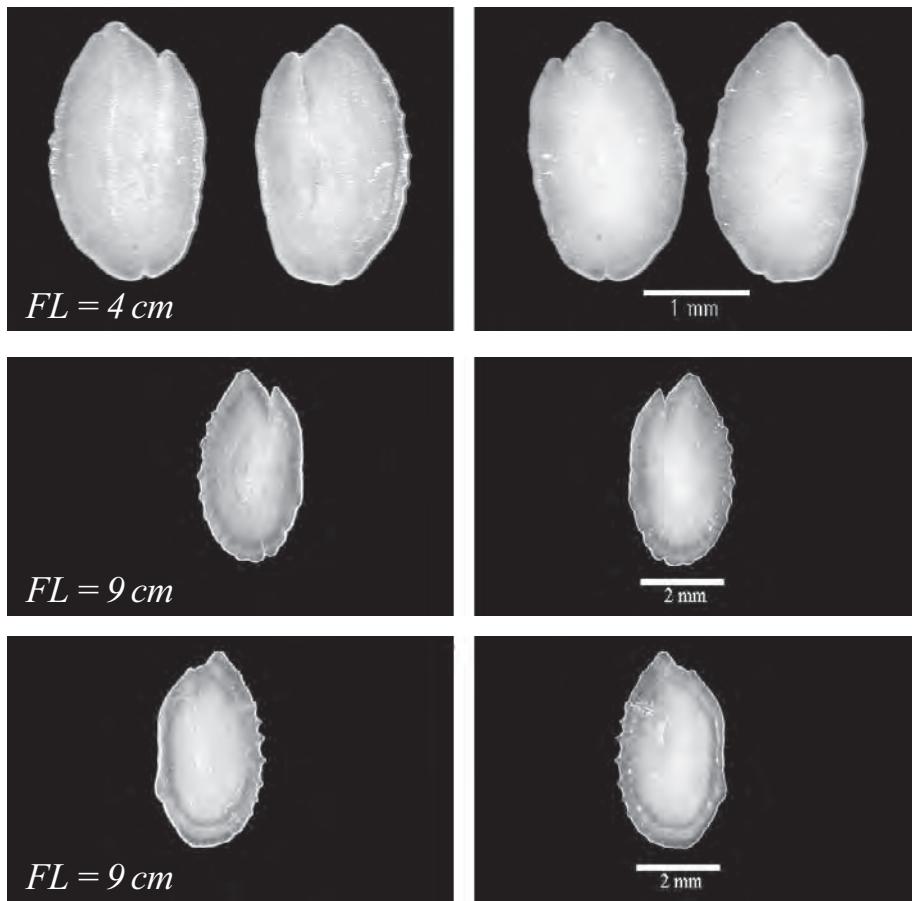


Myctophiformes

Myctophidae: *Notolychnus valdiviae*



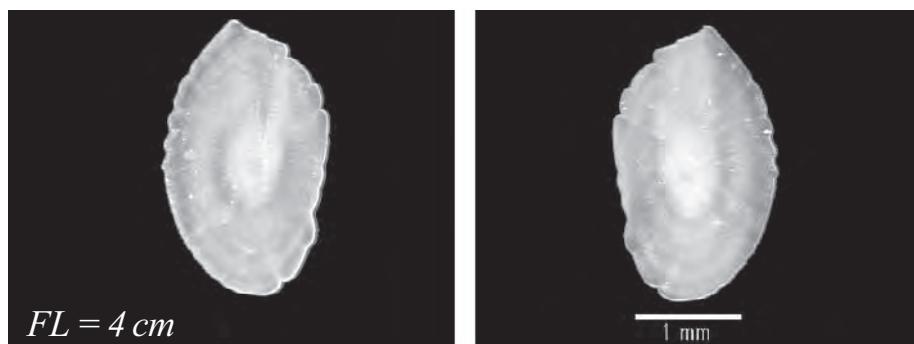
Myctophidae: *Notoscopelus bolini*



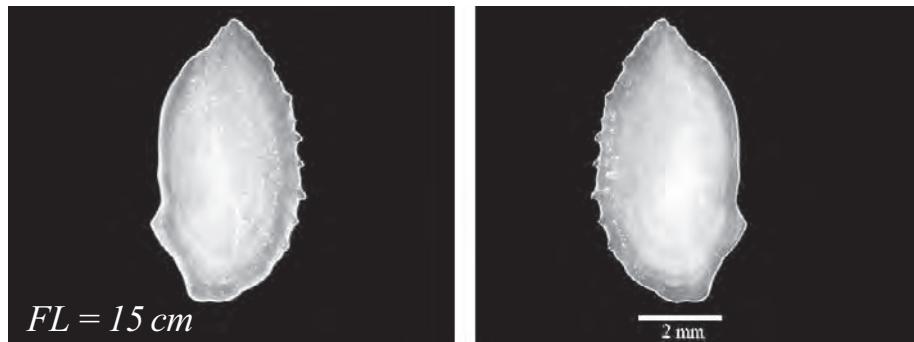
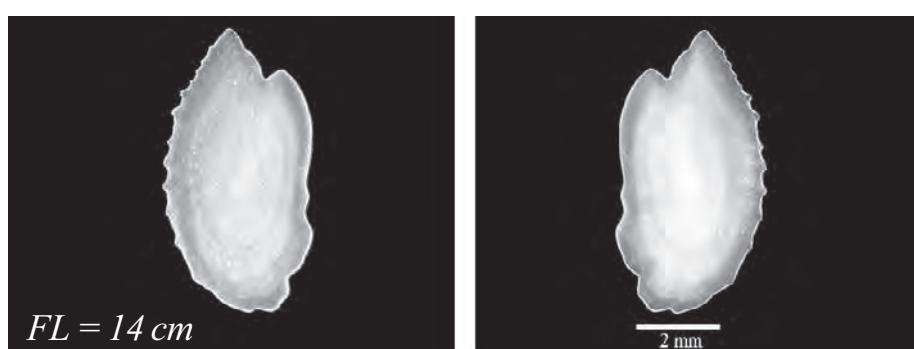
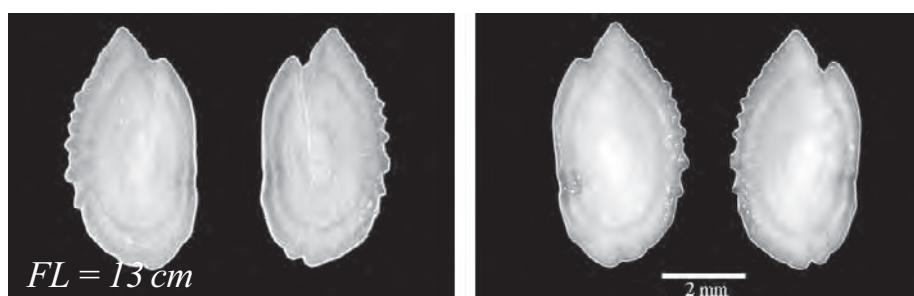
Myctophiformes

121

Myctophidae: *Notoscopelus caudispinosus*

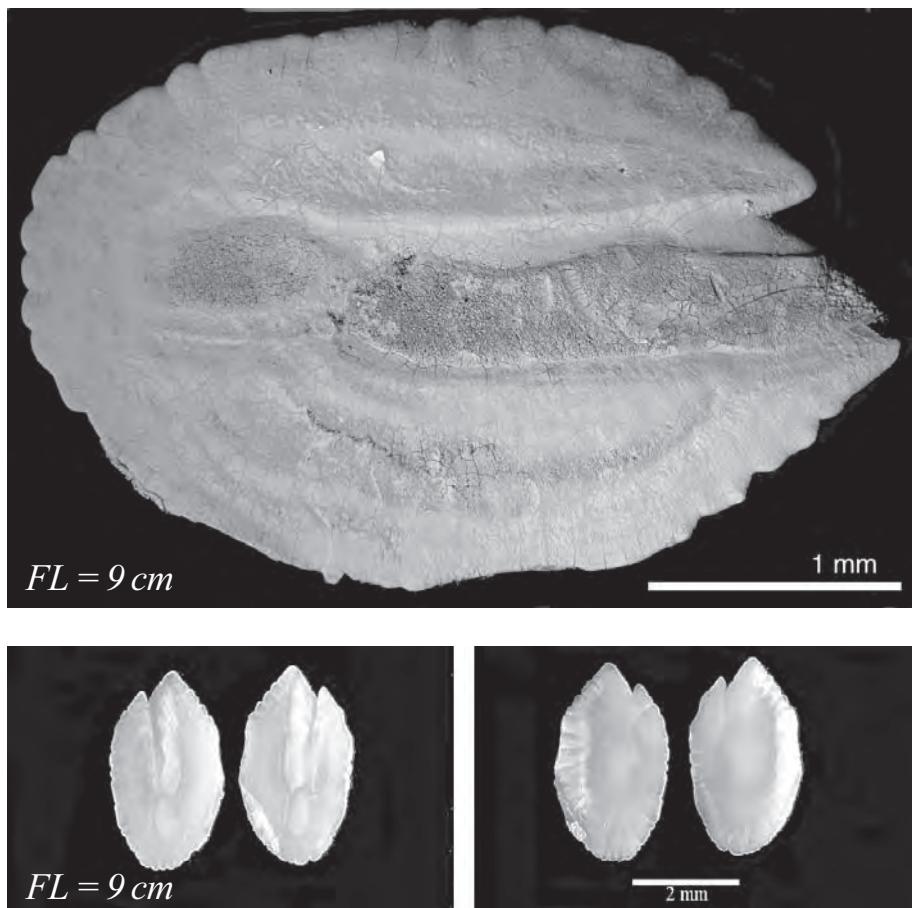


Myctophidae: *Notoscopelus elongatus kroeyerii*

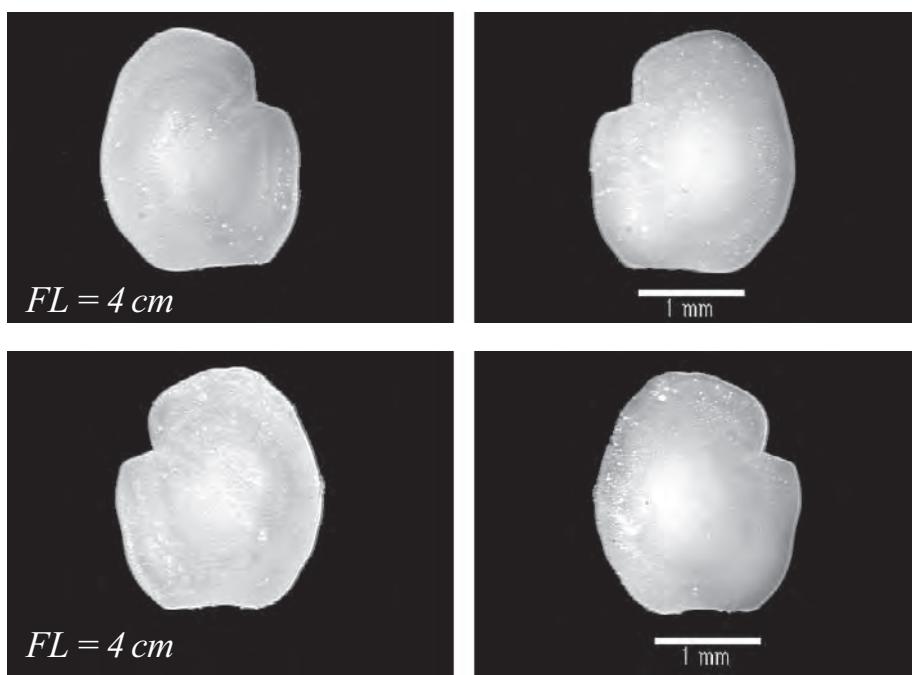


Myctophiformes

Myctophidae: *Notoscopelus resplendens*



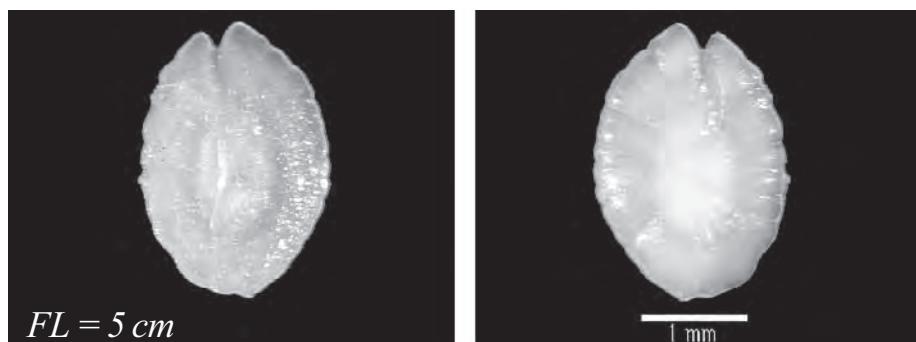
Myctophidae: *Protomyctophum arcticum*



Myctophiformes

123

Myctophidae: *Symbolophorus veranyi*



Myctophiformes

Myctophidae: *Taaningichthys bathyphilus*



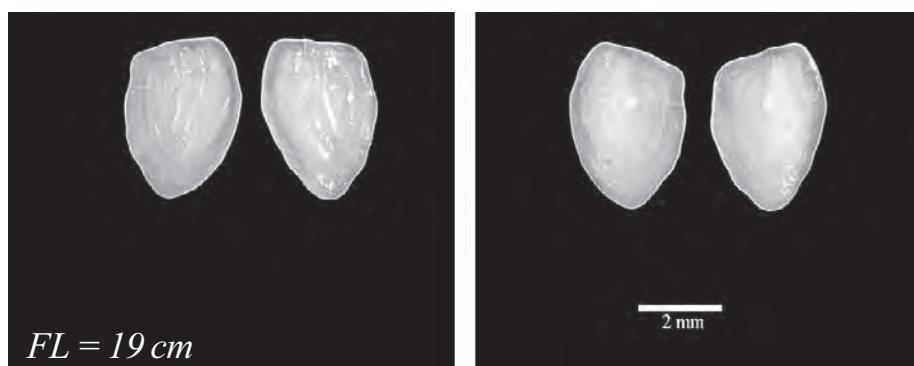
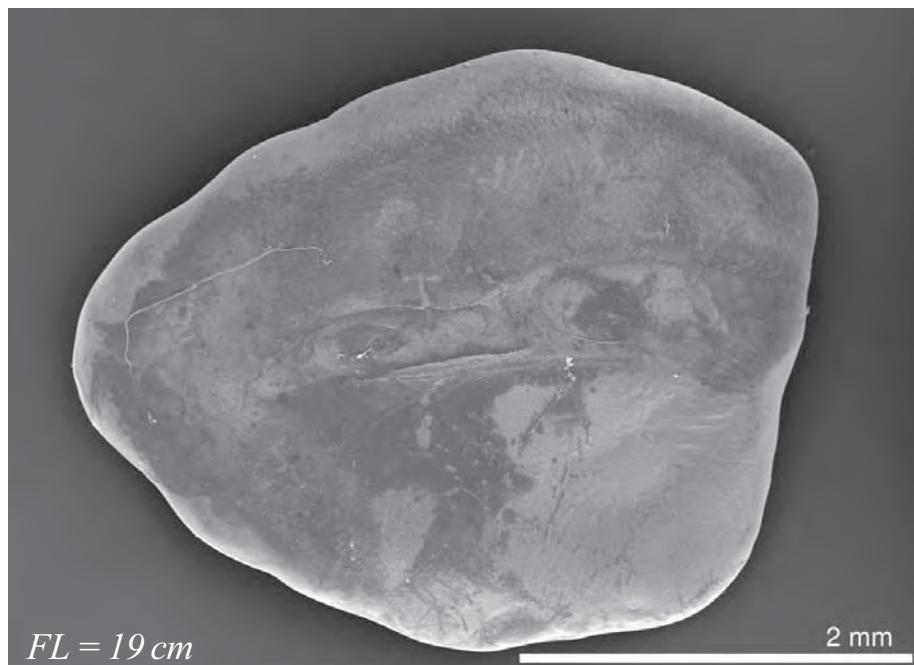
Myctophidae: *Taaningichthys minimus*



Ophidiiformes

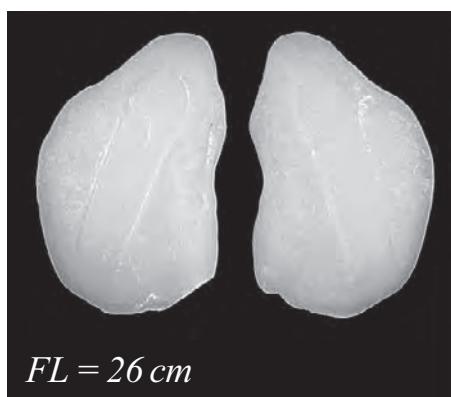
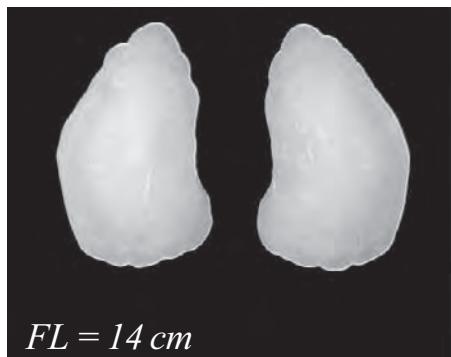
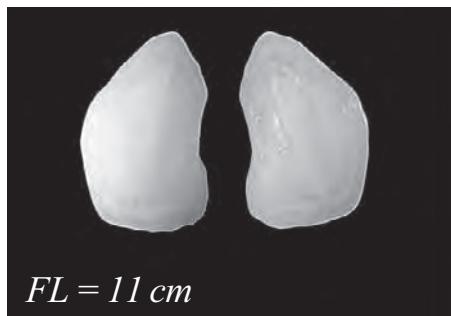
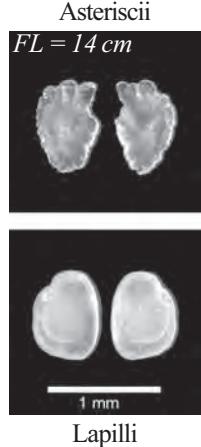
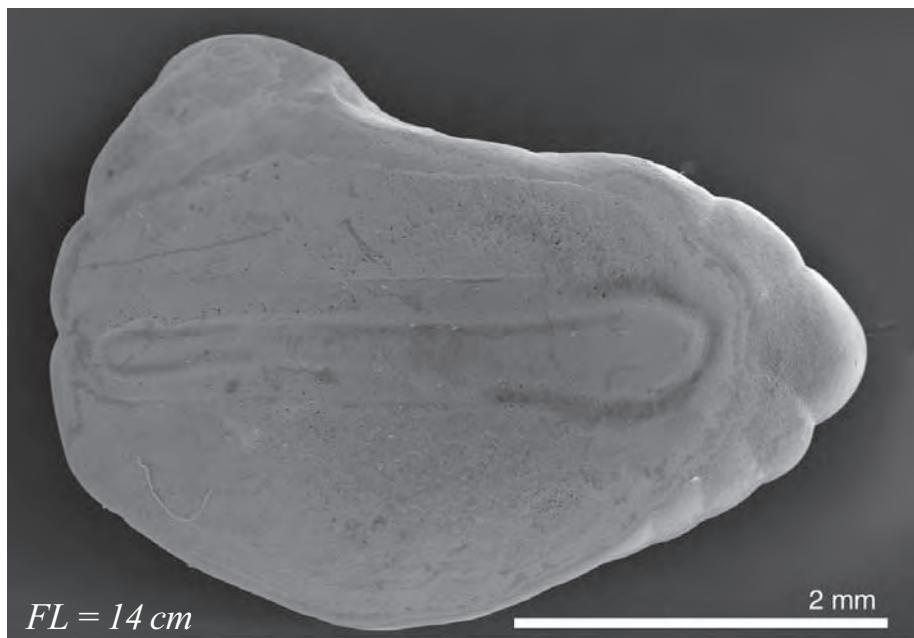
Ophidiidae: *Dicrolene intronigra*

125



Ophidiiformes

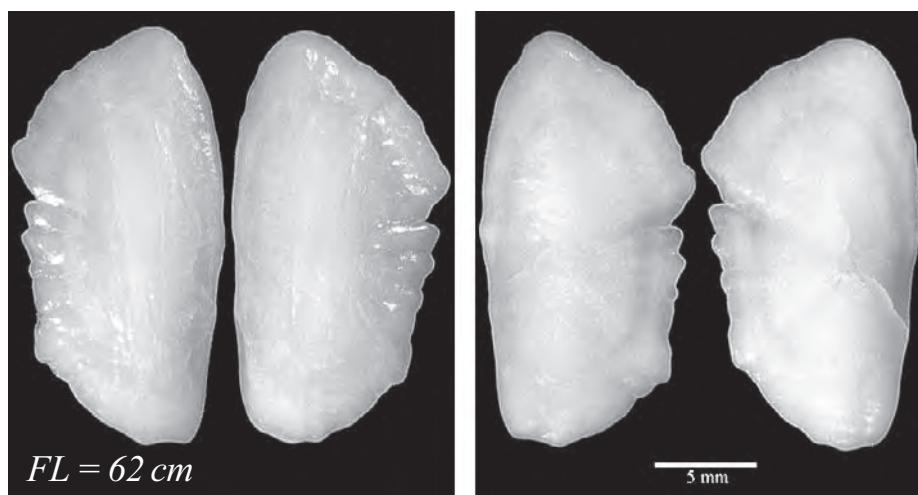
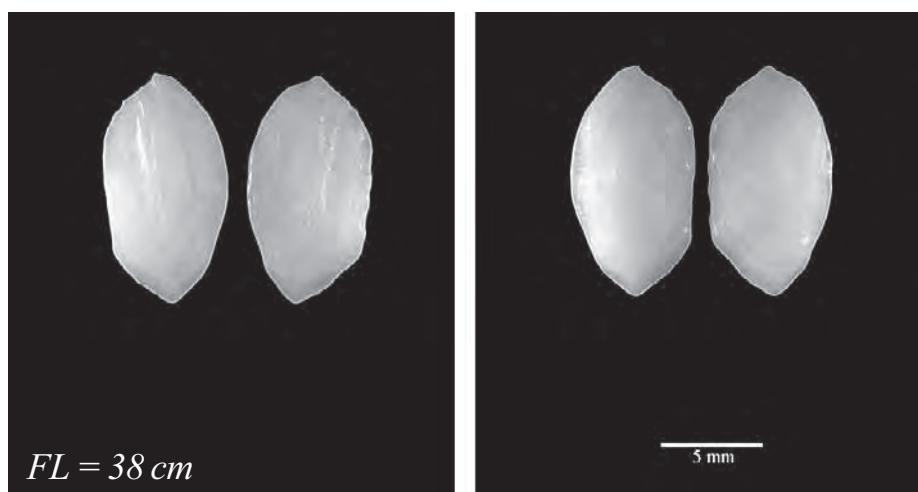
Ophidiidae: *Lepophidium cervinum*



Ophidiiformes

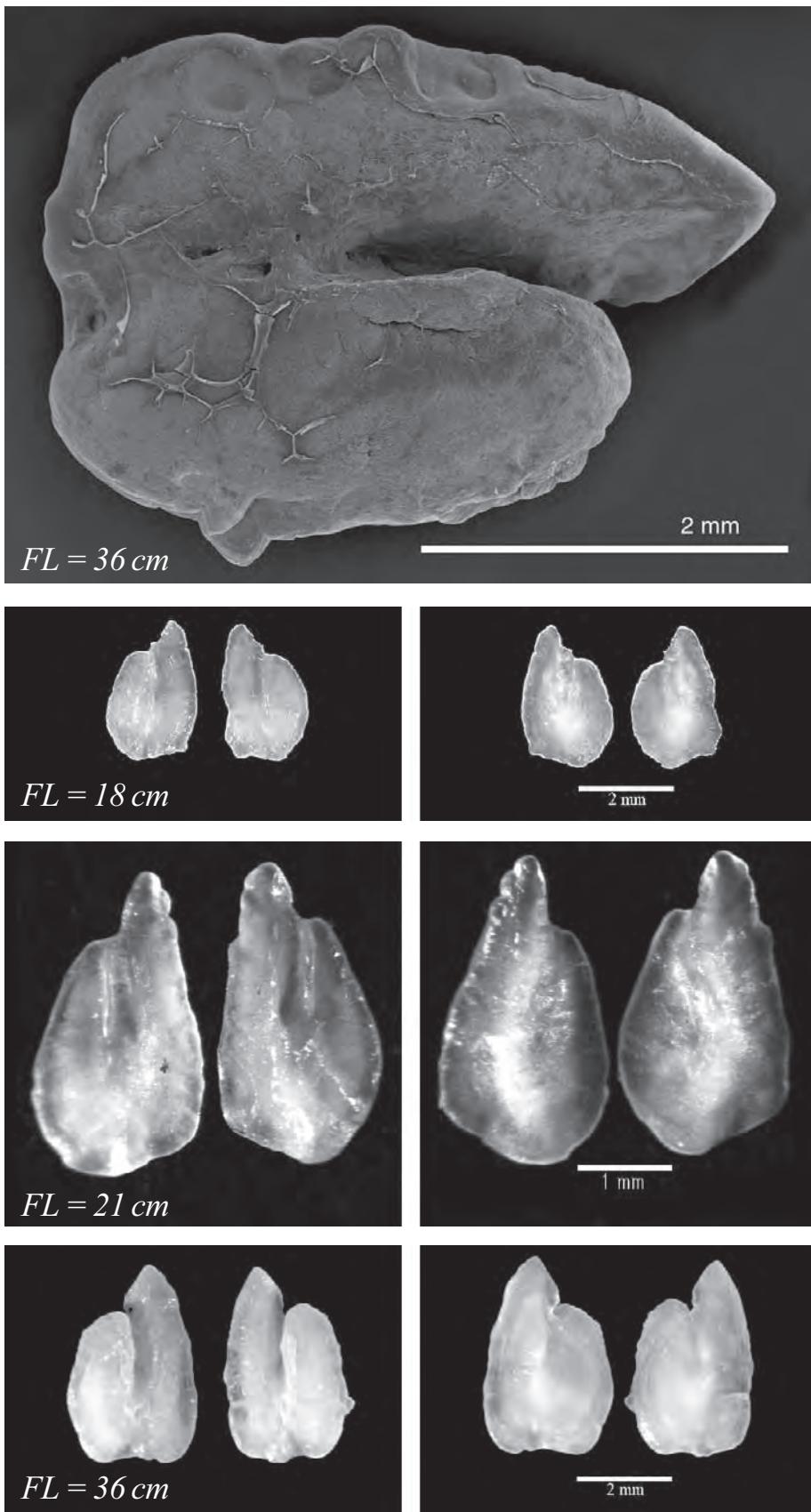
127

Ophidiidae: *Spectrunculus grandis*



Osmeriformes

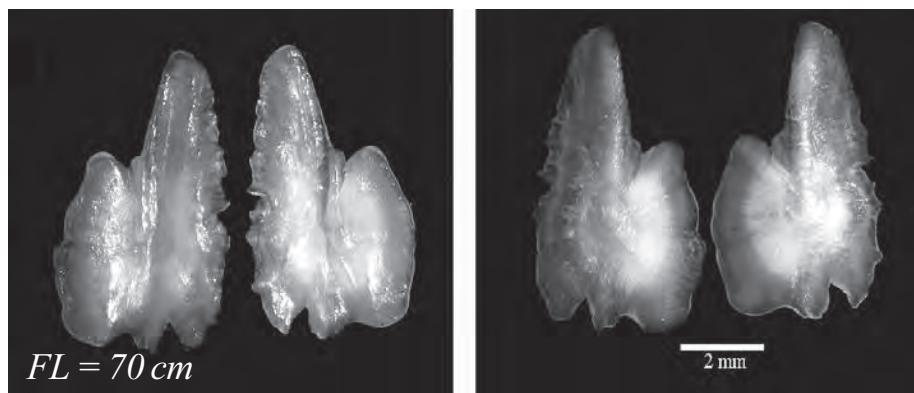
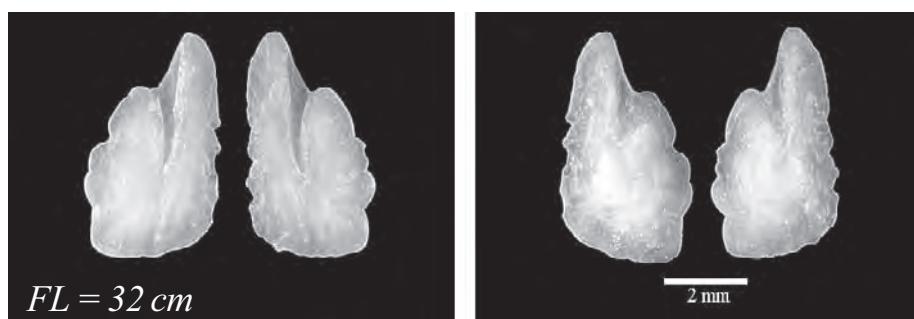
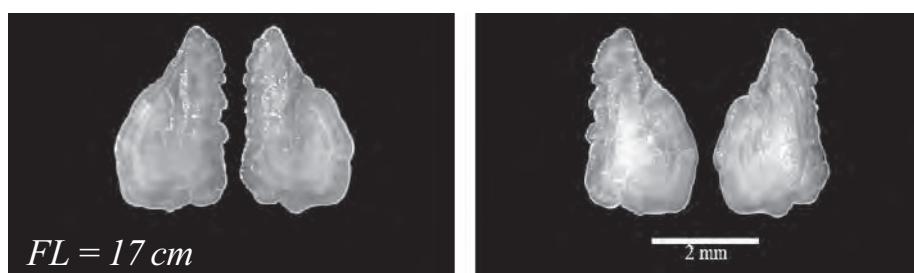
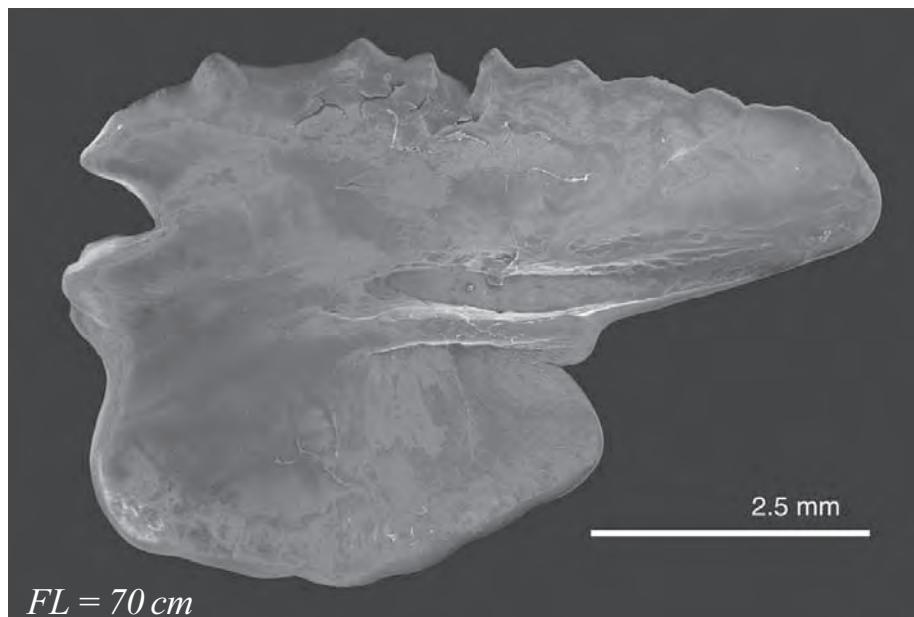
Alepocephalidae: *Alepocephalus agassizii*



Osmeriformes

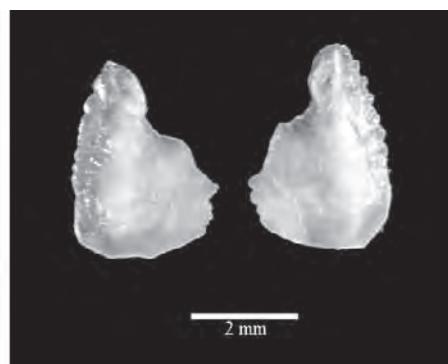
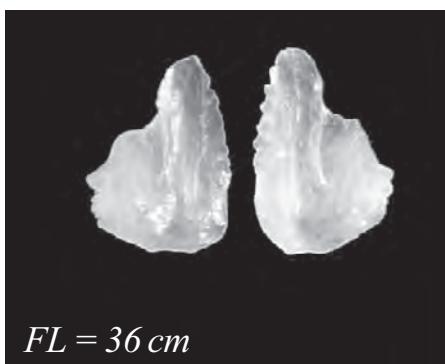
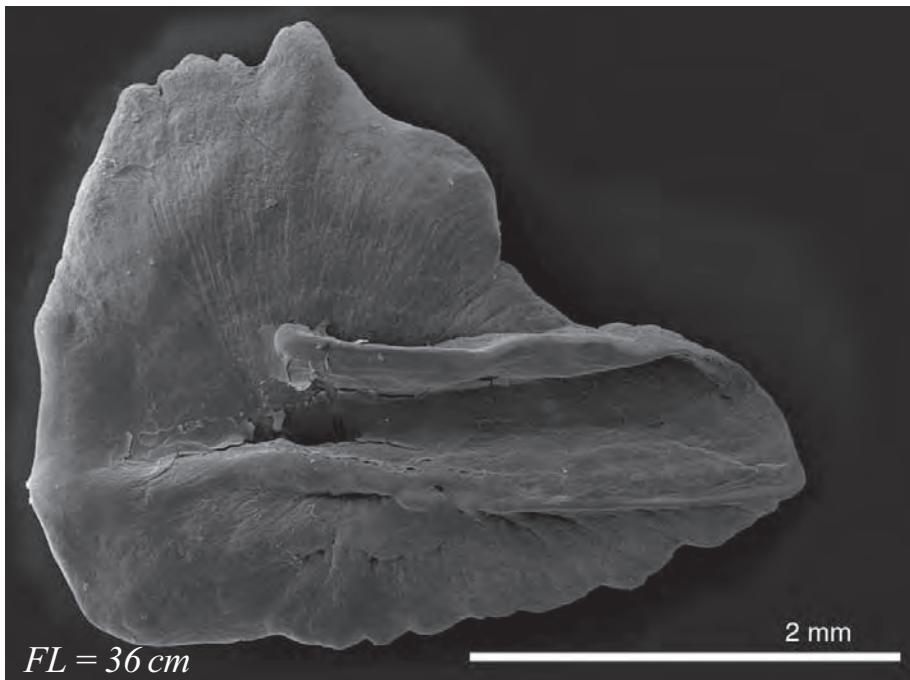
129

Alepocephalidae: *Alepocephalus bairdii*



Osmeriformes

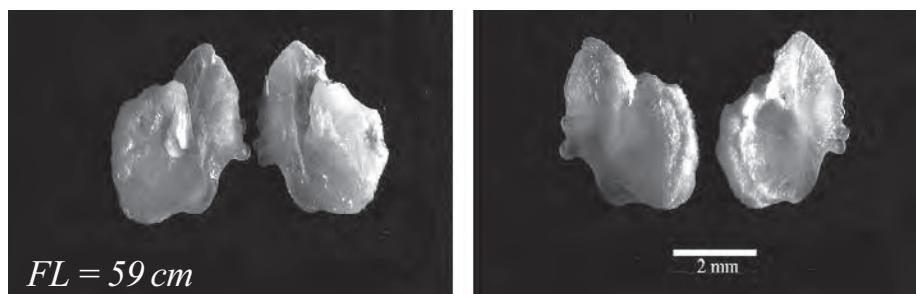
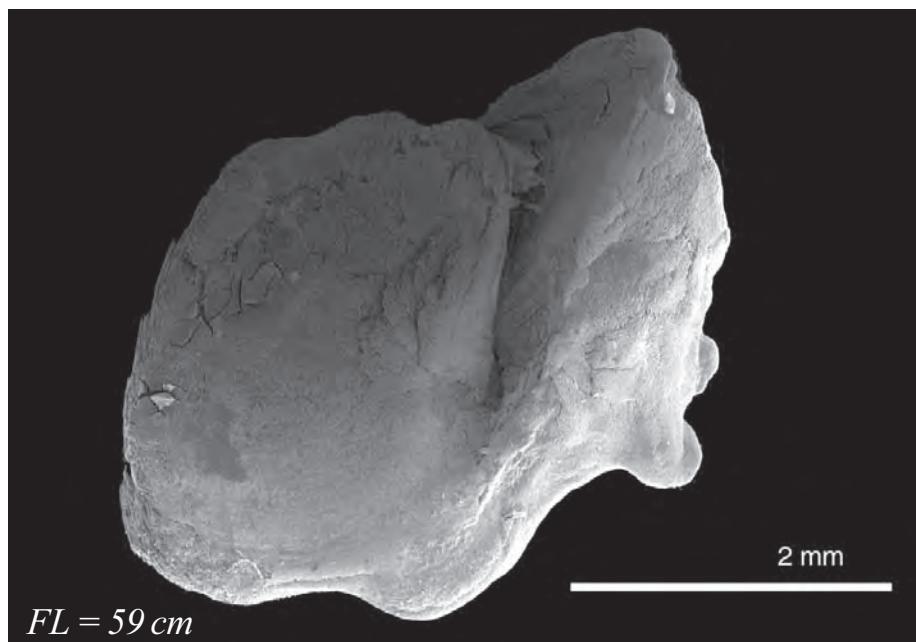
Alepocephalidae: *Bajacalifornia megalops*



Osmeriformes

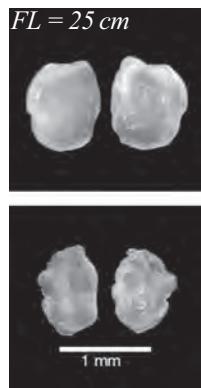
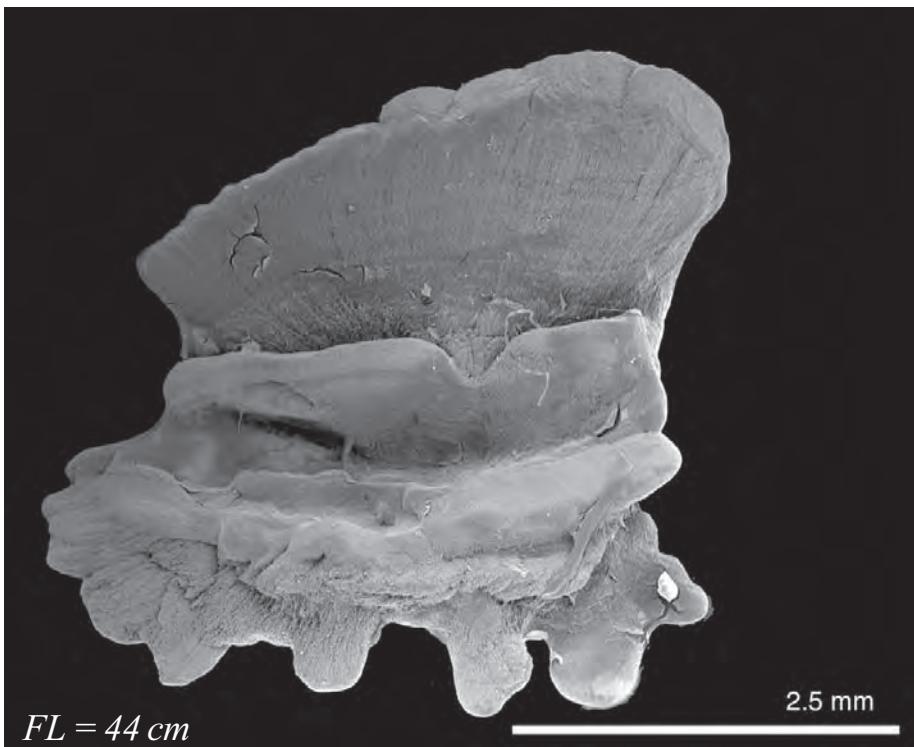
131

Alepocephalidae: *Narcetes stomias*

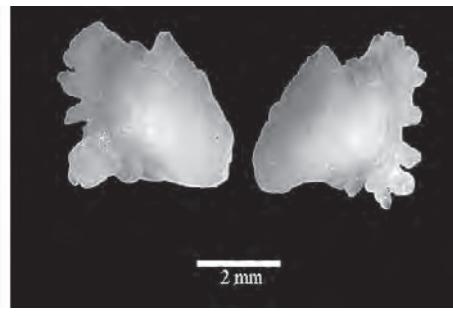
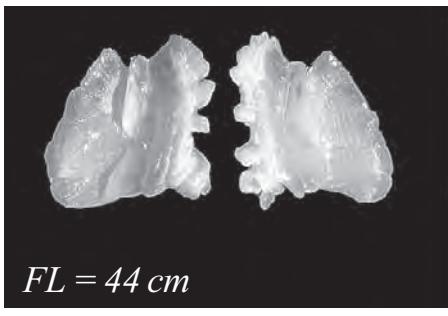
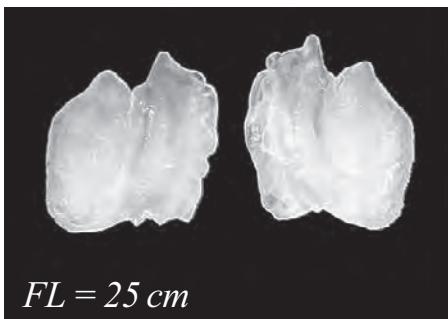


Osmeriformes

Alepocephalidae: *Rouleina attrita*



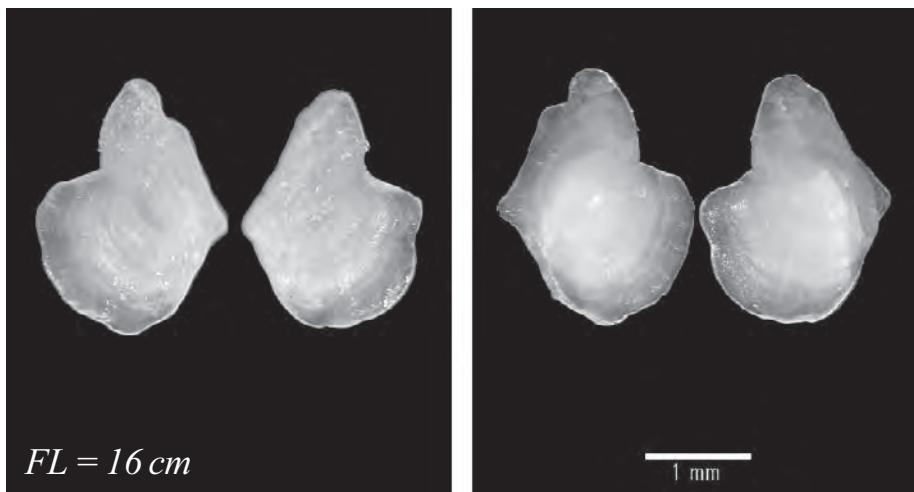
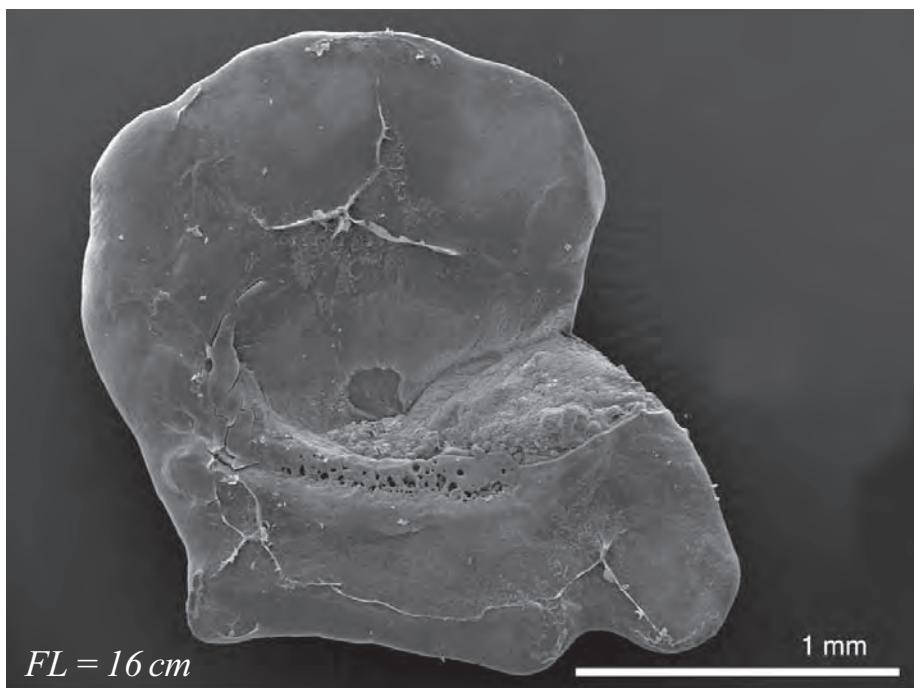
Asteriscii
& Lapilli



Osmeriformes

133

Alepocephalidae: *Xenodermichthys copei*



Osmeriformes

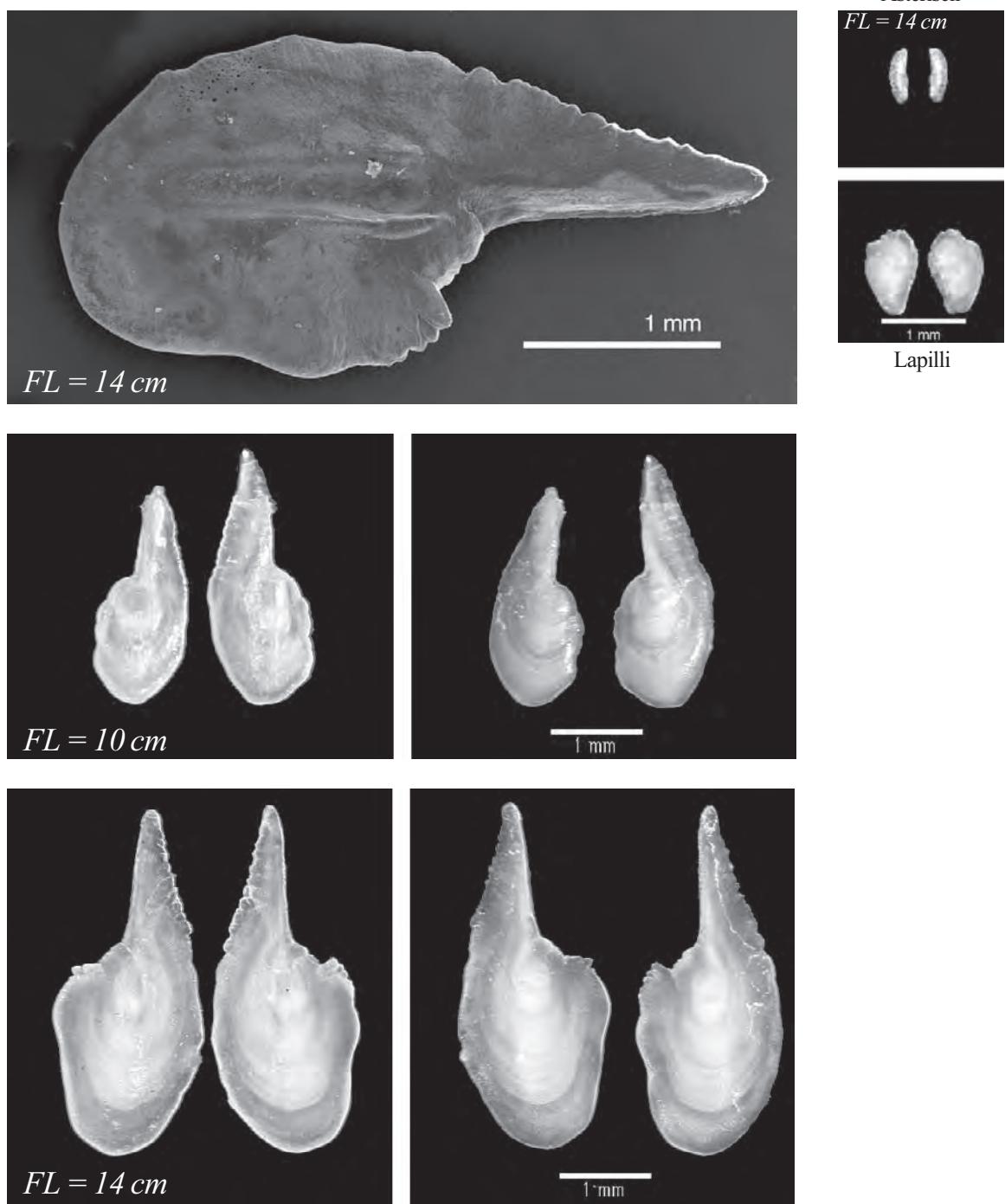
Argentinidae: *Argentina silus*



Osmeriformes

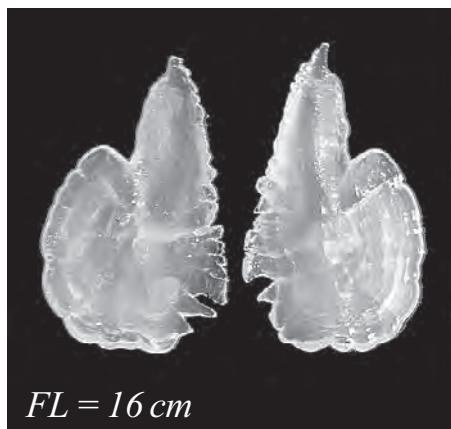
135

Bathylagidae: *Bathylagus euryops*



Osmeriformes

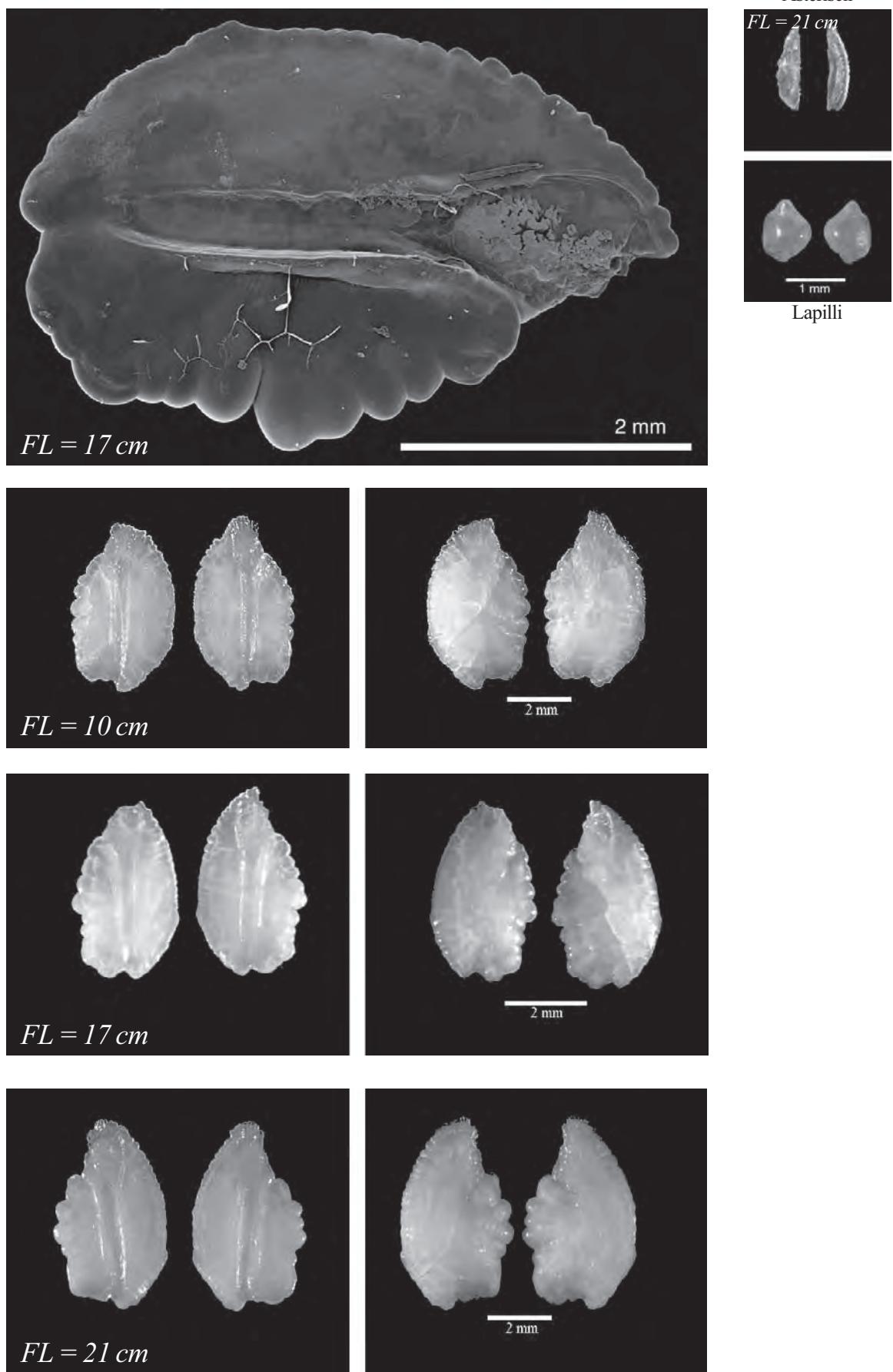
Osmeridae: *Mallotus villosus*



Osmeriformes

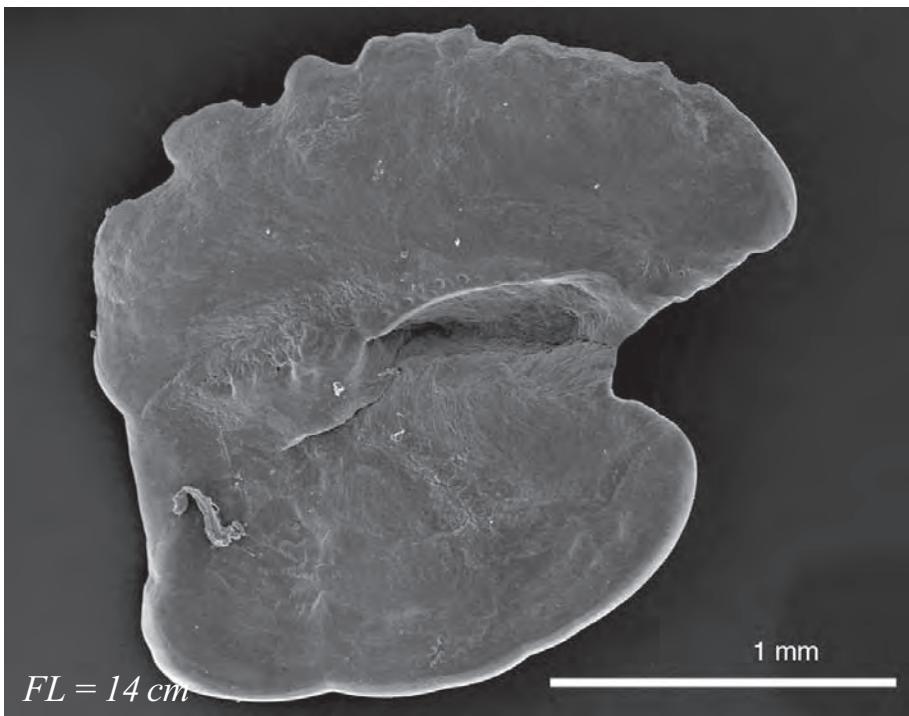
Osmeridae: *Osmerus mordax*

137



Osmeriformes

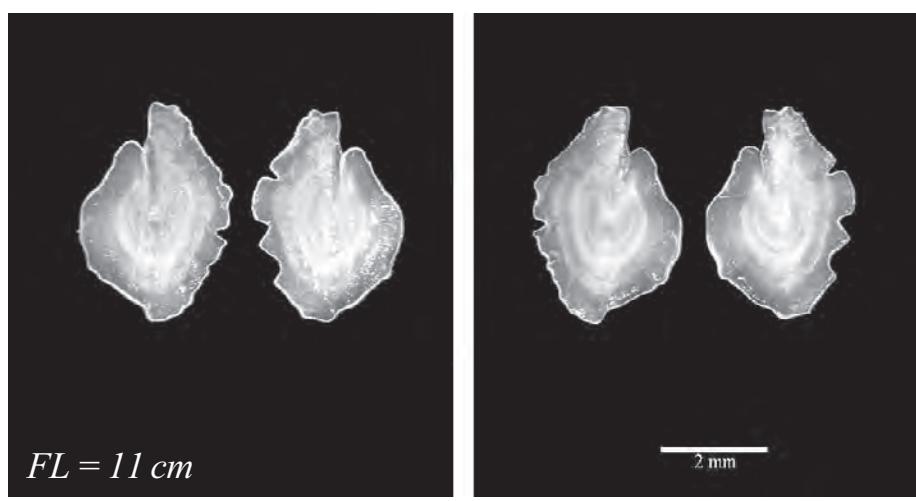
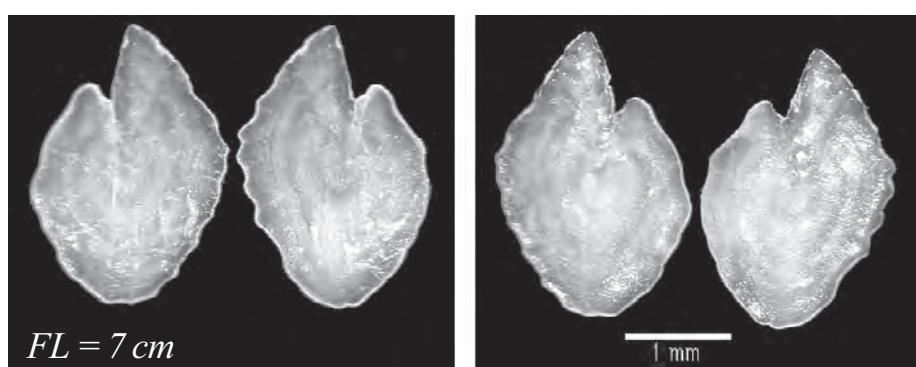
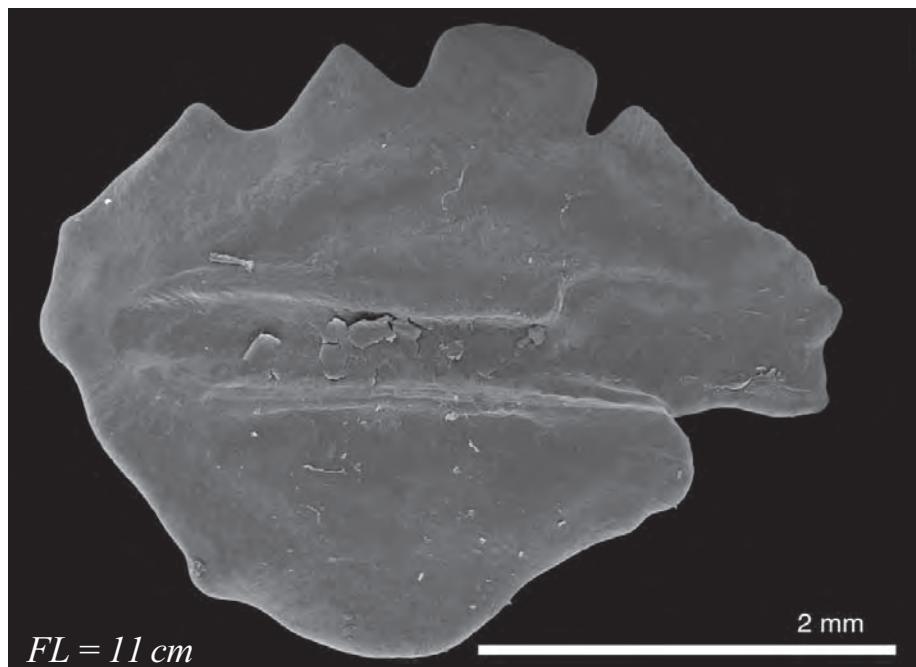
Platytroctidae: *Holtbyrnia macrops*



Perciformes

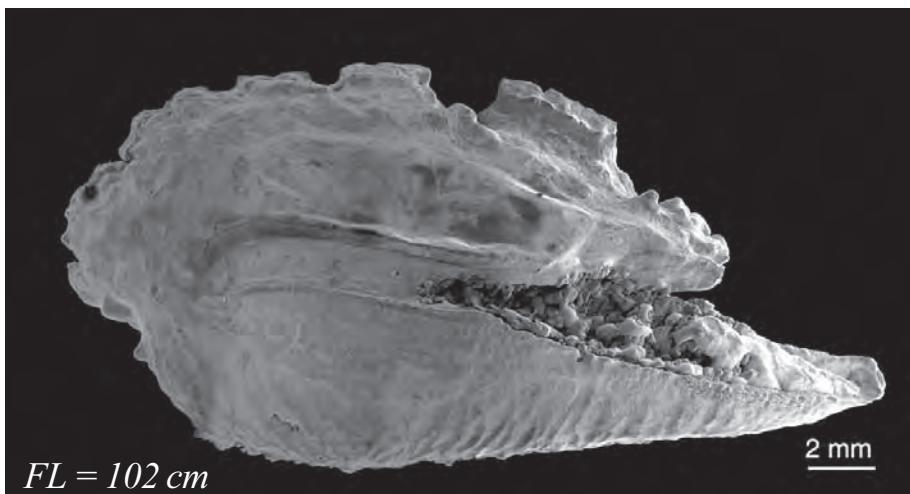
139

Acropomatidae: *Howella sherborni*



Perciformes

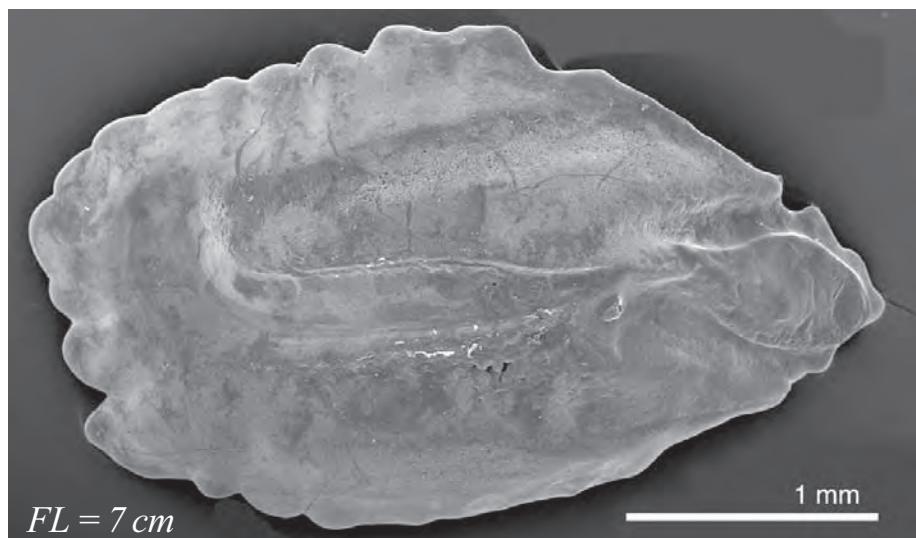
Acropomatidae: *Polyprion americanus*



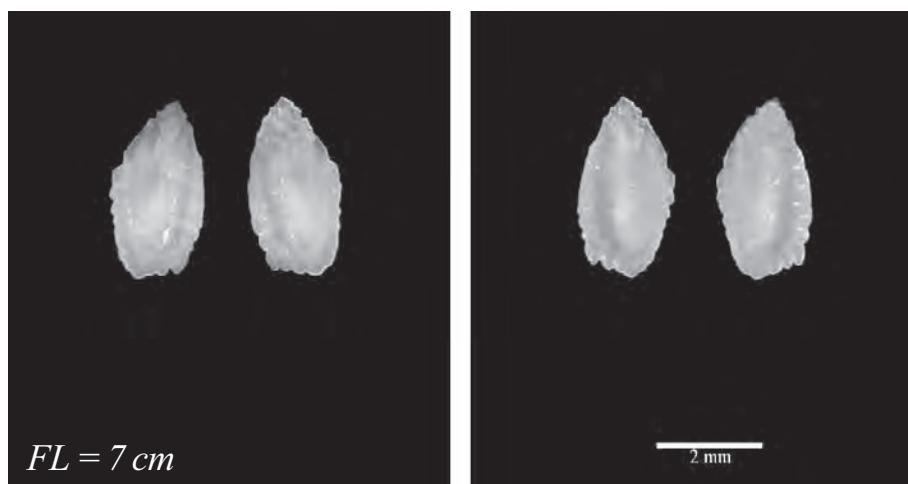
Perciformes

141

Acropomatidae: *Synagrops bellus*



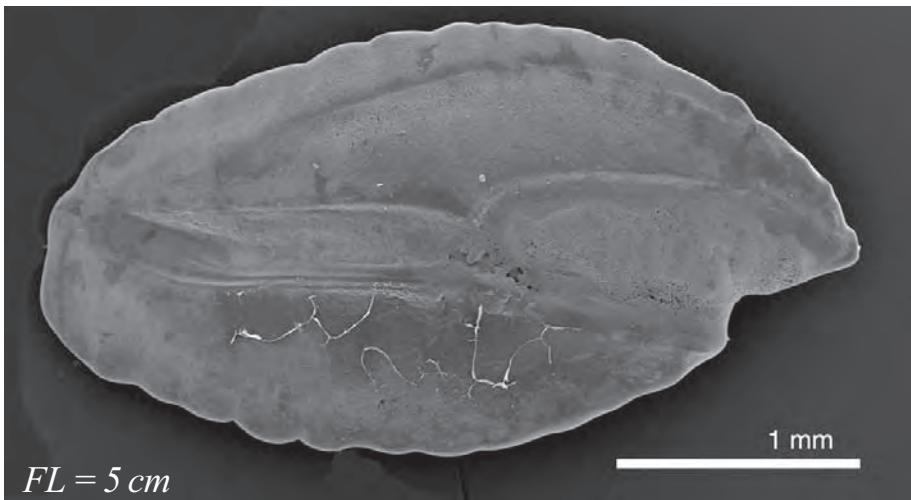
FL = 7 cm



FL = 7 cm

Perciformes

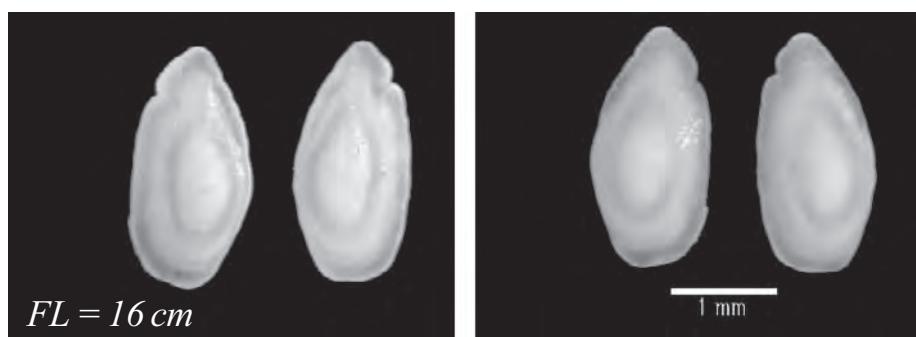
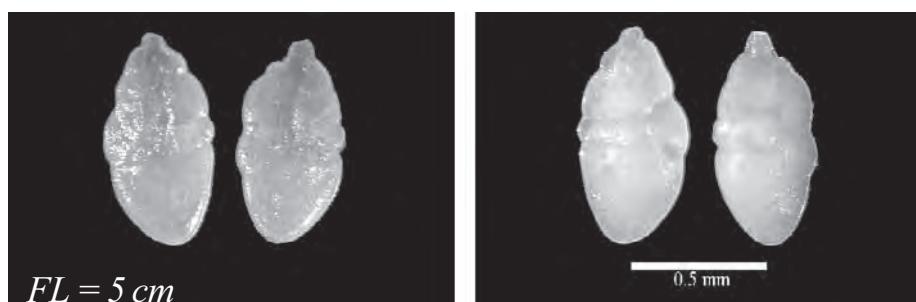
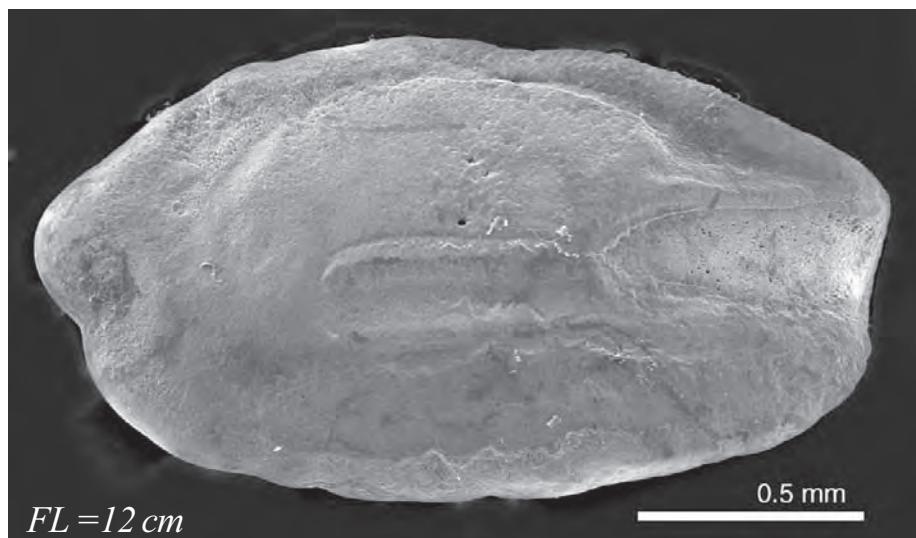
Acropomatidae: *Synagrops spinosa*



Perciformes

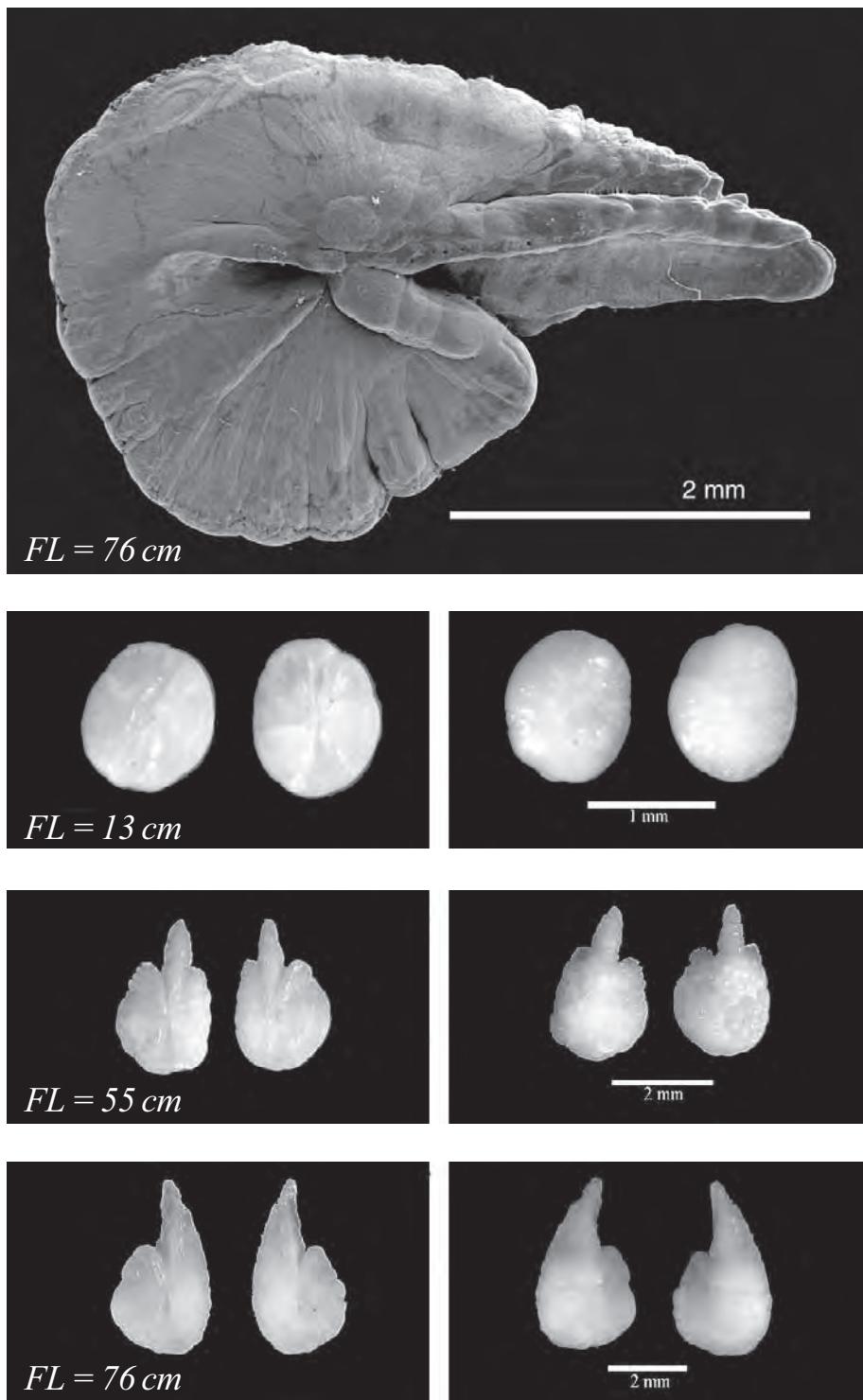
143

Ammodytidae: *Ammodytes americanus / dubius*

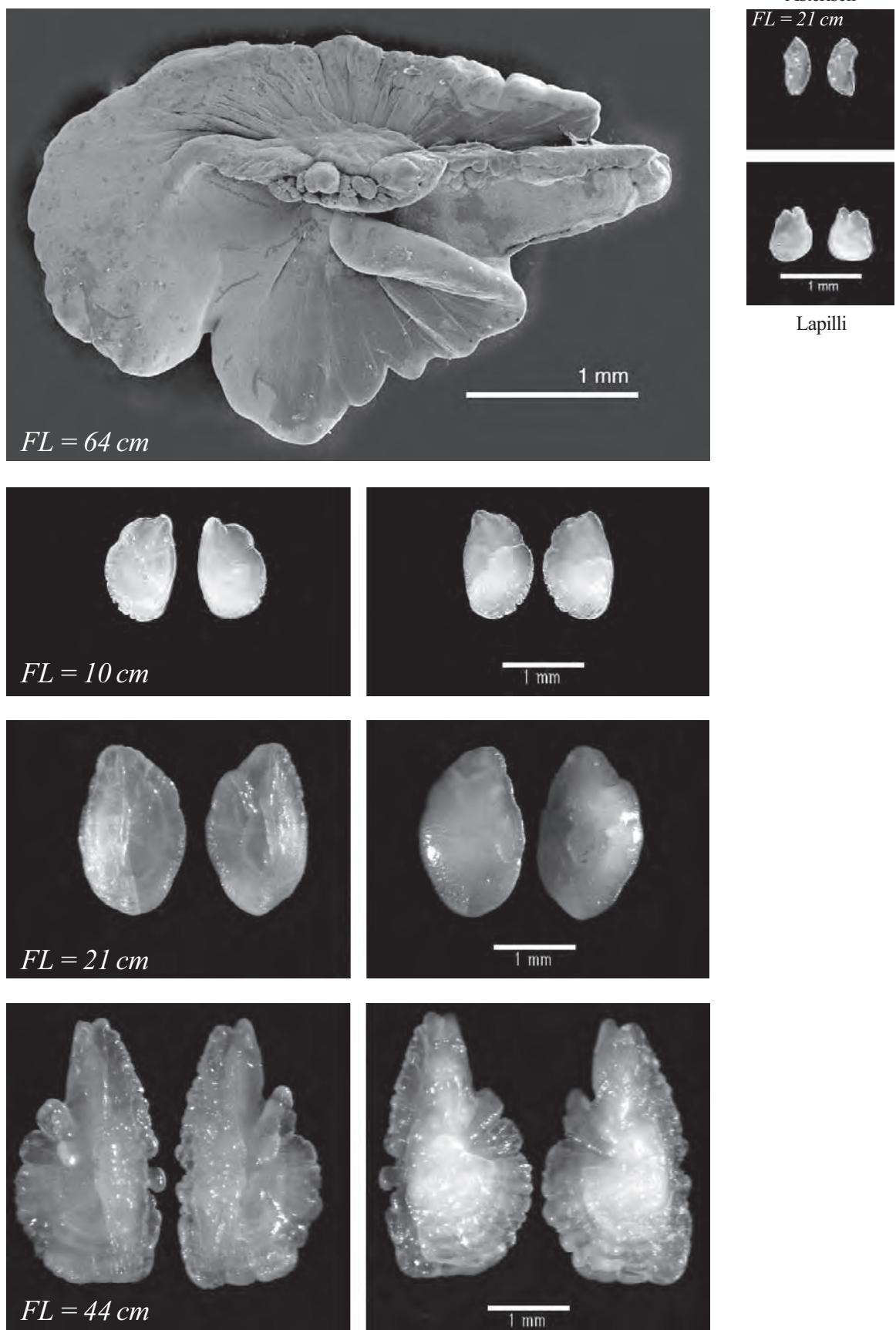


Perciformes

Anarhichadidae: *Anarhichas denticulatus*

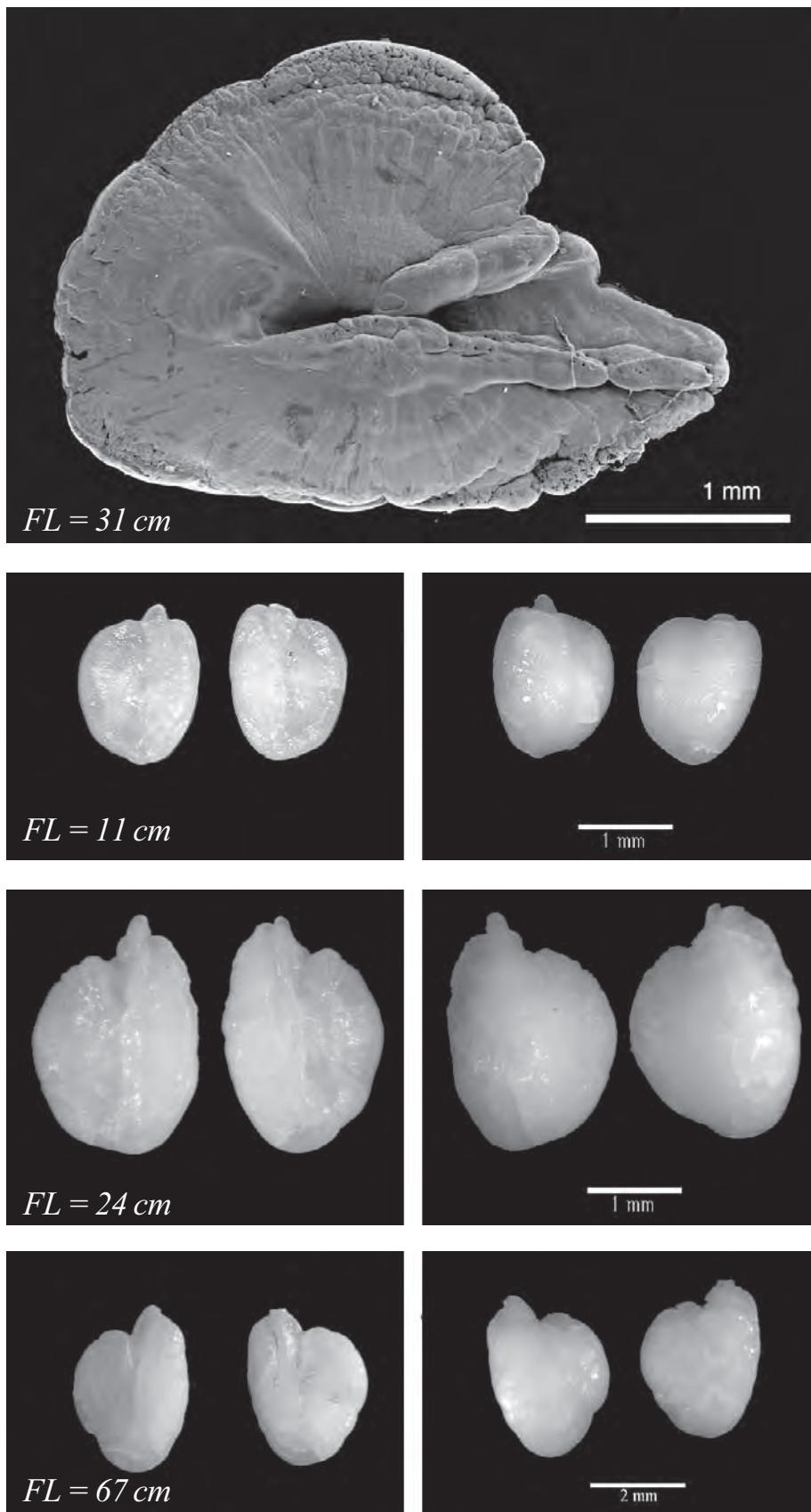


Anarhichadidae: *Anarhichas lupus*



Perciformes

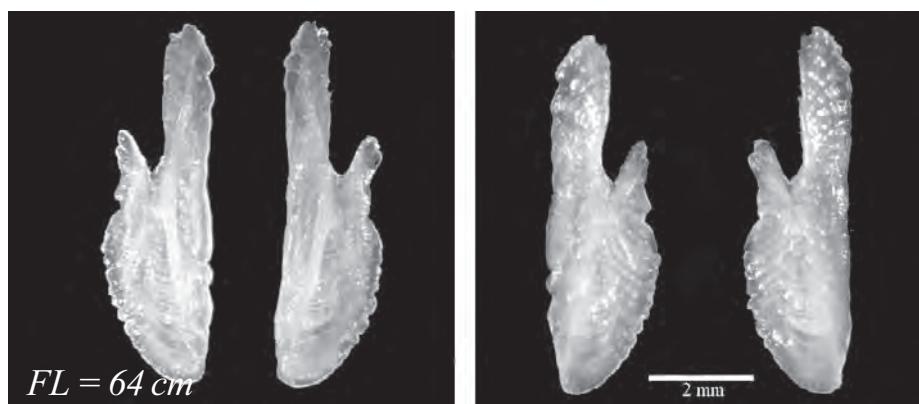
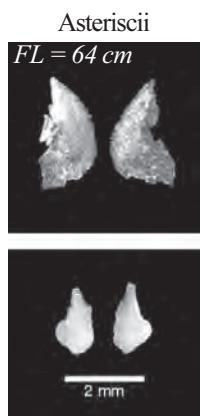
Anarhichadidae: *Anarhichas minor*



Perciformes

147

Bramidae: *Brama brama*

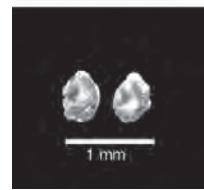
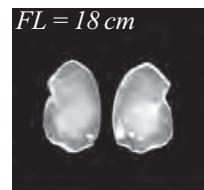


Bramidae: *Taractichthys longipinnis*



Perciformes

Callionymidae: *Callionymus agassizi*



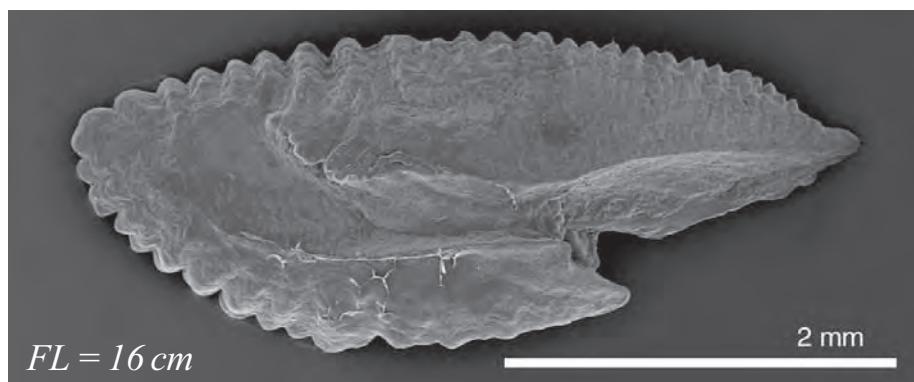
Asterisci
& Lapilli



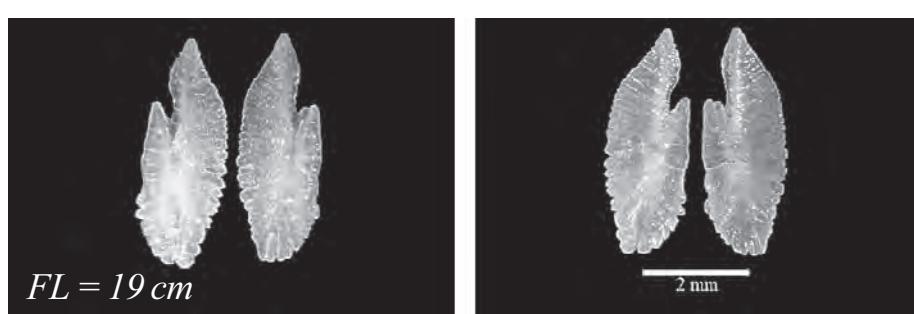
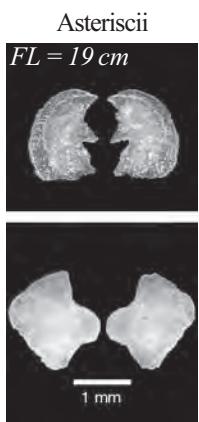
Perciformes

149

Carangidae: *Caranx crysos*

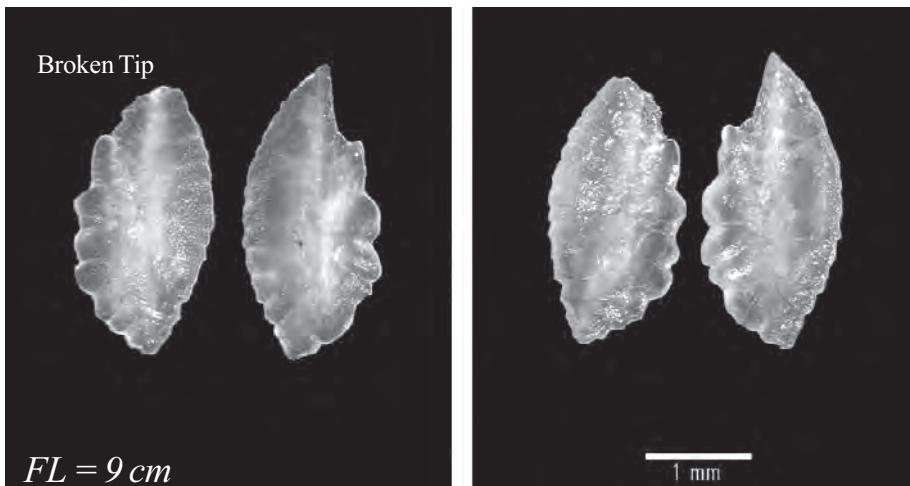
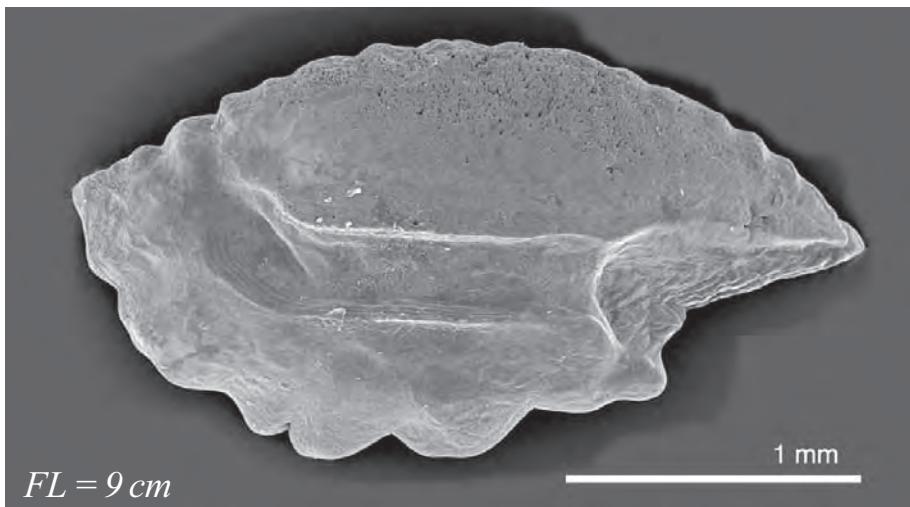


Carangidae: *Caranx hippos*



Perciformes

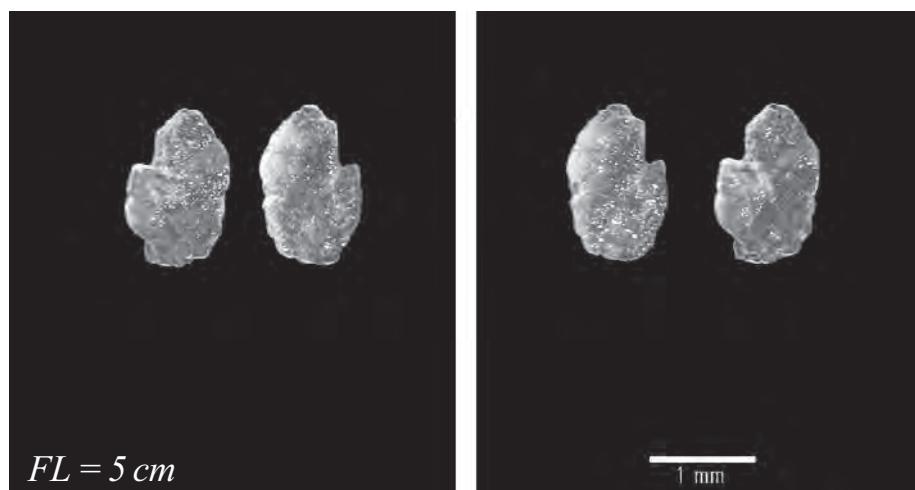
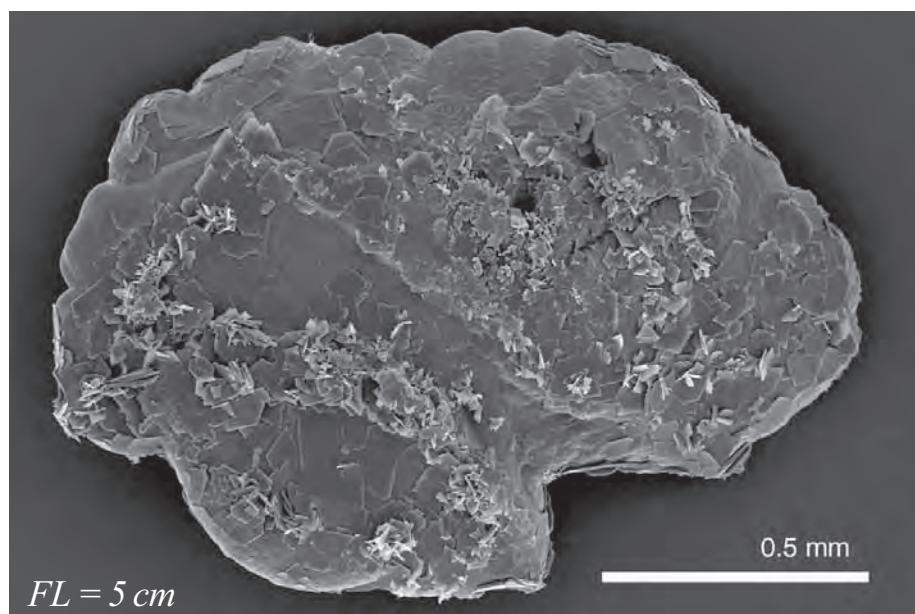
Carangidae: *Decapterus macarellus*



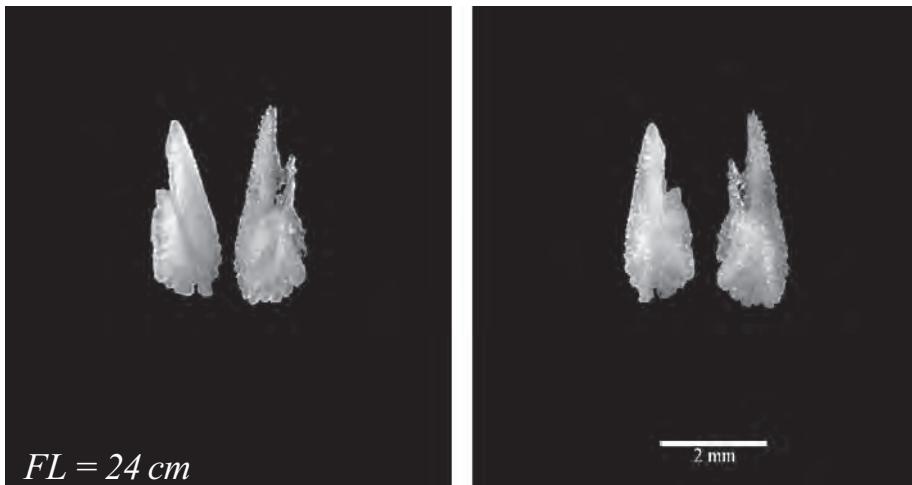
Perciformes

151

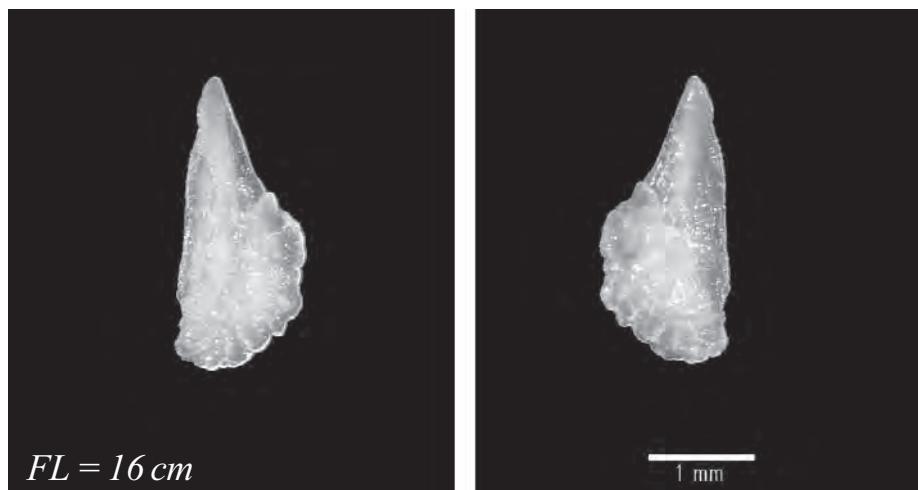
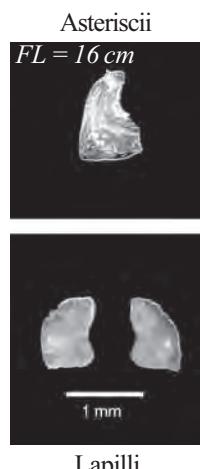
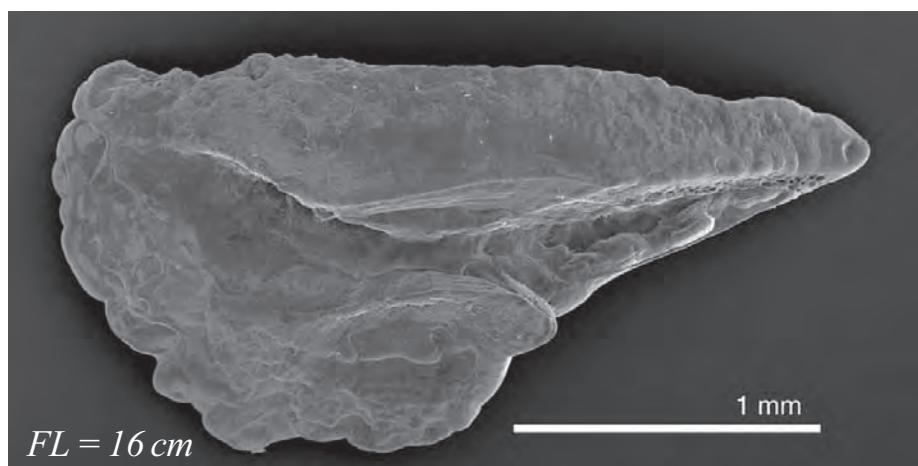
Carangidae: *Selene setapinnis*



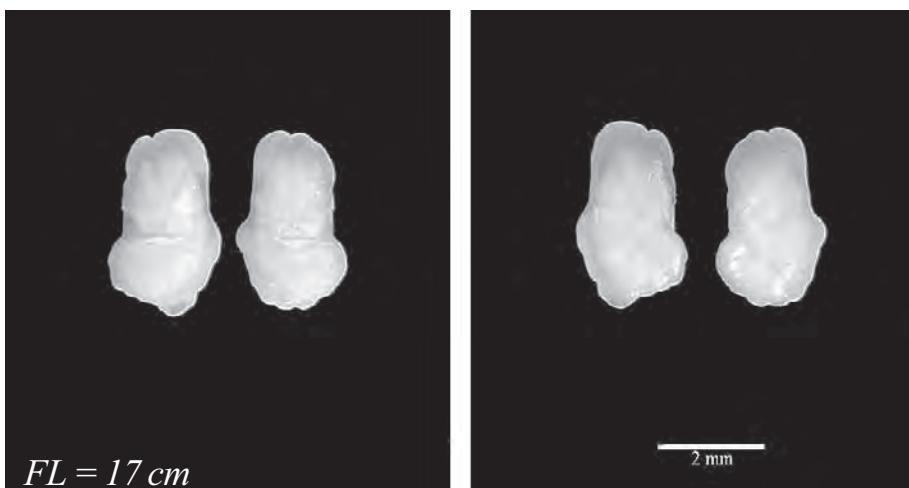
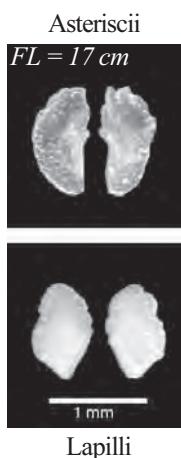
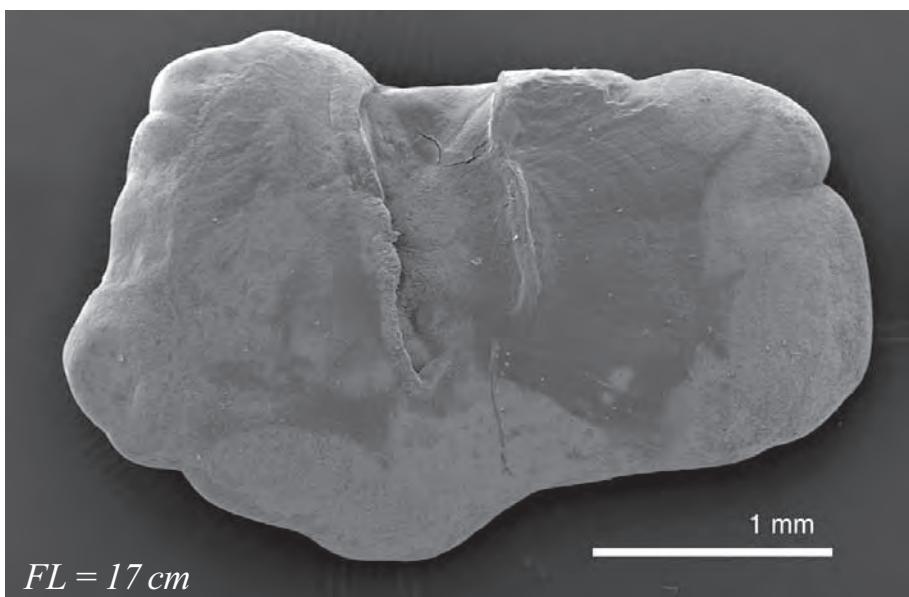
Perciformes
Carangidae: *Seriola dumerili*



Carangidae: *Seriola zonata*



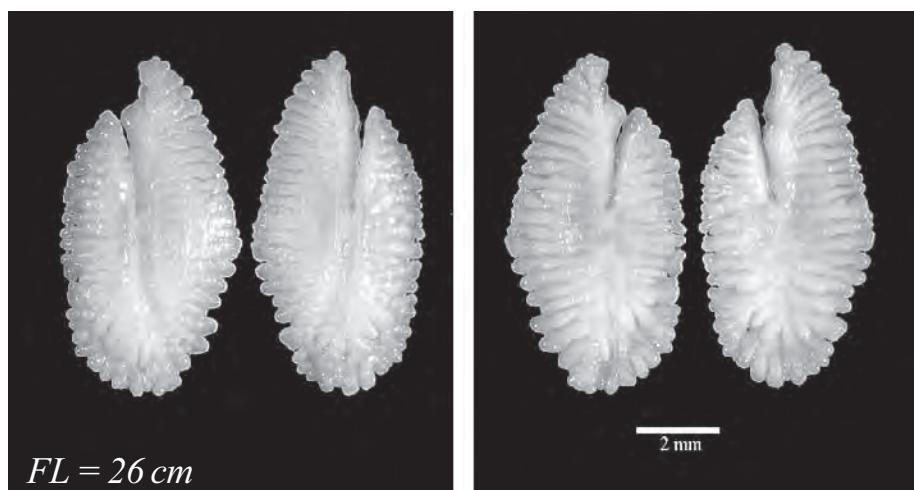
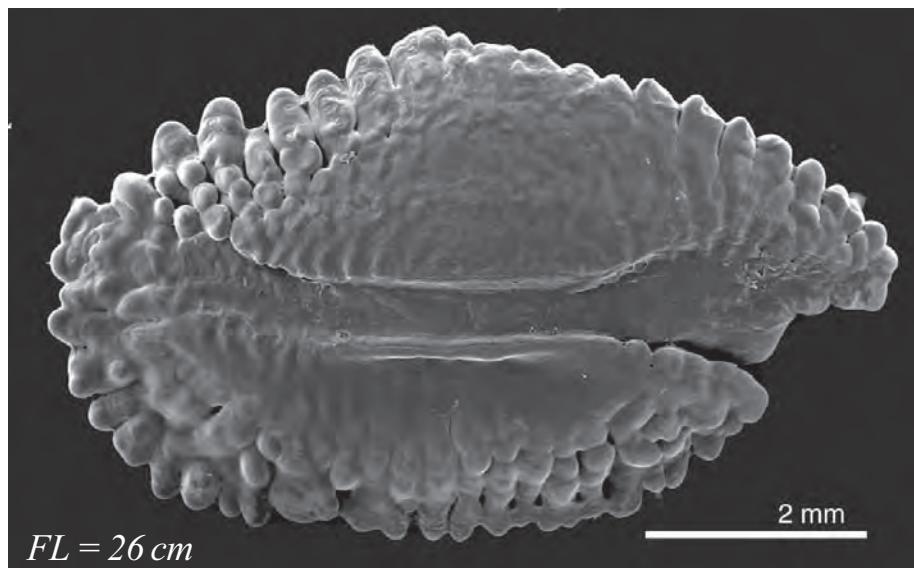
Perciformes

Caristiidae: *Caristius groenlandicus*

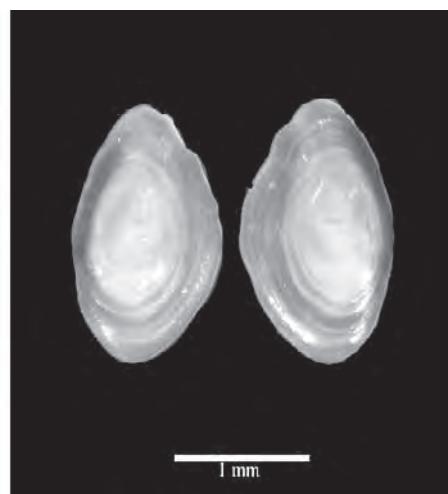
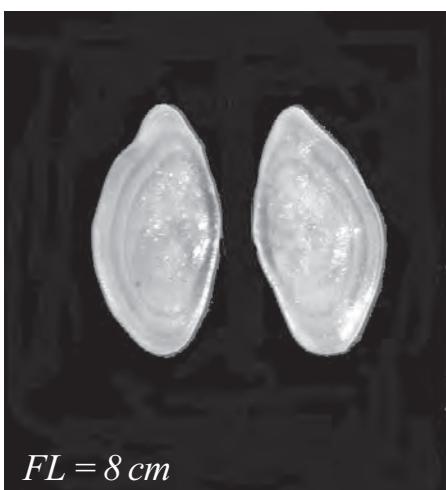
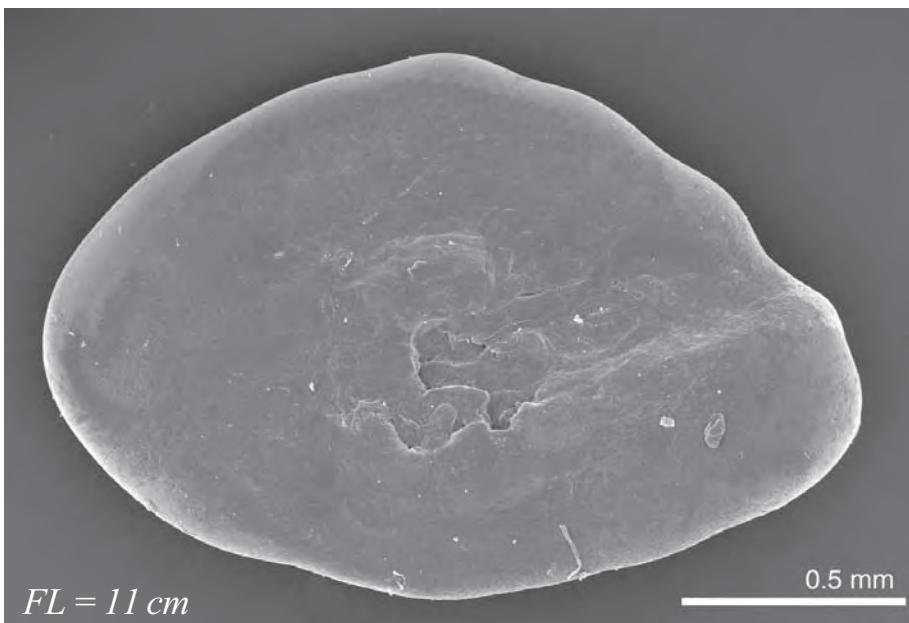
Perciformes

155

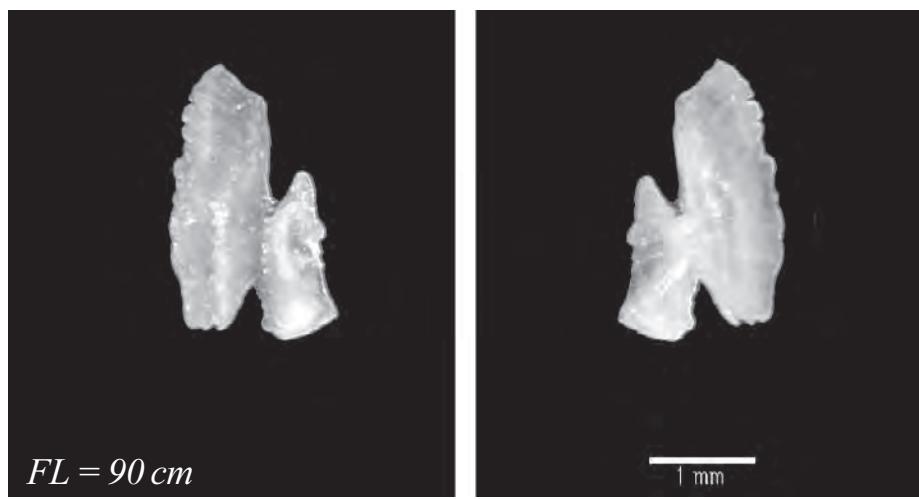
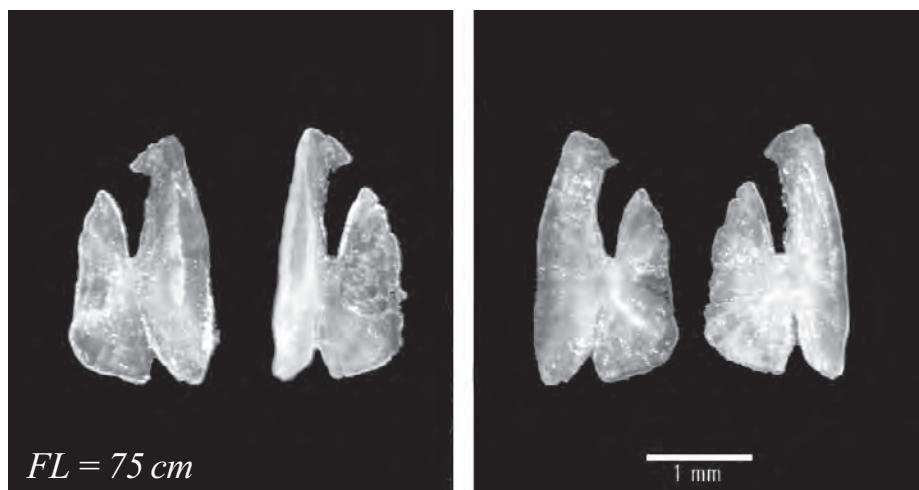
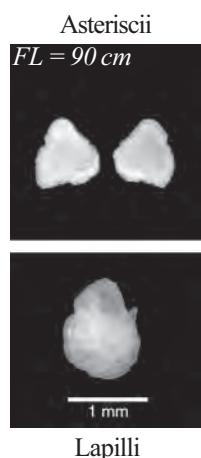
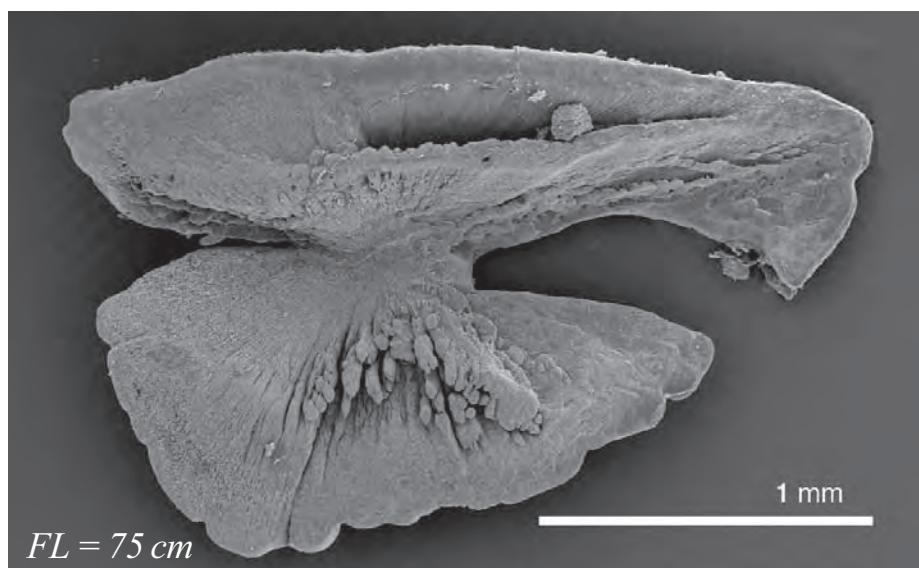
Centrolophidae: *Hyperoglyphe perciformis*



Perciformes
Chiasmodontidae: *Chiasmodon niger*

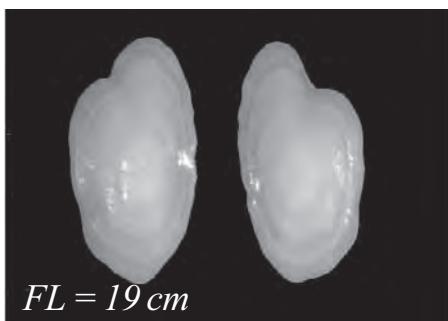
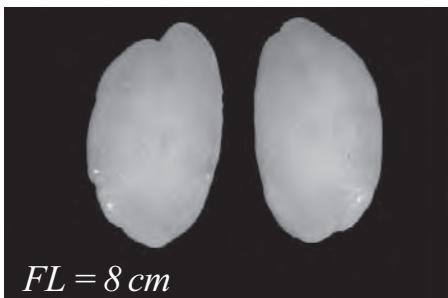
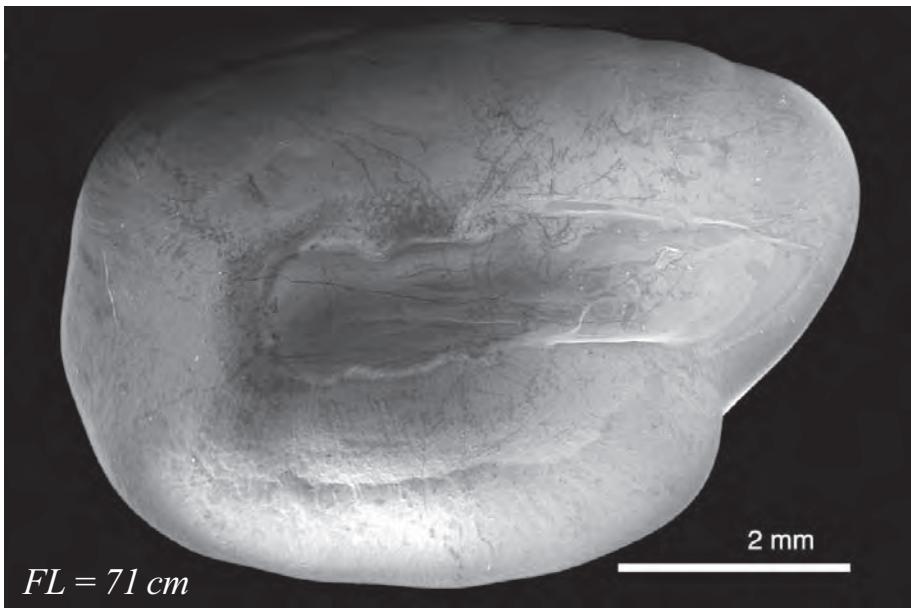


Coryphaenidae: *Coryphaena hippurus*

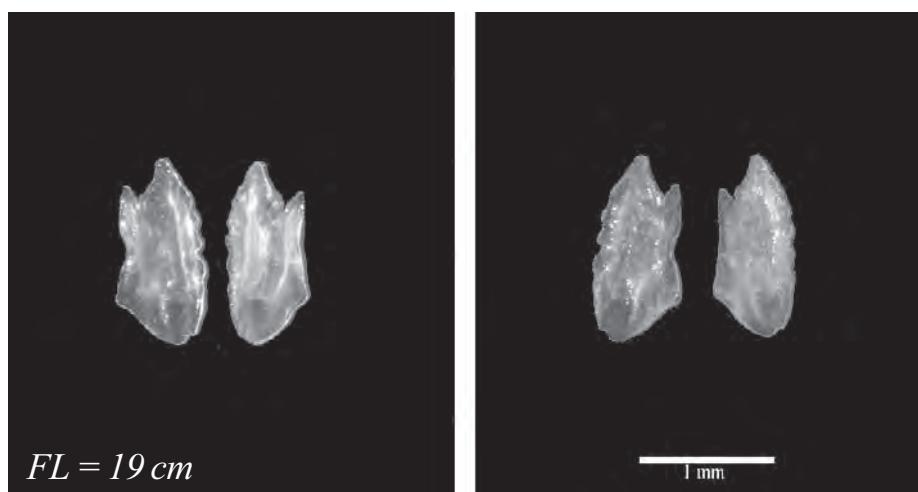
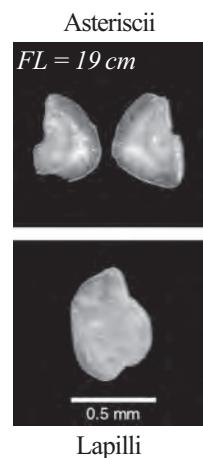
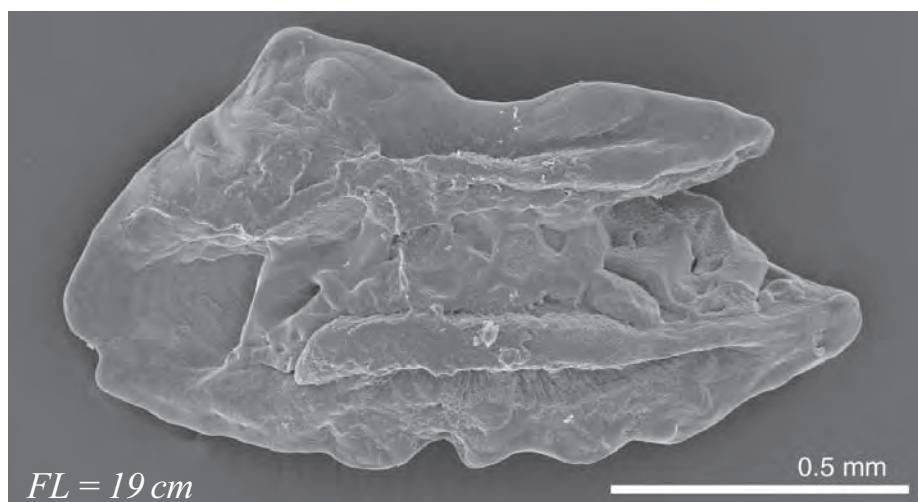


Perciformes

Cryptacanthodidae: *Cryptacanthodes maculatus*

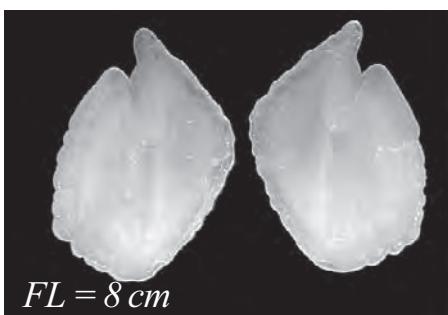
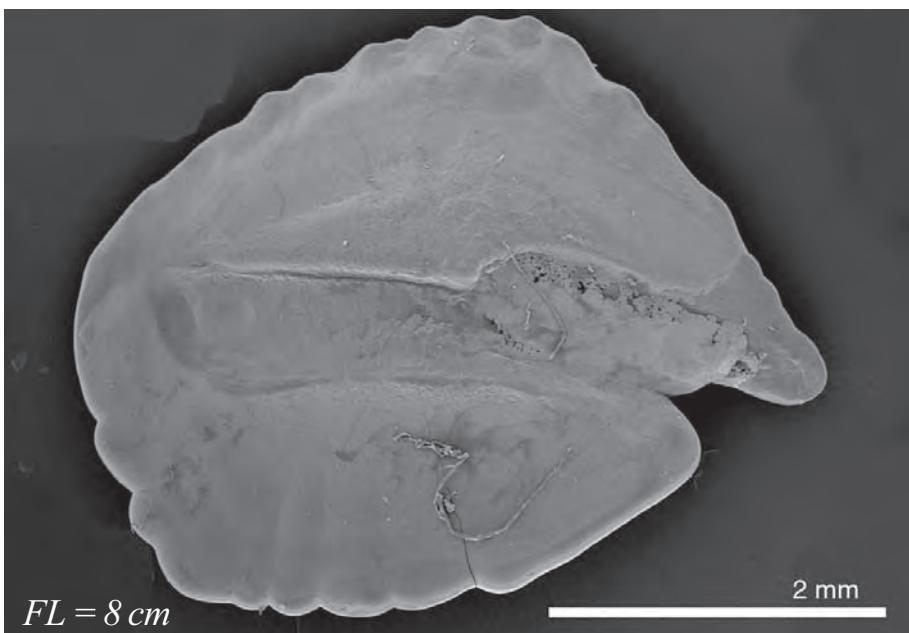


Echeneidae: *Echeneis naucrates*



Perciformes

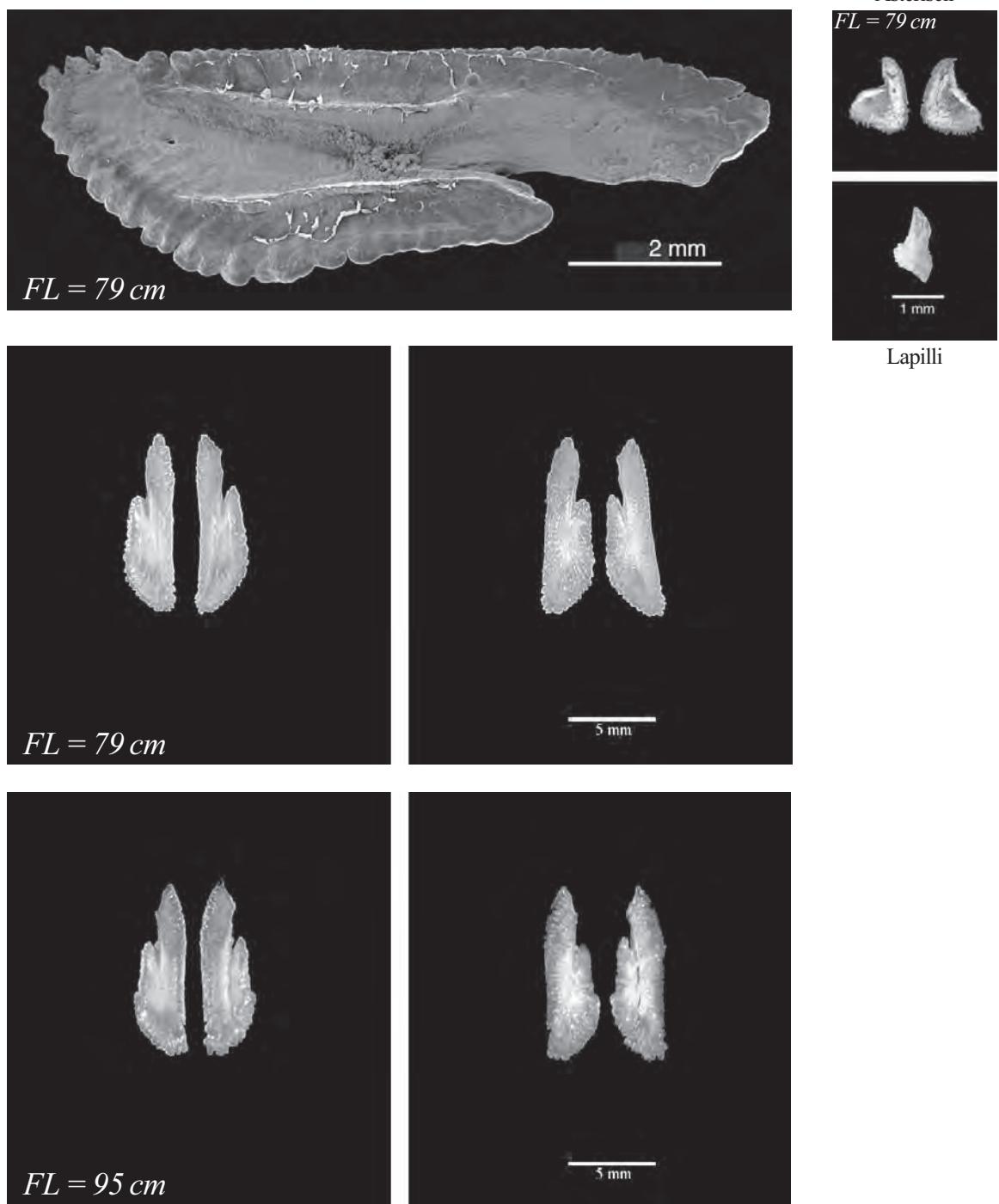
Epigonidae: *Epigonus telescopus*



Perciformes

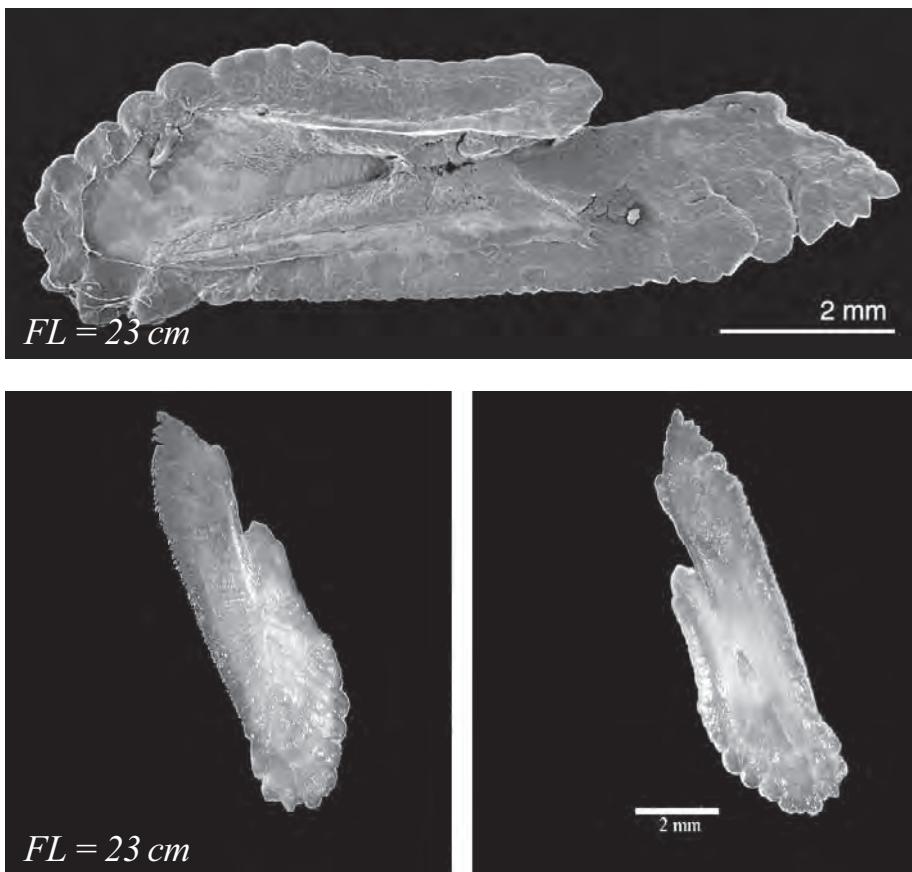
161

Gempylidae: *Lepidocybium flavobrunneum*

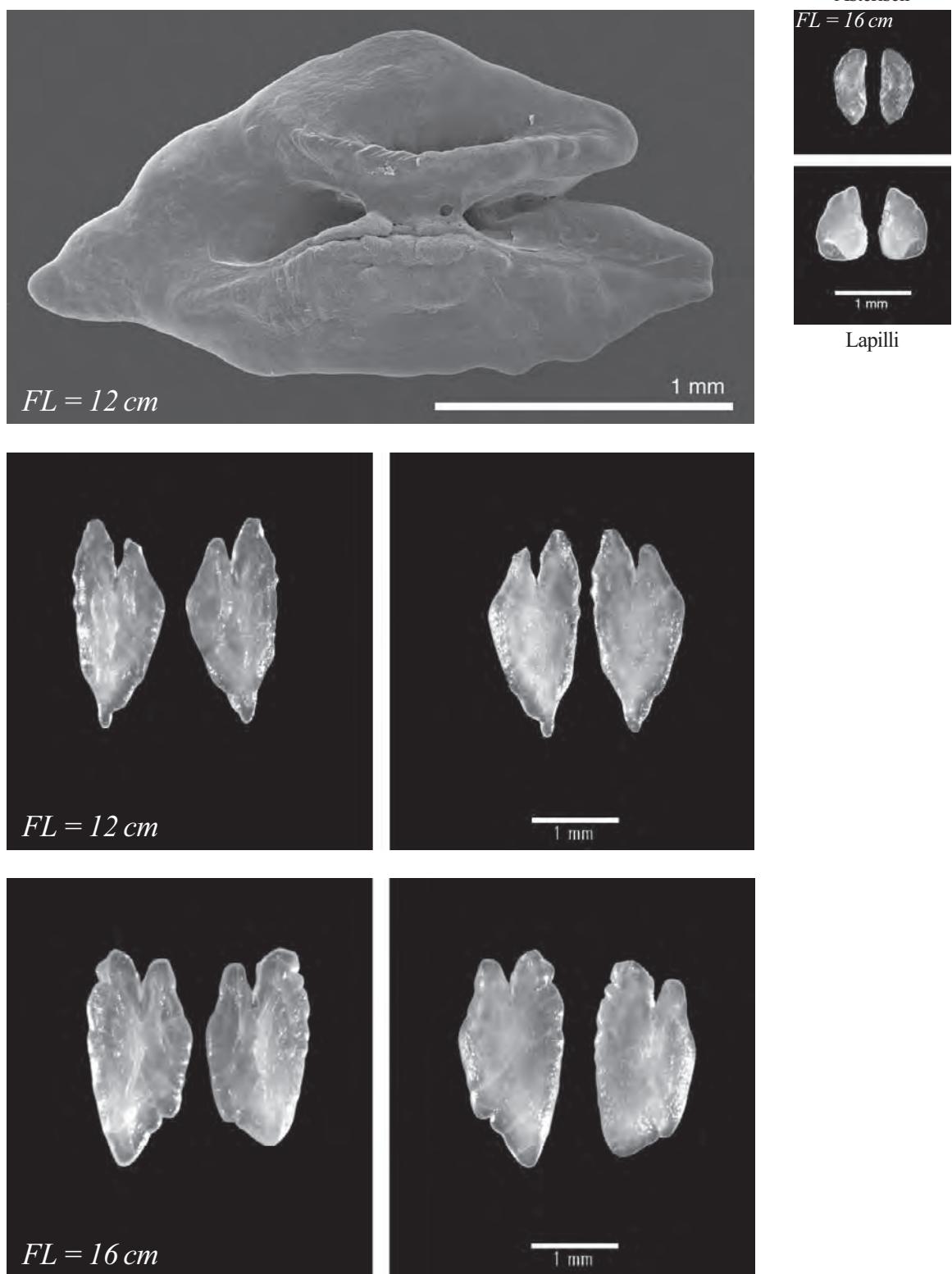


Perciformes

Gempylidae: *Ruvettus pretiosus*

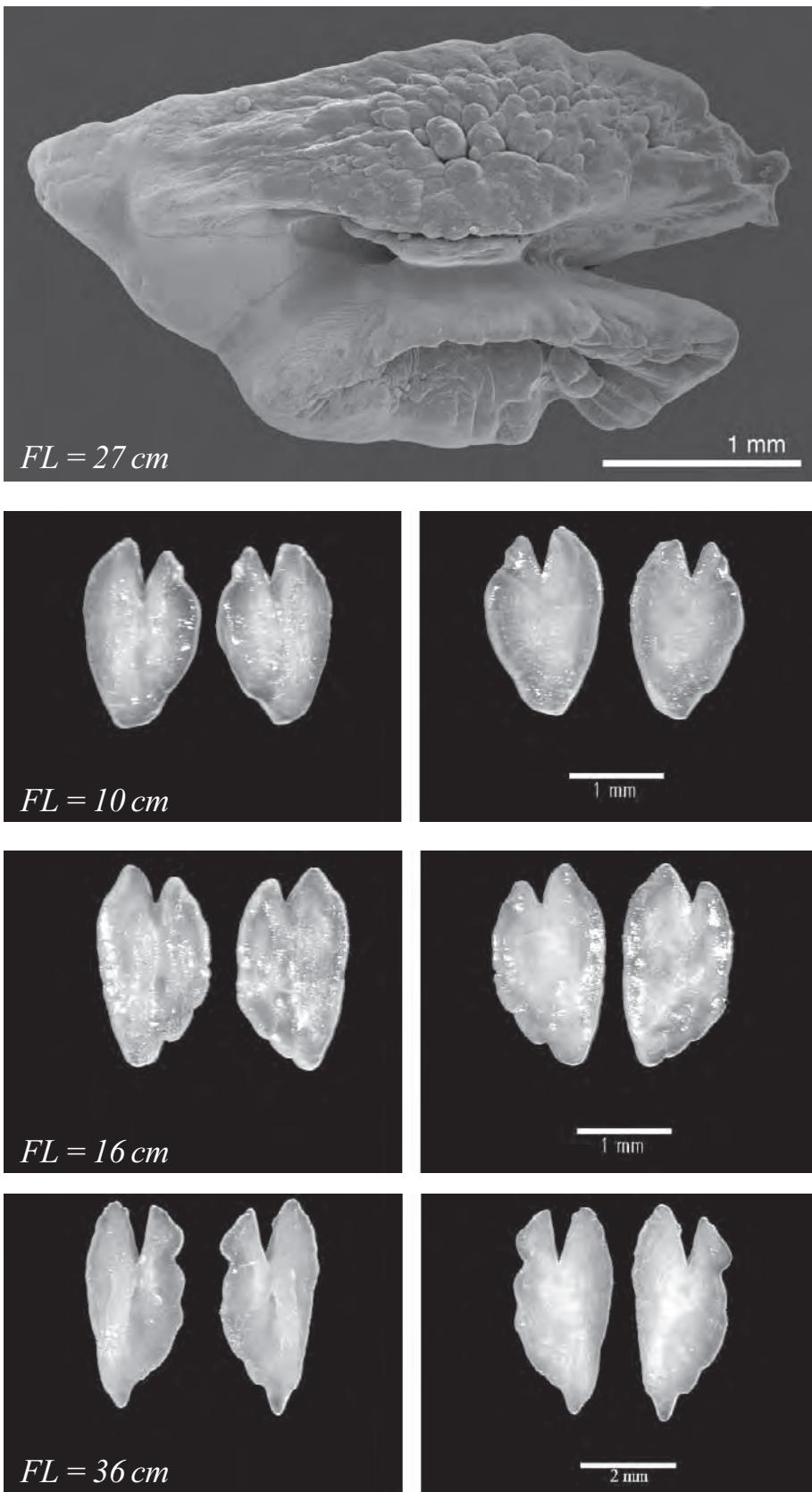


Labridae: *Tautoga onitis*



Perciformes

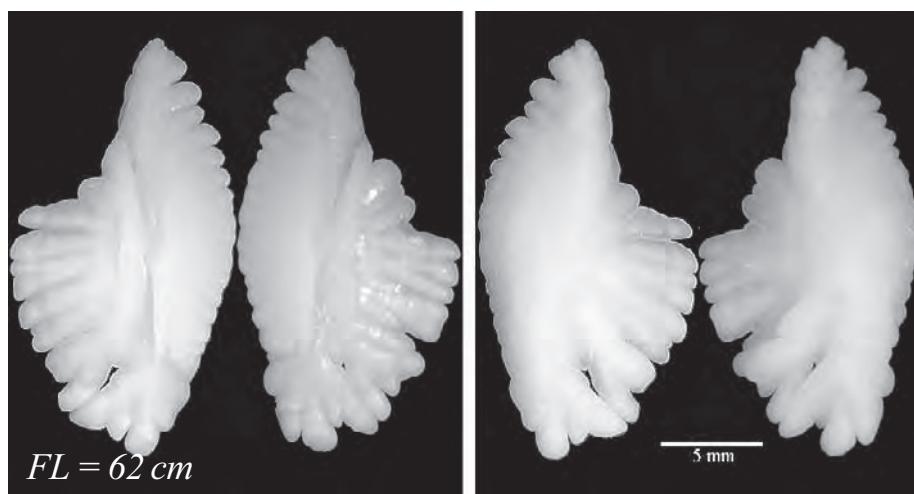
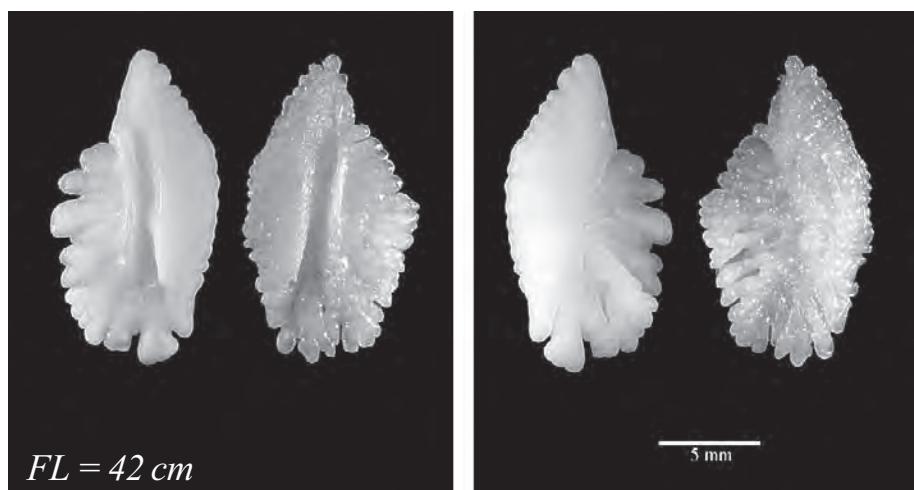
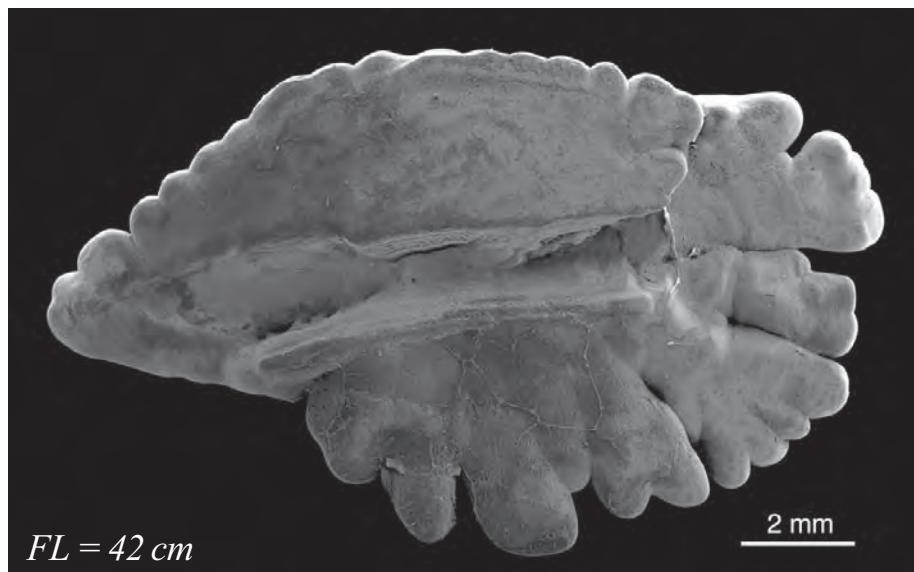
Labridae: *Tautogolabrus adspersus*



Perciformes

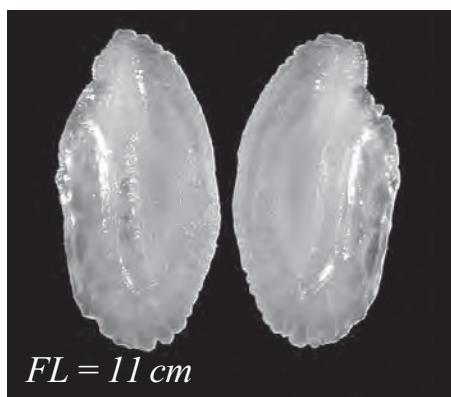
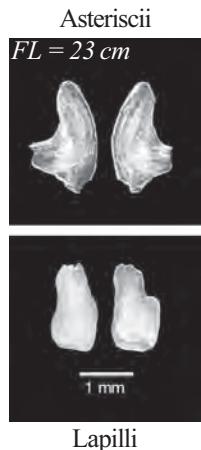
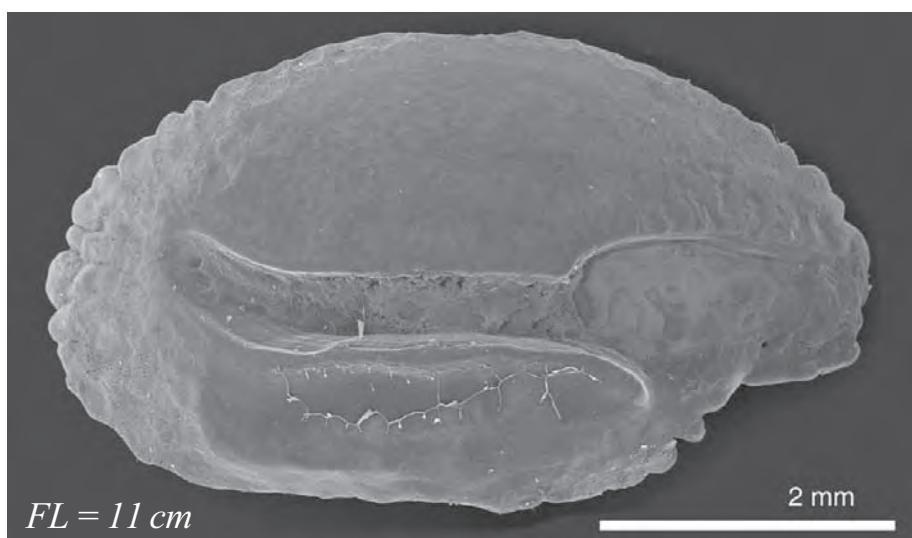
165

Malacanthidae: *Lopholatilus chamaeleonticeps*



Perciformes

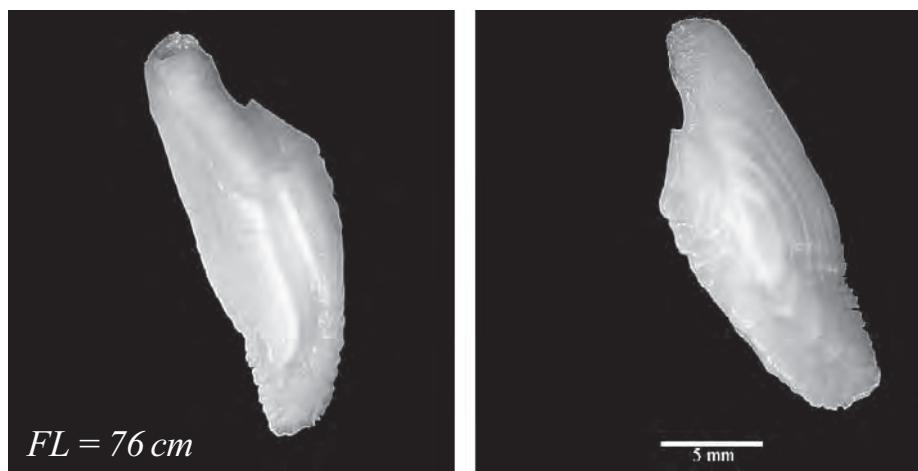
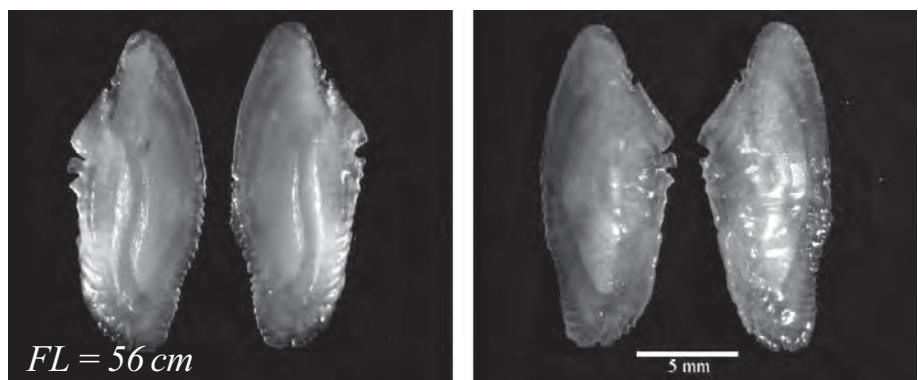
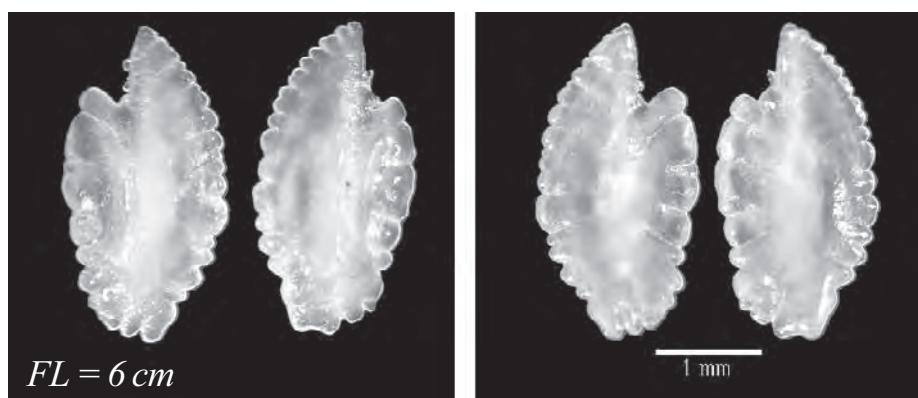
Moronidae: *Morone americana*



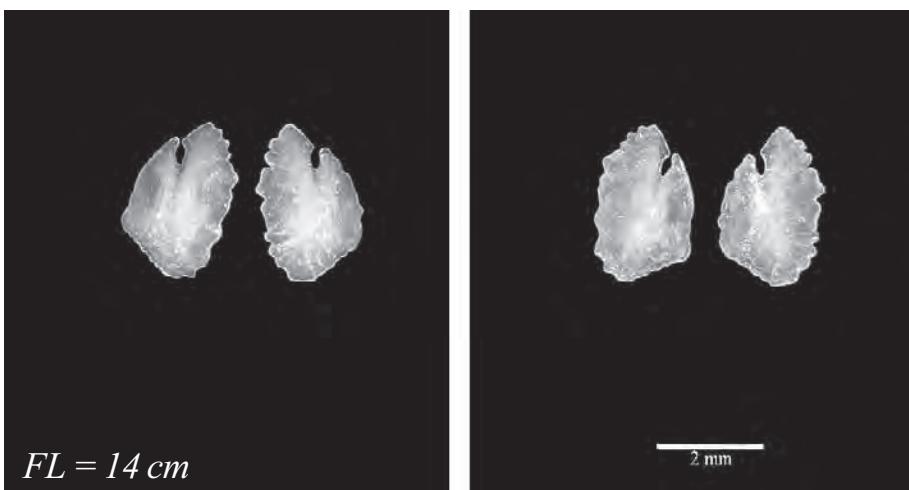
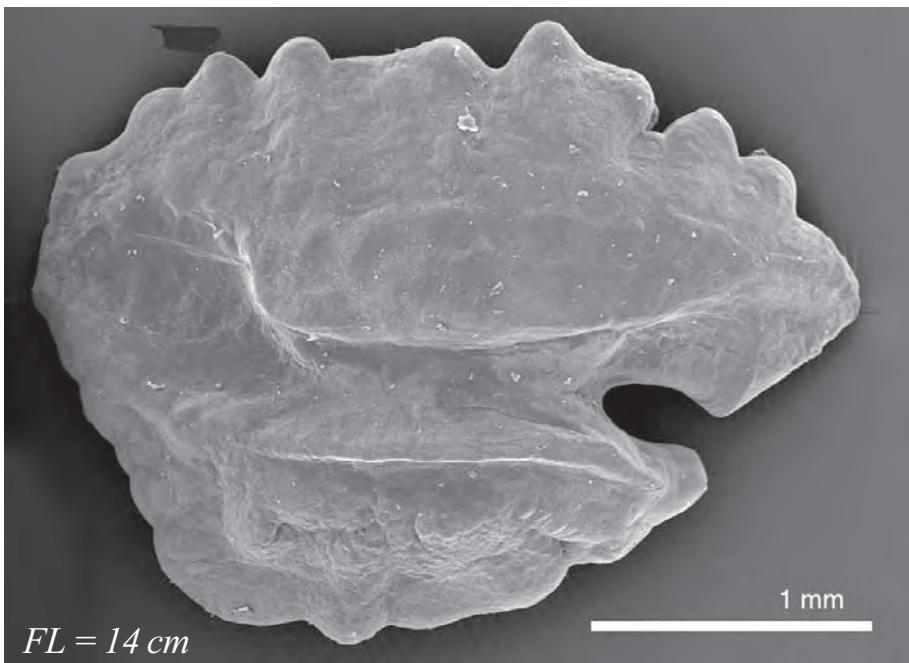
Perciformes

167

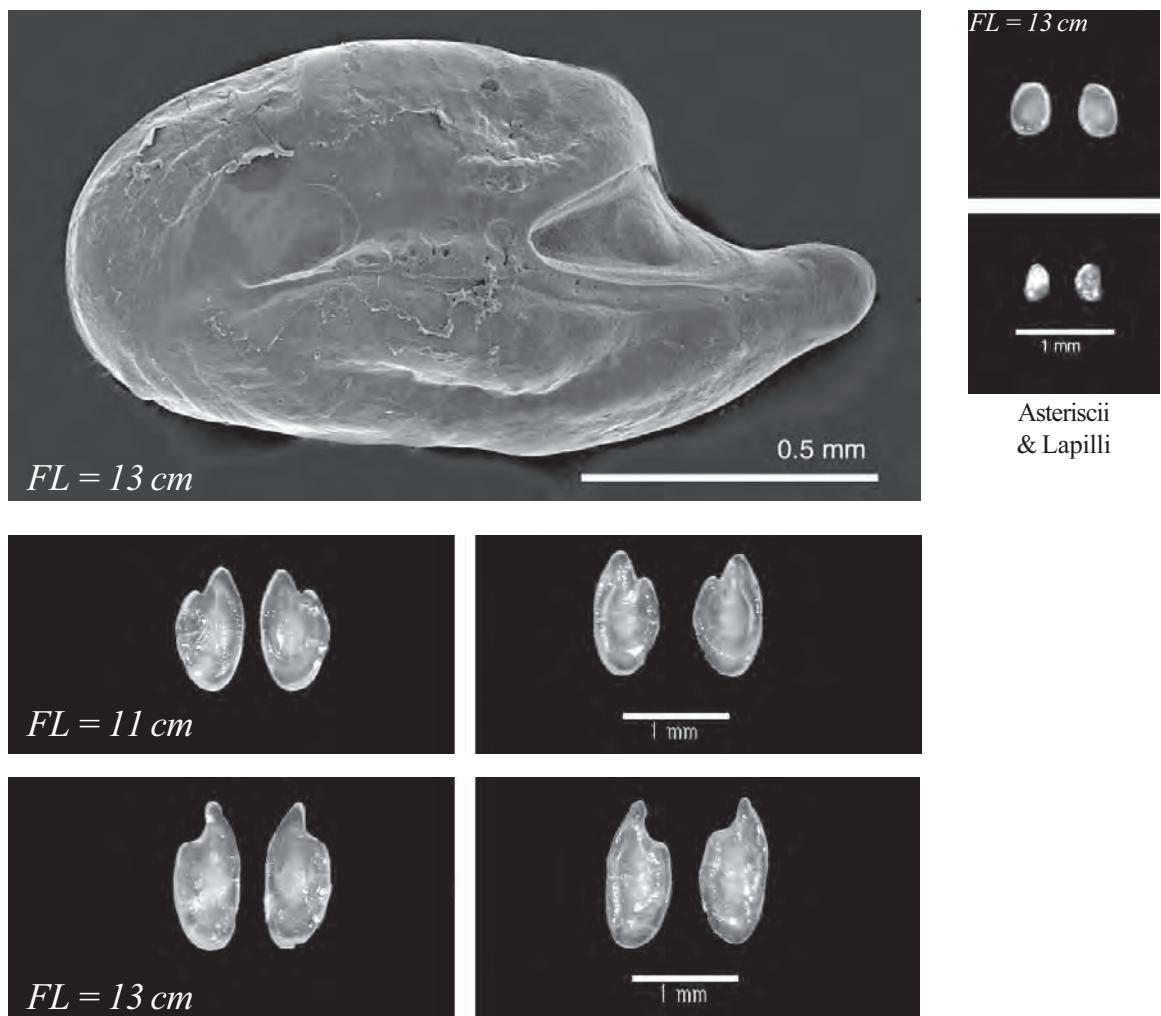
Moronidae: *Morone saxatilis*



Perciformes
Mullidae: *Mullus auratus*



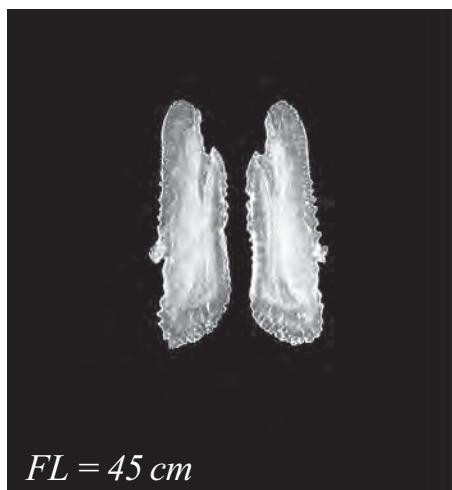
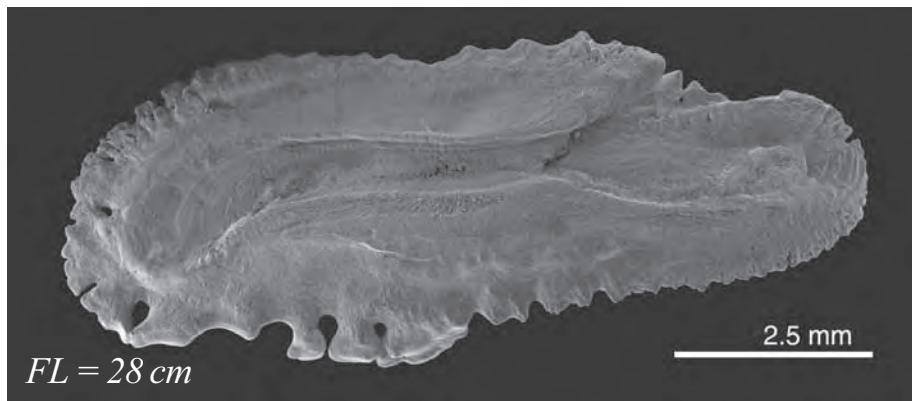
Pholidae: *Pholis gunnellus*



Asterisci
& Lapilli

Perciformes

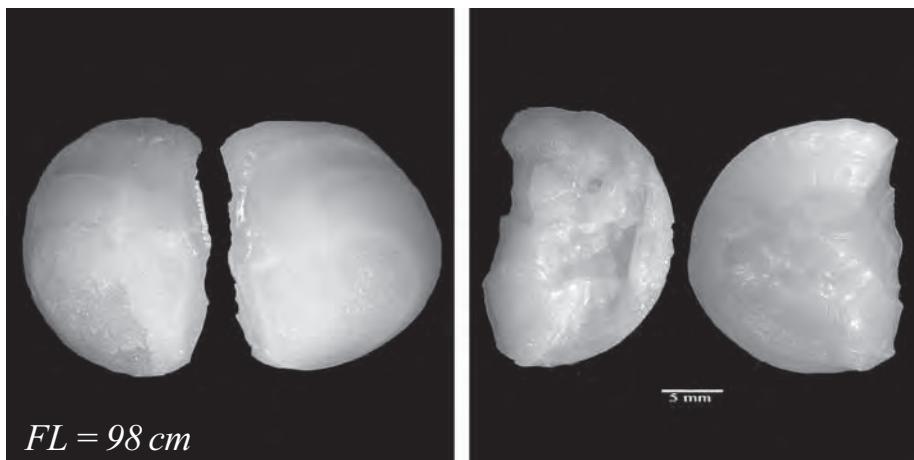
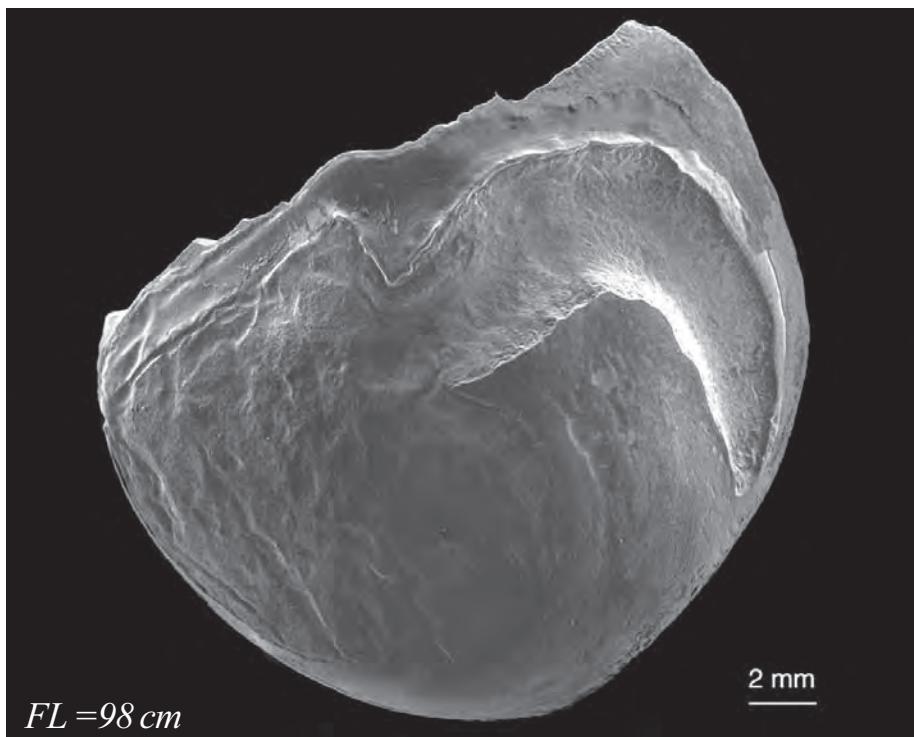
Pomatomidae: *Pomatomus saltatrix*



Perciformes

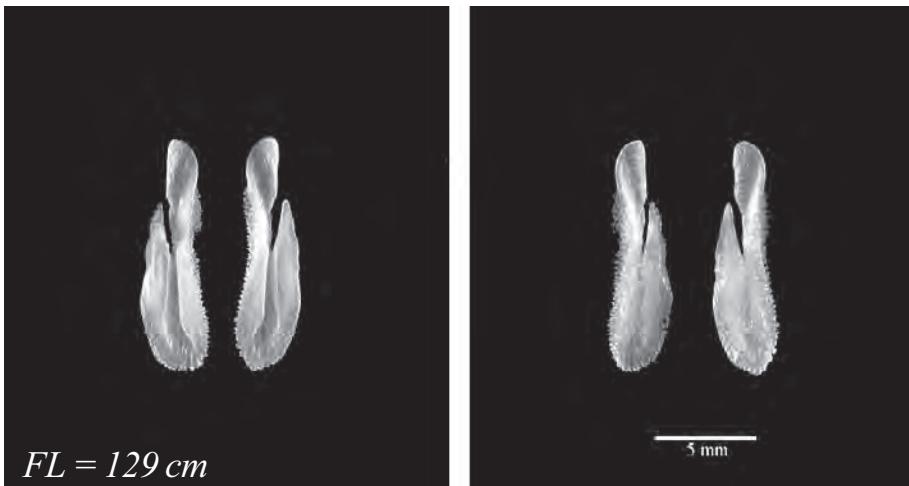
171

Sciaenidae: *Pogonias cromis*

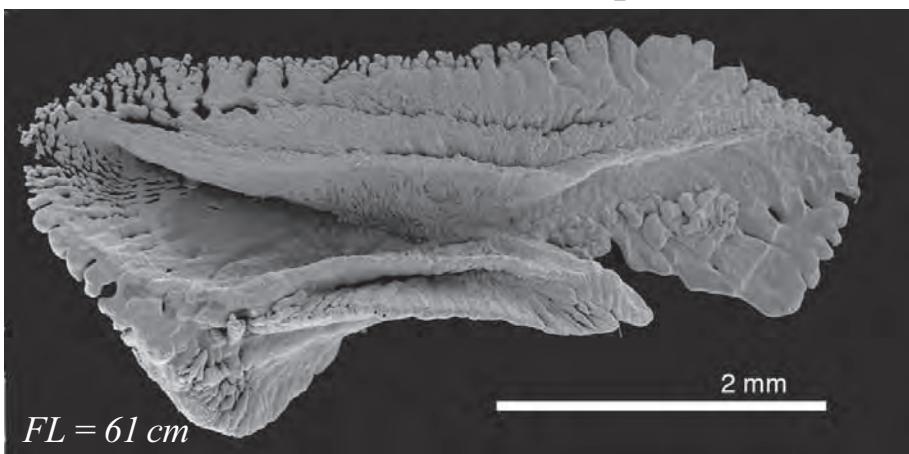


Perciformes

Scombridae: *Acanthocybium solandri*



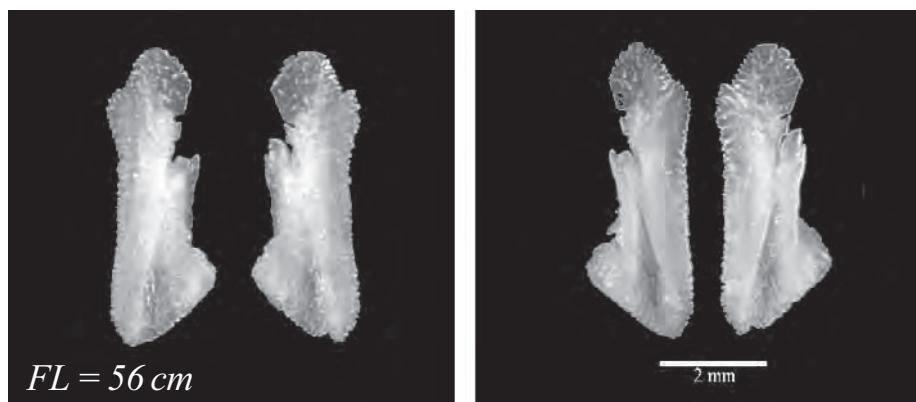
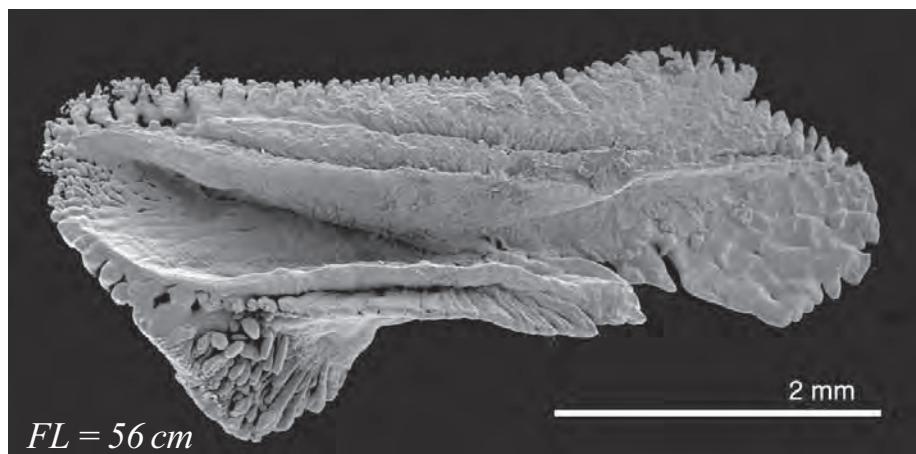
Scombridae: *Katsuwonus pelamis*



Perciformes

173

Scombridae: *Sarda sarda*



Perciformes

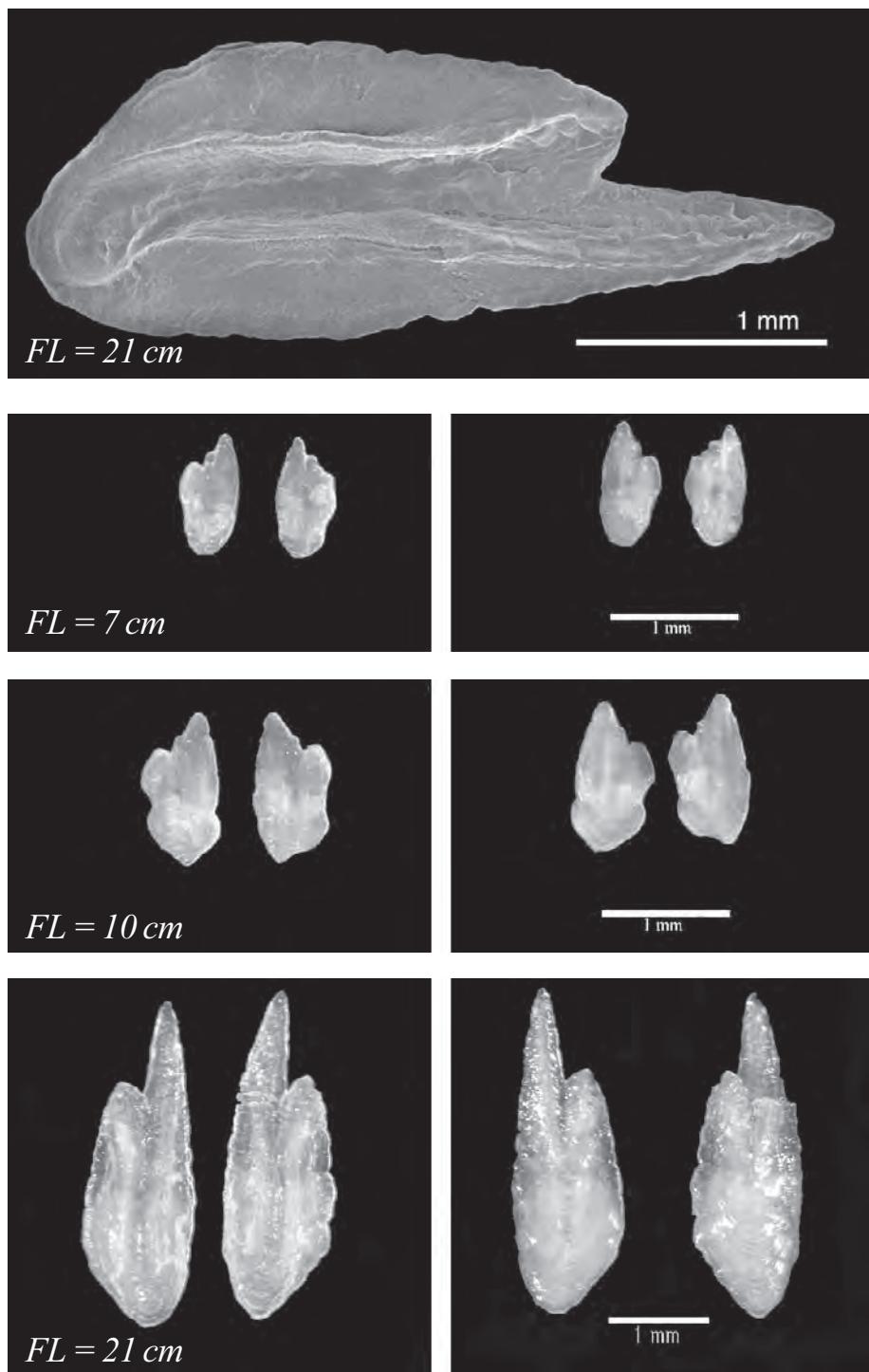
Scombridae: *Scomber japonicus*



Perciformes

175

Scombridae: *Scomber scombrus*

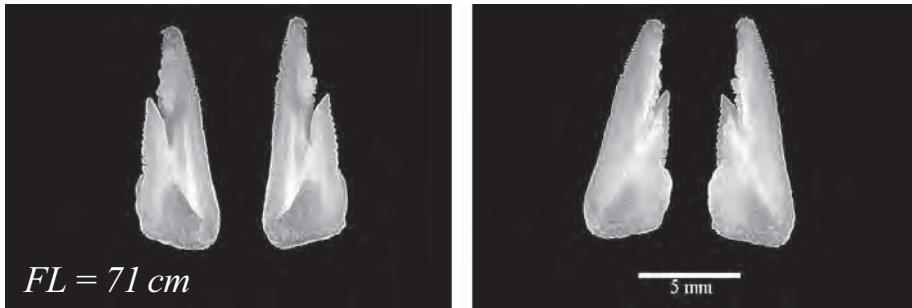
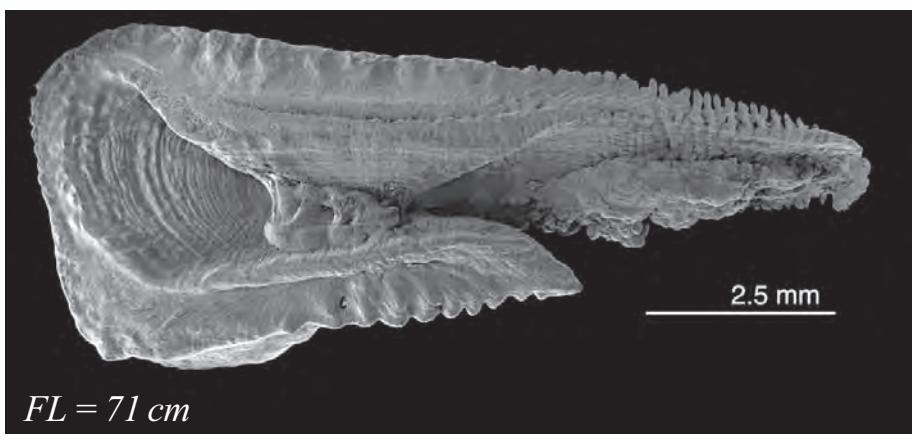


Perciformes

Scombridae: *Scomberomorus brasiliensis*



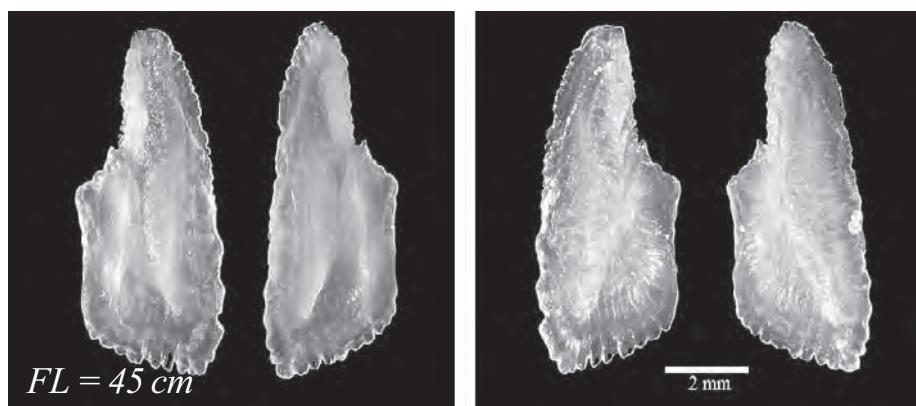
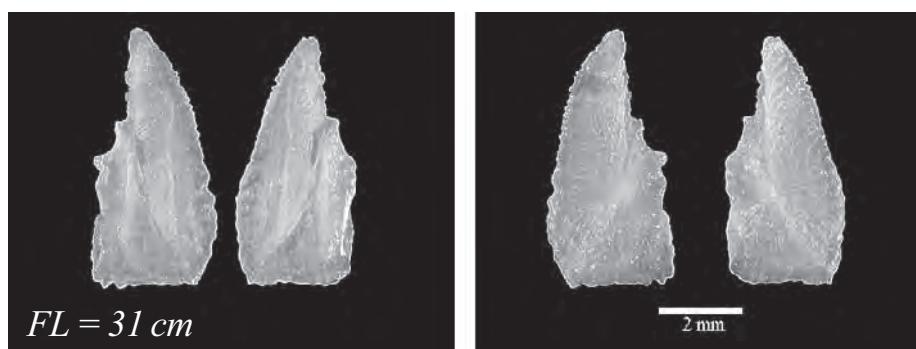
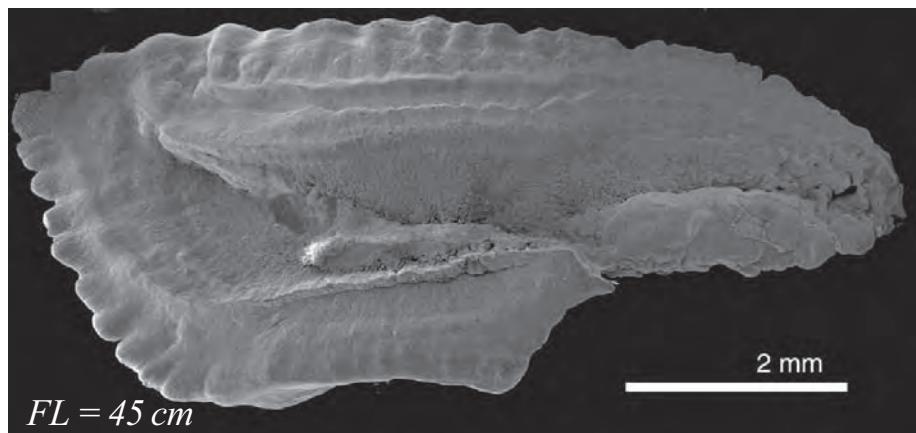
Scombridae: *Scomberomorus cavalla*



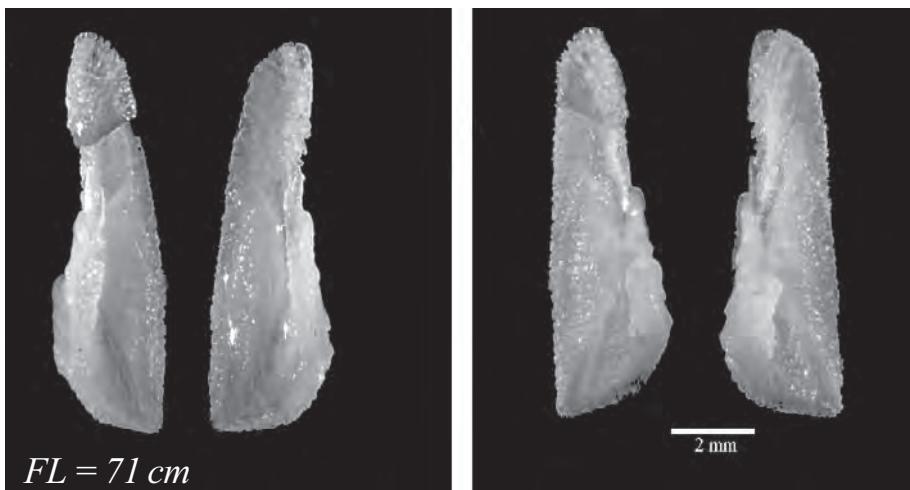
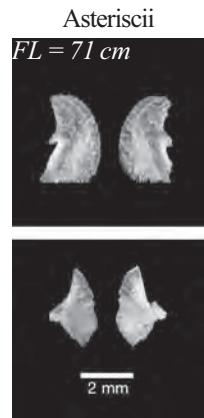
Perciformes

177

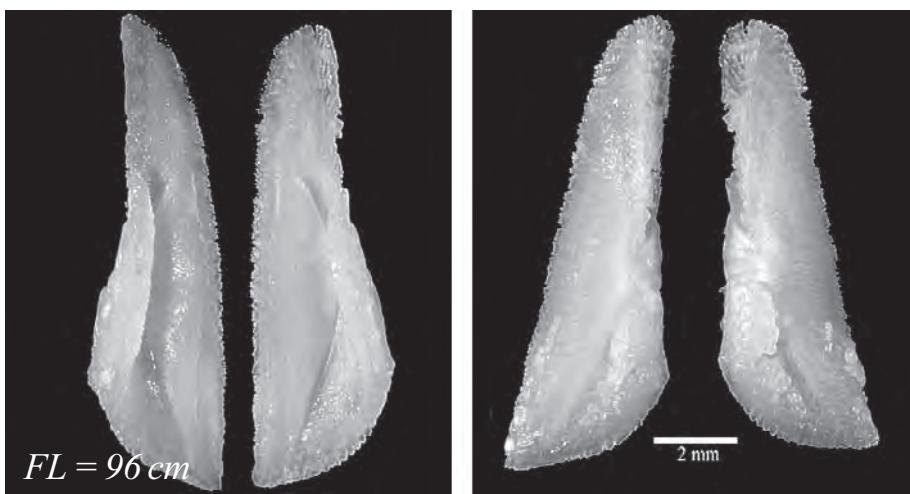
Scombridae: *Scomberomorus maculatus*



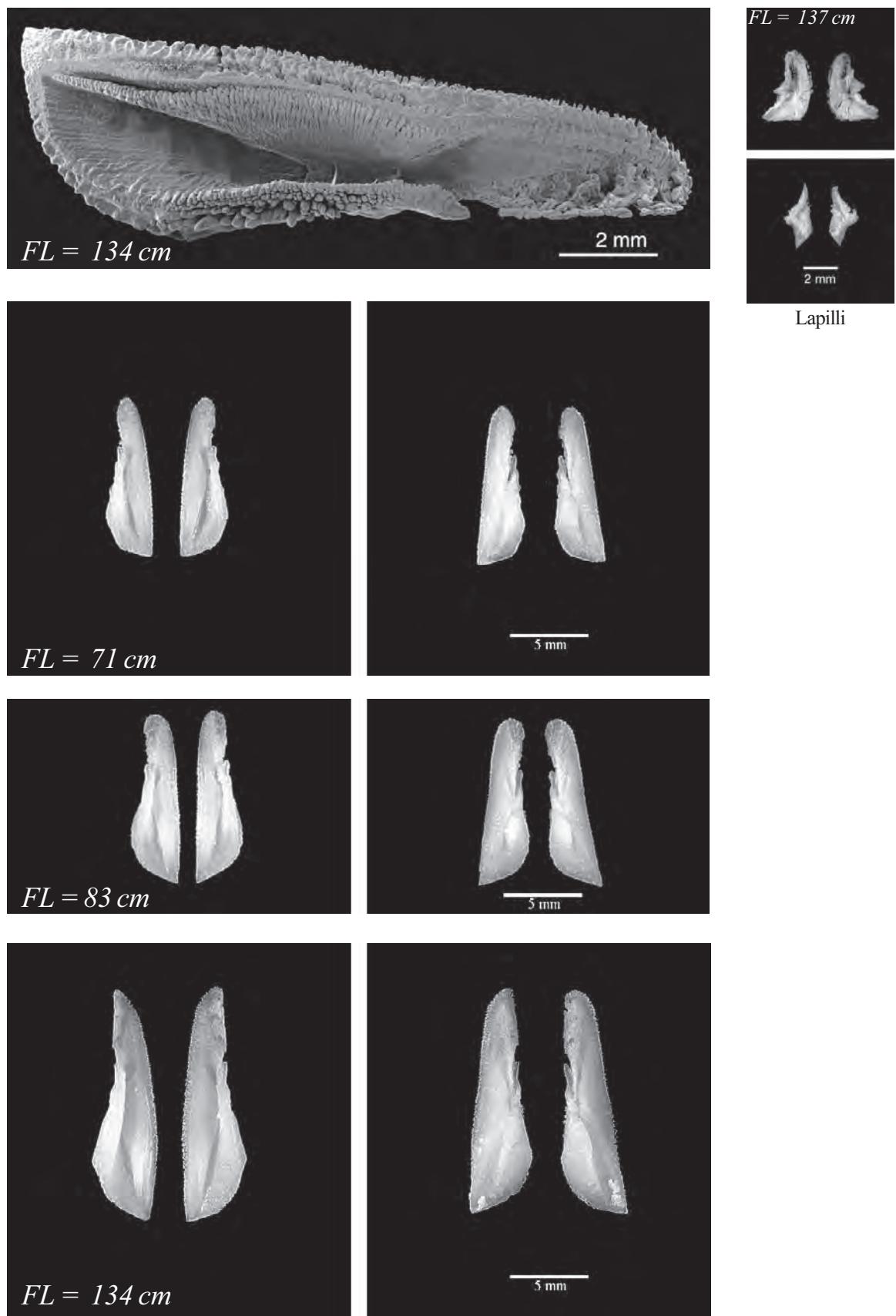
Perciformes

Scombridae: *Thunnus alalunga*

Lapilli

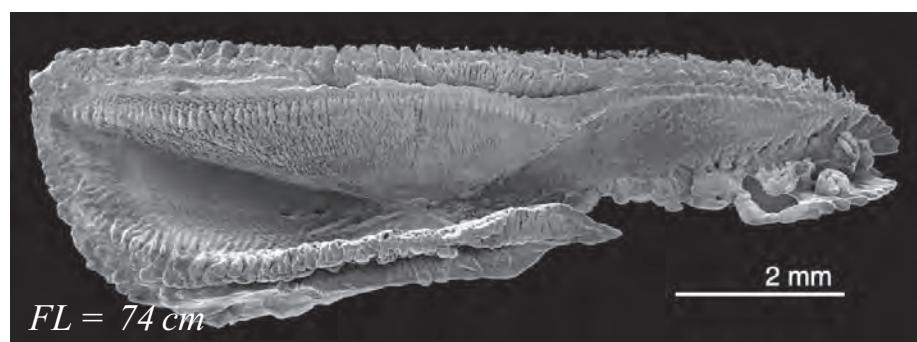


Scombridae: *Thunnus albacares*

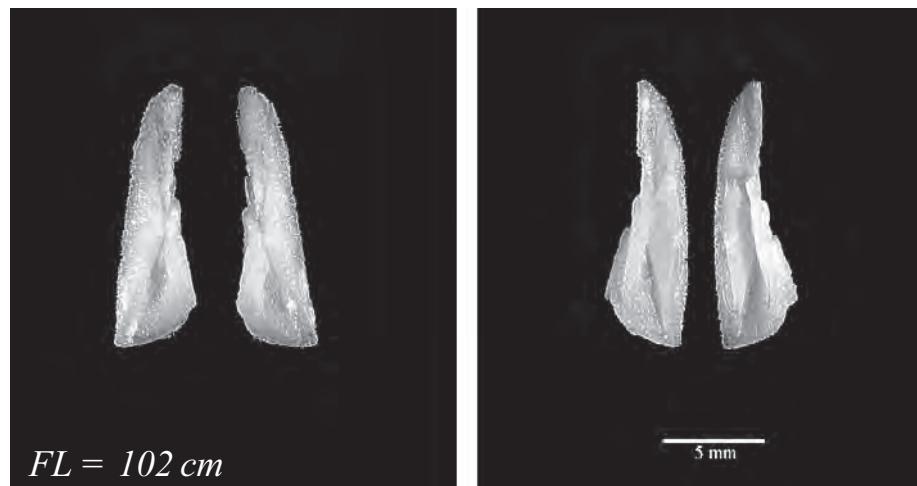


Perciformes

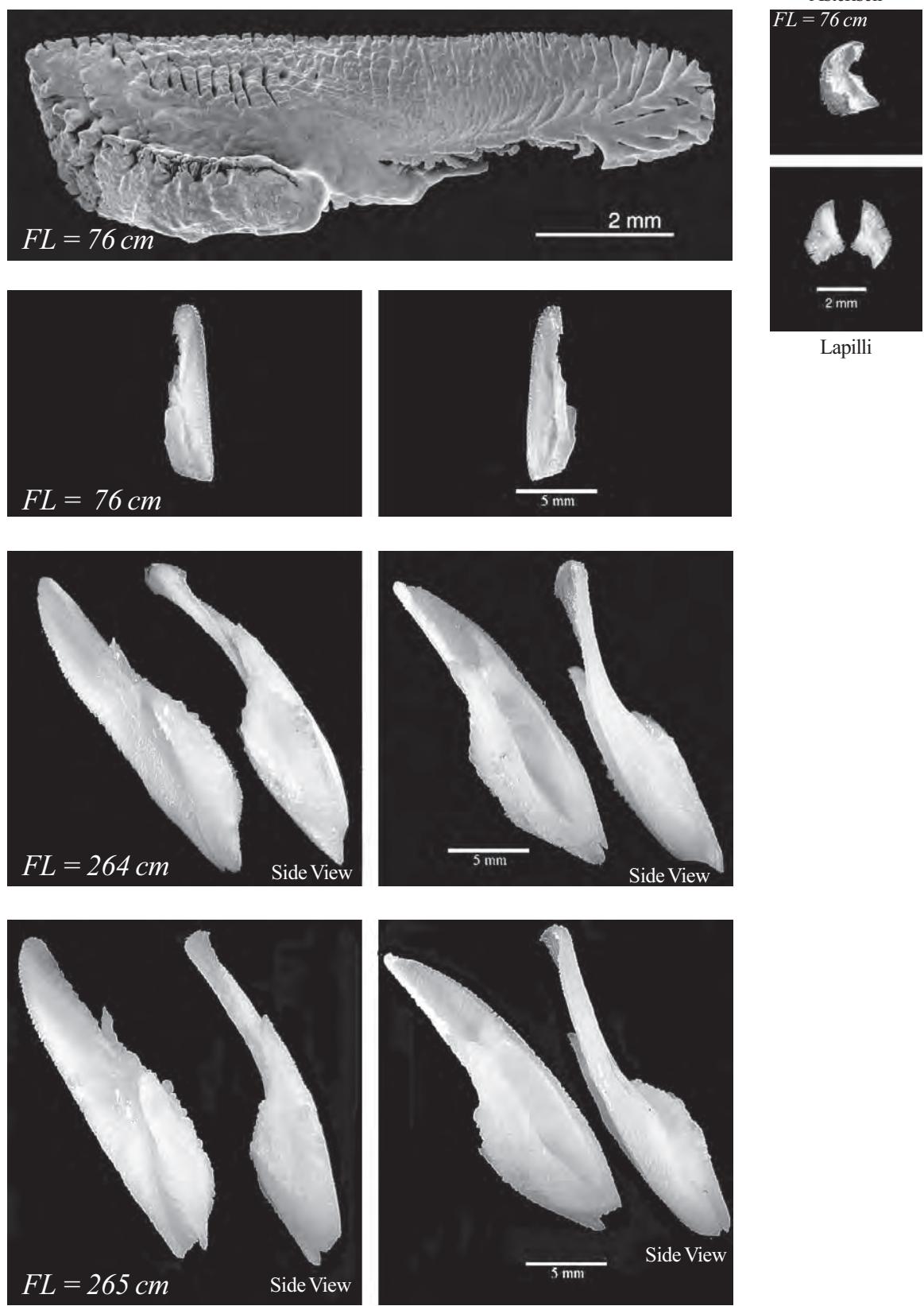
Scombridae: *Thunnus obesus*



Lapilli

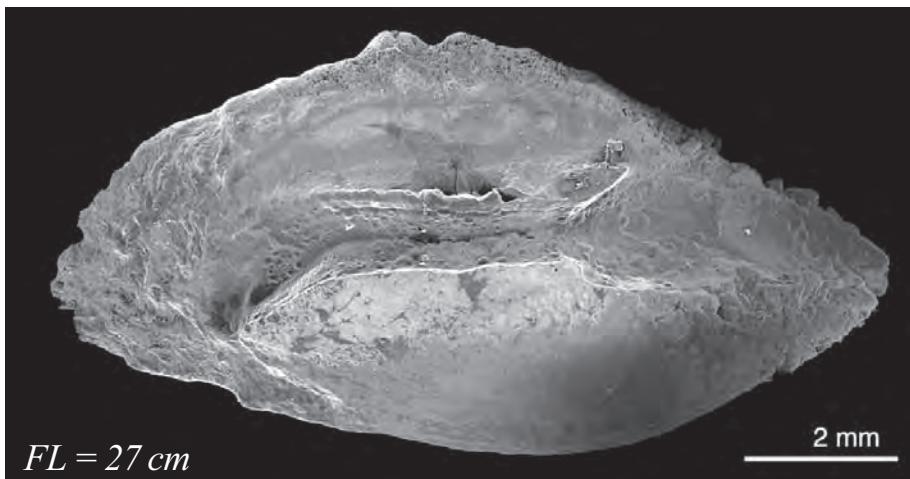


Scombridae: *Thunnus thynnus*

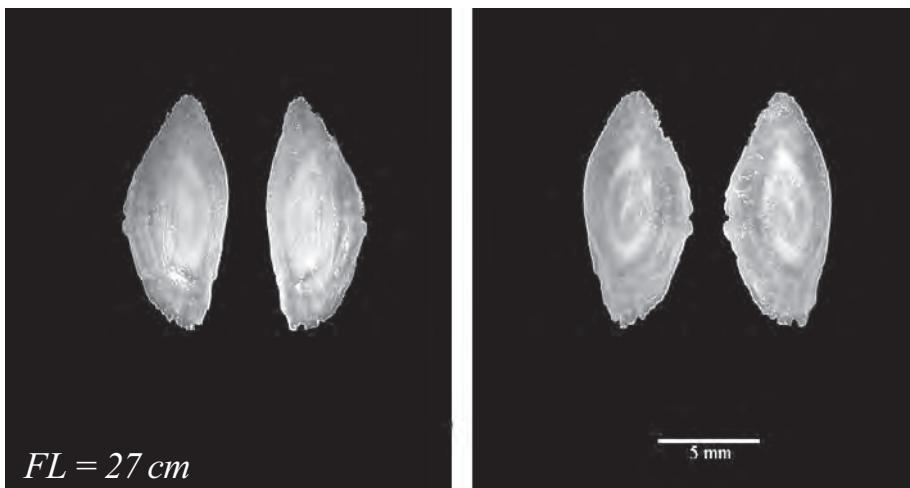


Perciformes

Serranidae: *Centropristes striata*

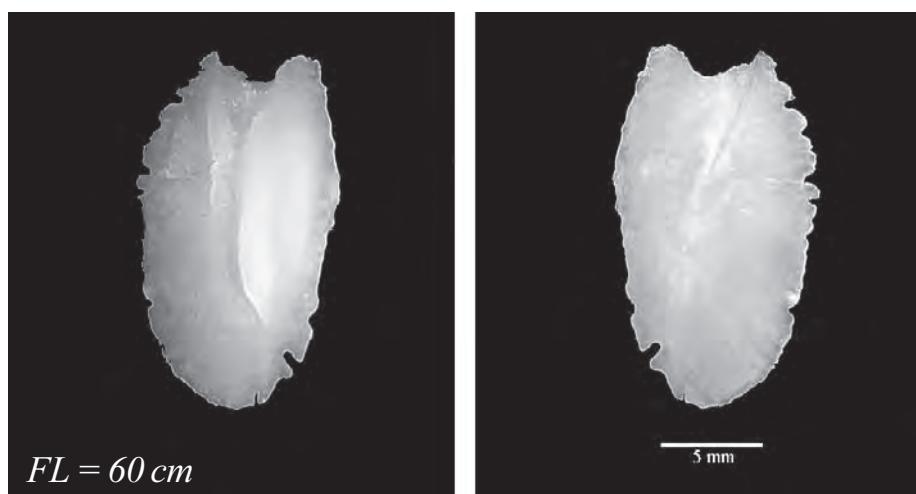
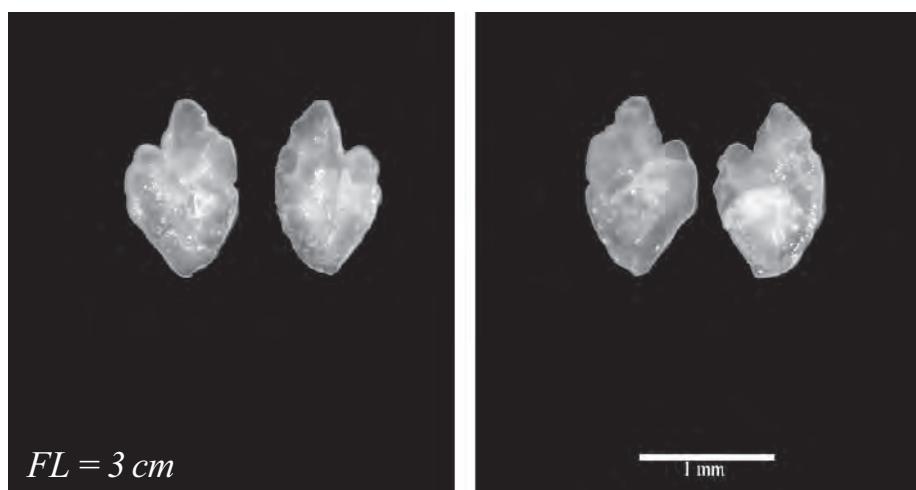
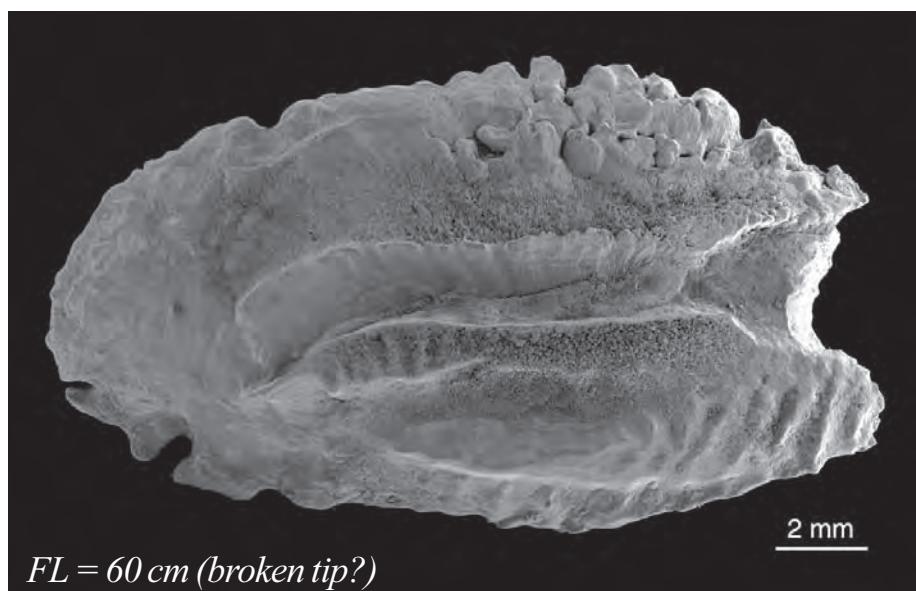


FL = 27 cm

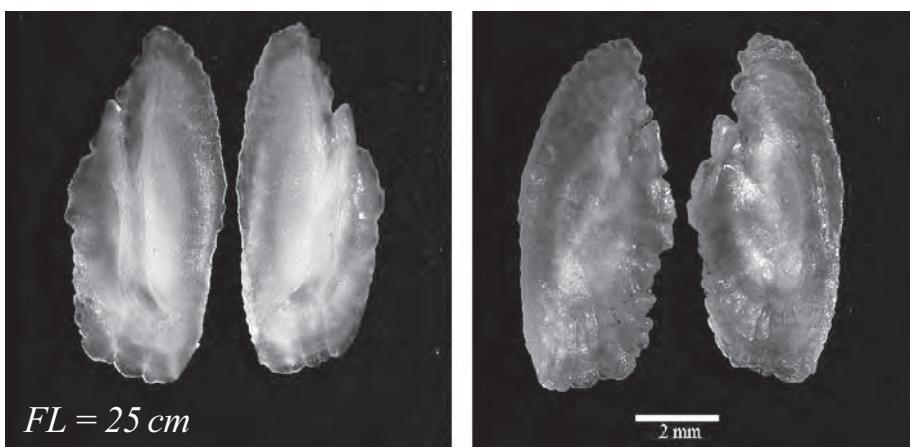
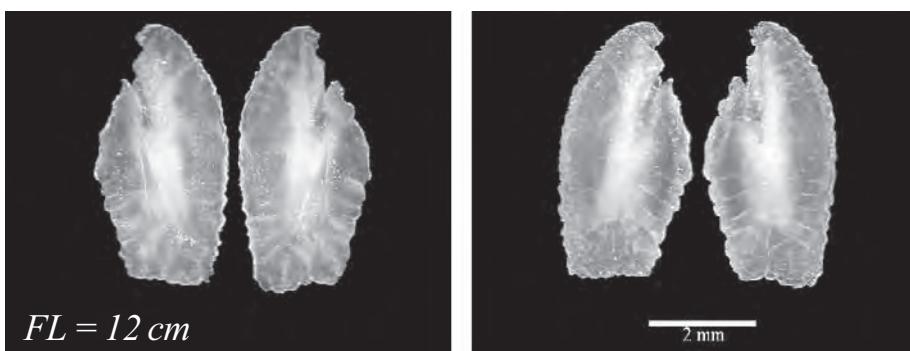
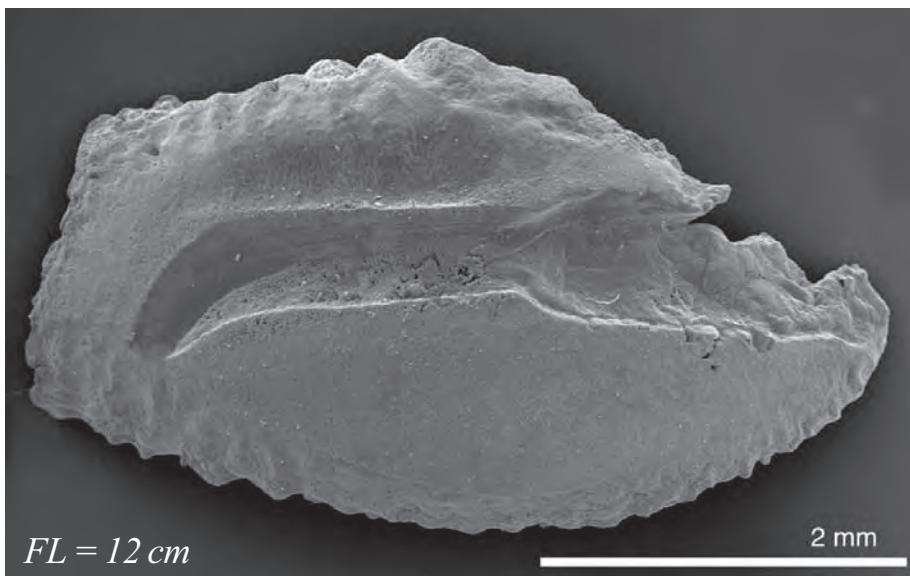


FL = 27 cm

Serranidae: *Epinephelus niveatus*



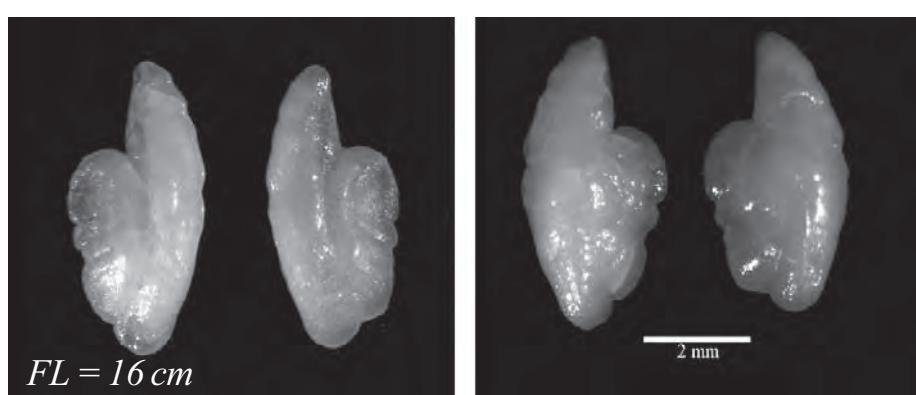
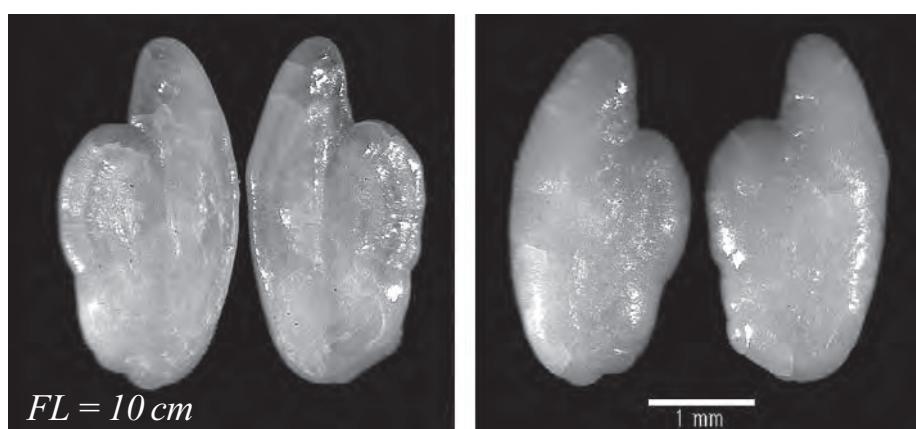
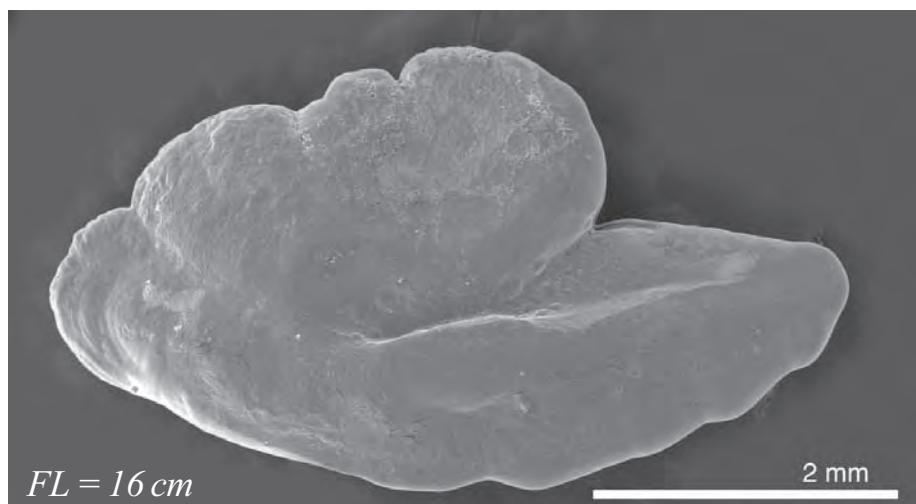
Perciformes
Sparidae: *Stenotomus chrysops*



Perciformes

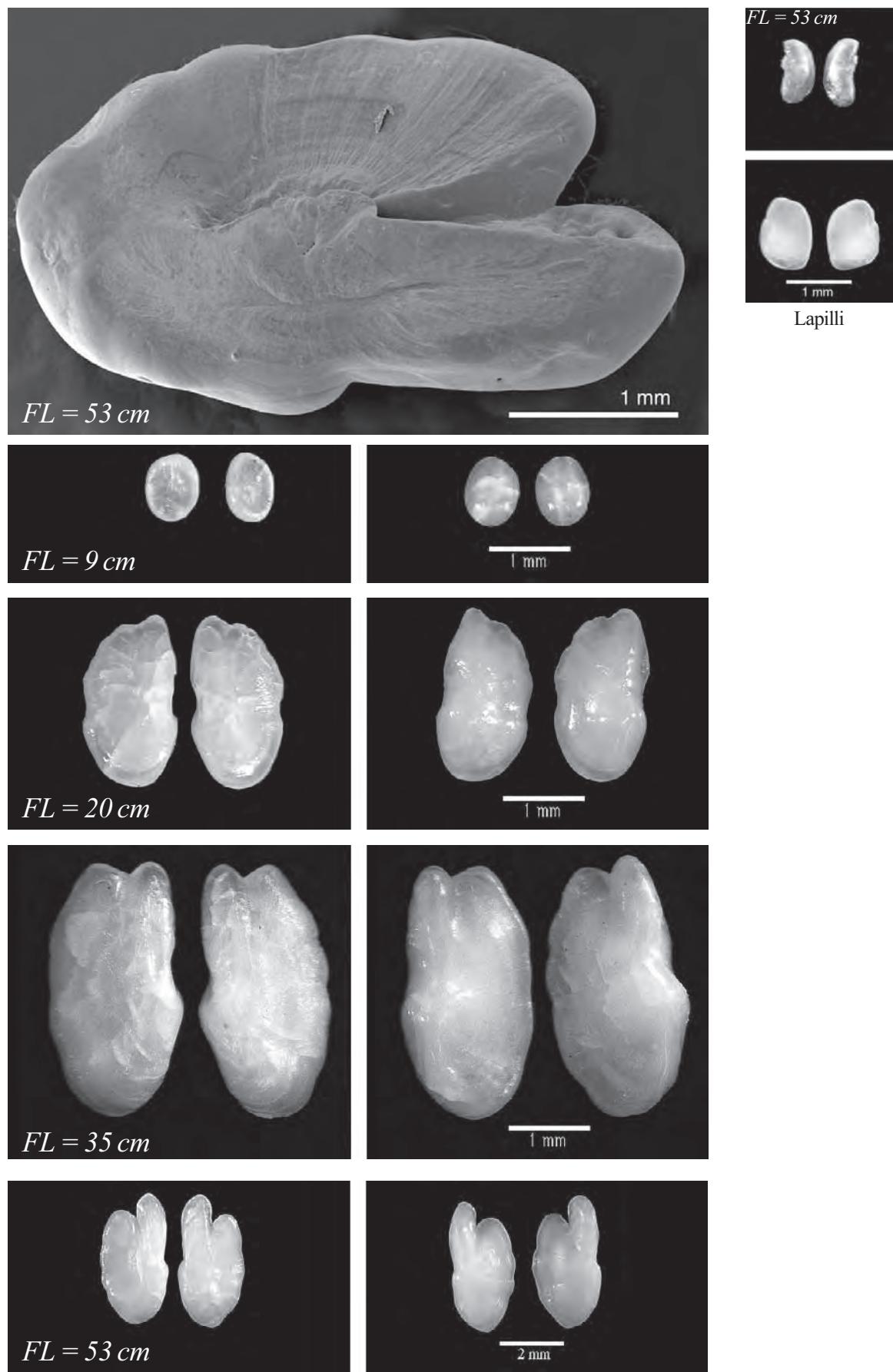
185

Stichaeidae: *Eumesogrammus praecisus*



Perciformes

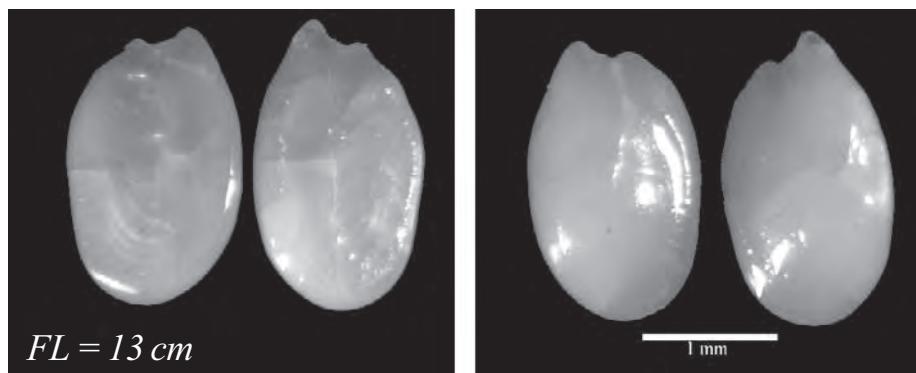
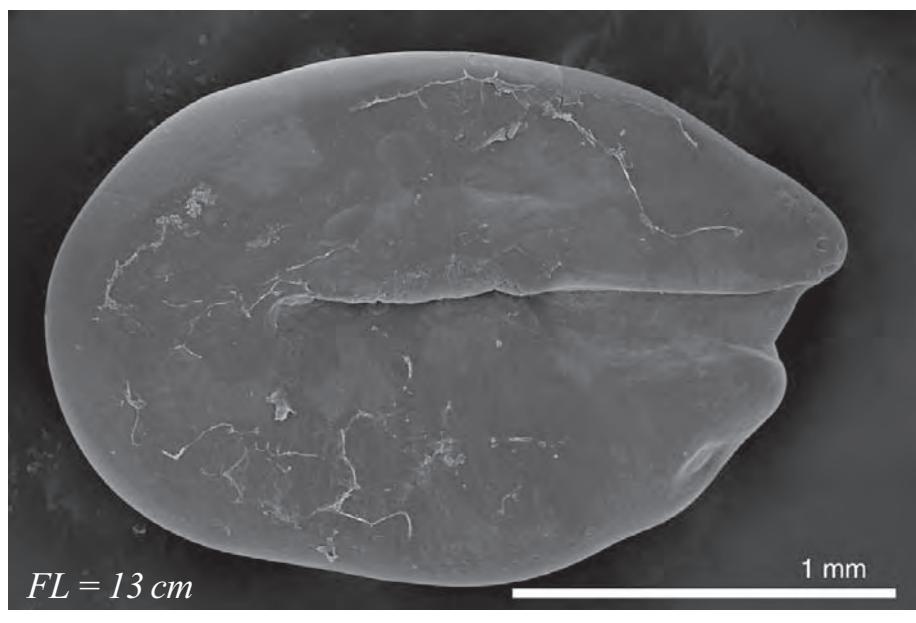
Stichaeidae: *Lumpenus lumpretaeformis*



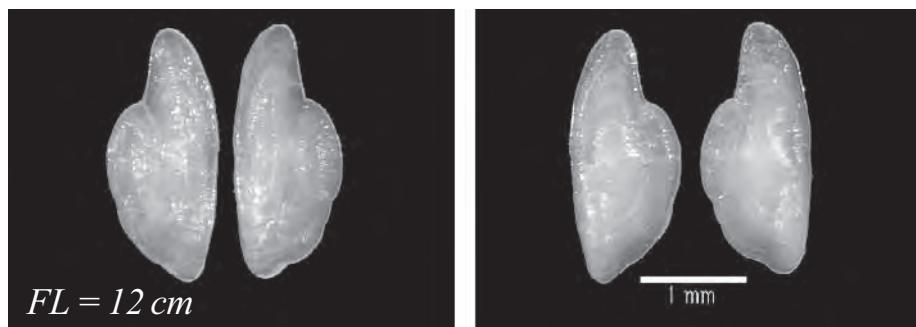
Perciformes

187

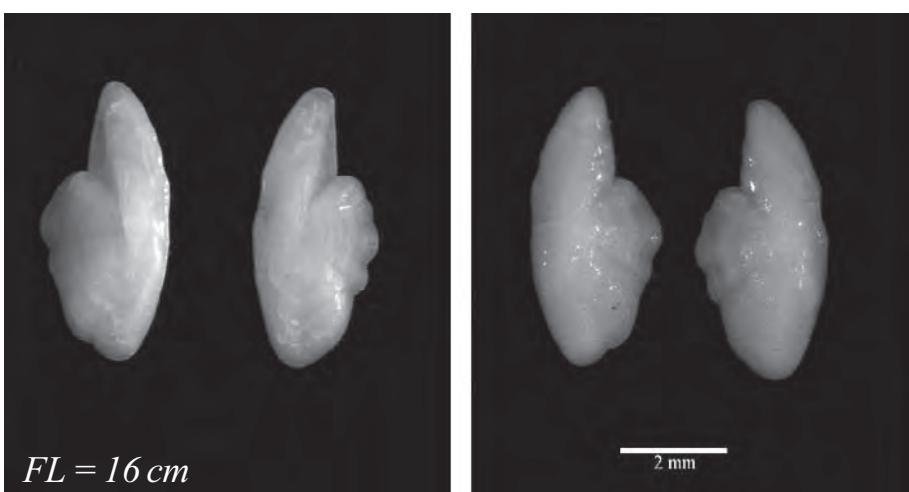
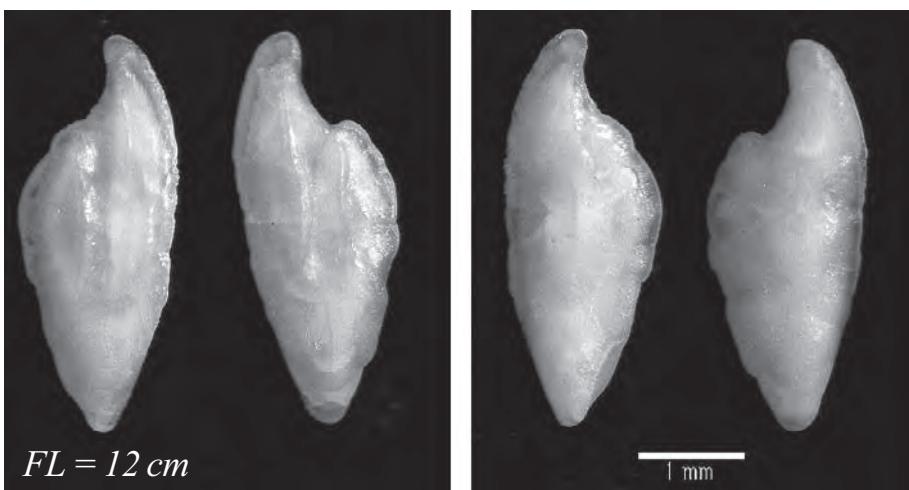
Stichaeidae: *Lumpenus maculatus*



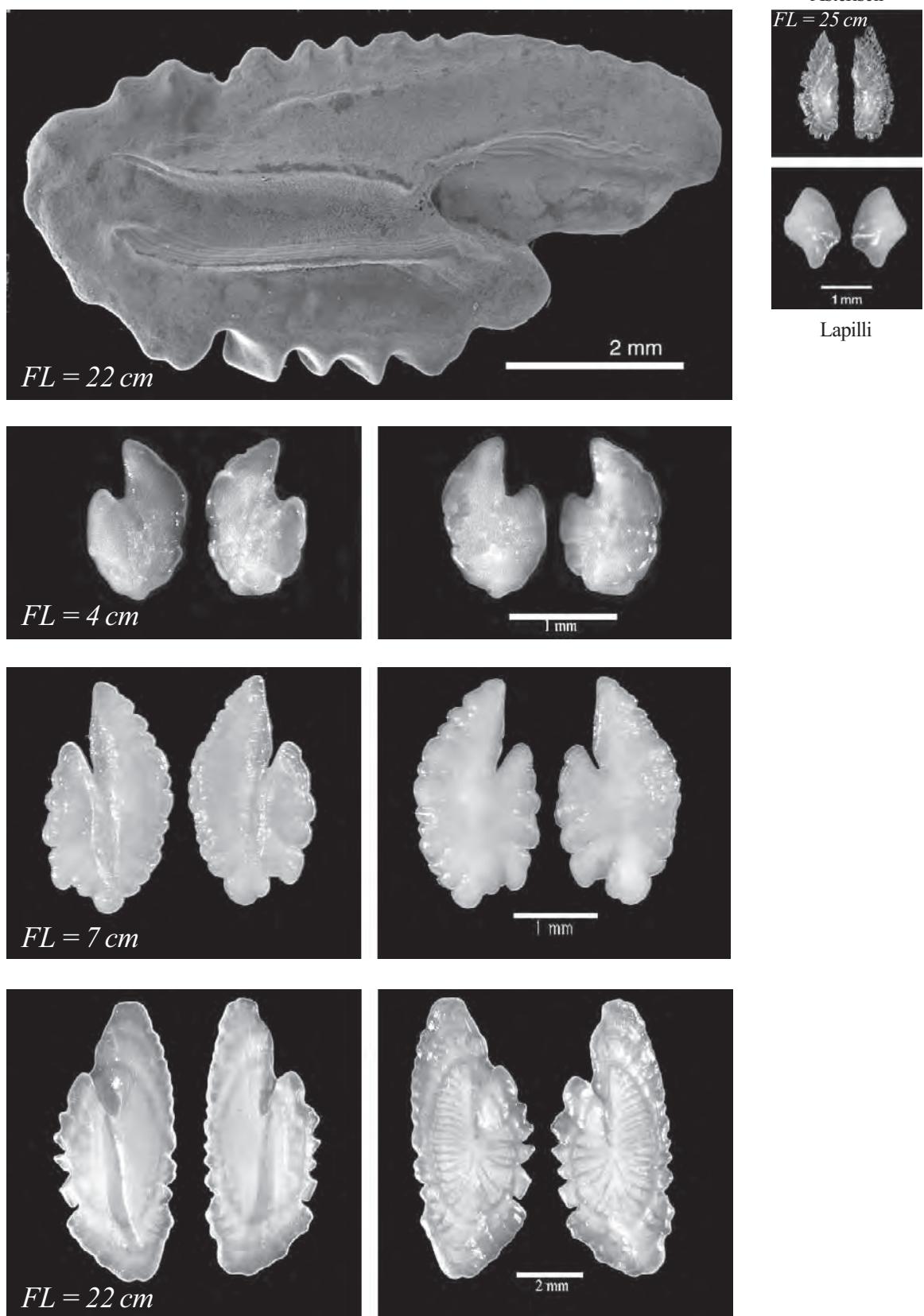
Stichaeidae: *Stichaeus punctatus*



Perciformes

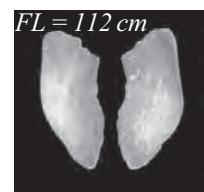
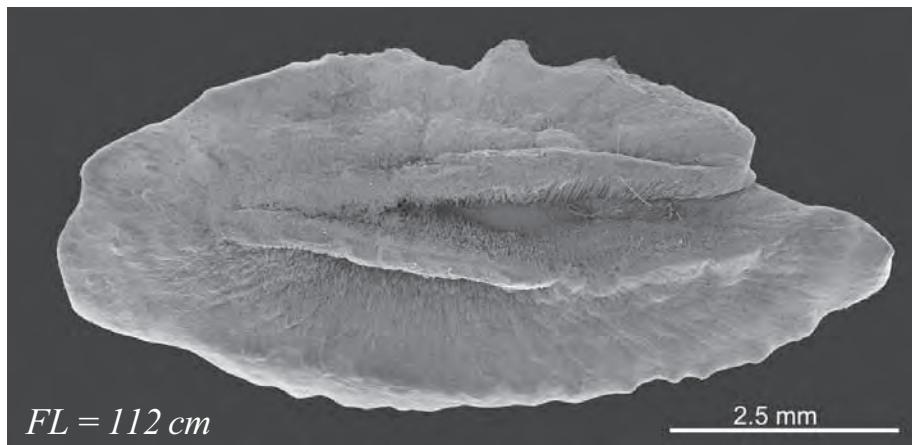
Stichaeidae: *Ulvaria subbifurcata*

Stromateidae: *Peprilus tricanthus*

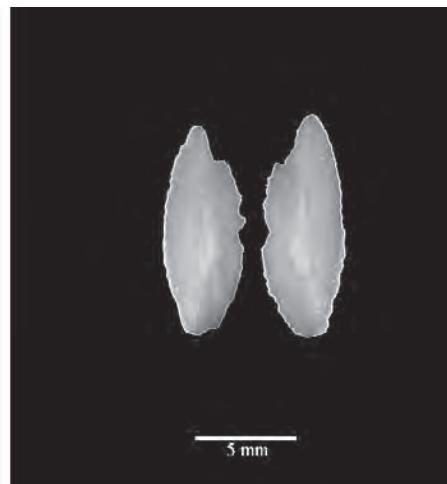


Perciformes

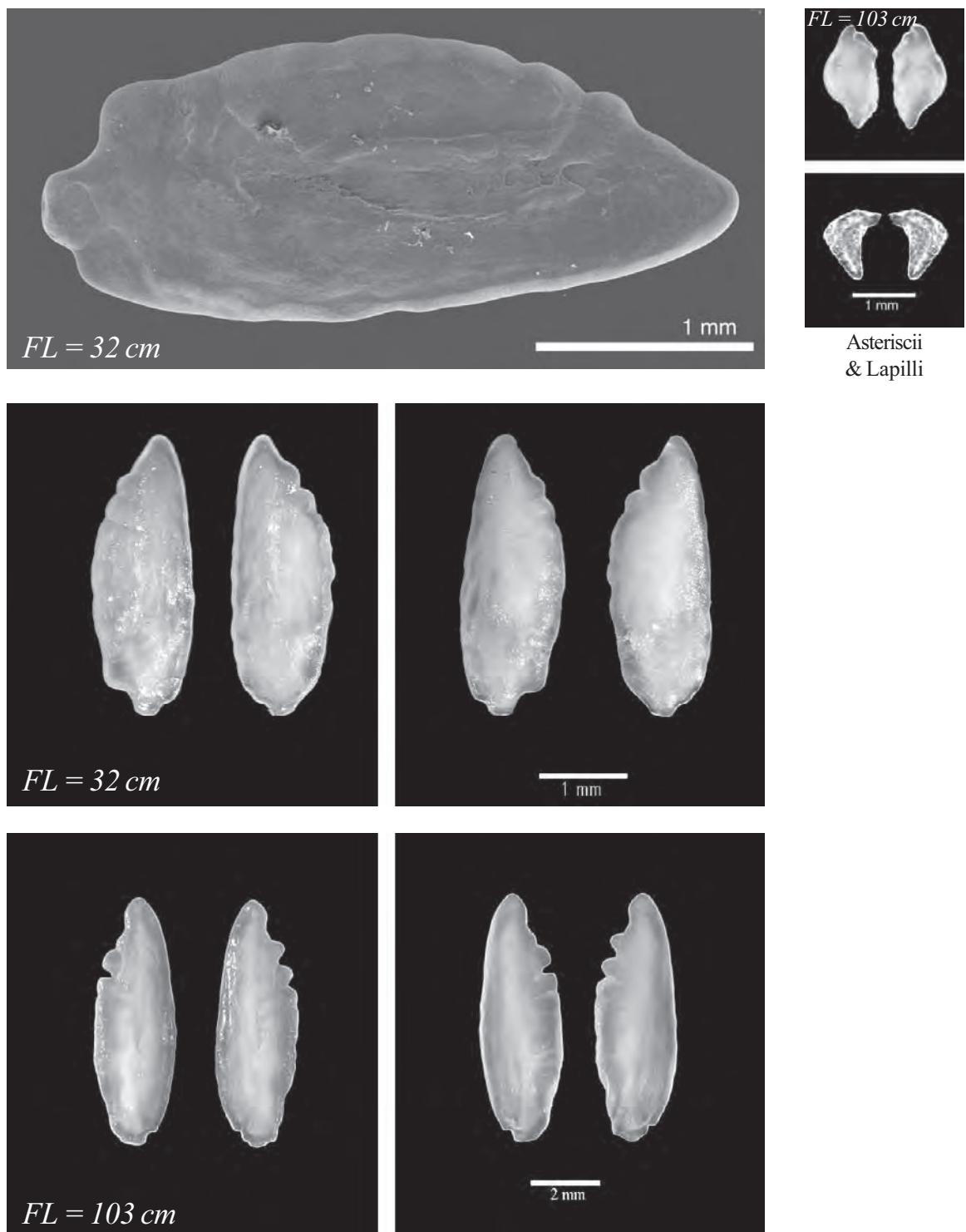
Trichiuridae: *Aphanopus carbo*



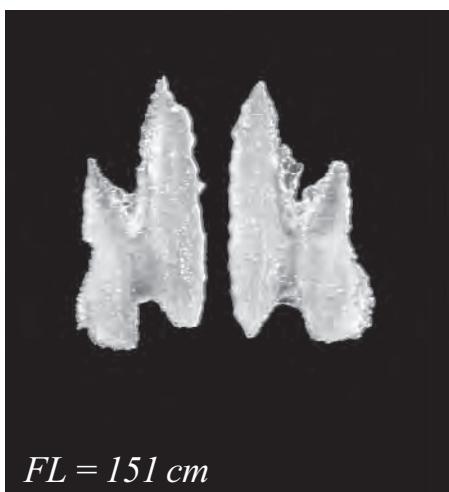
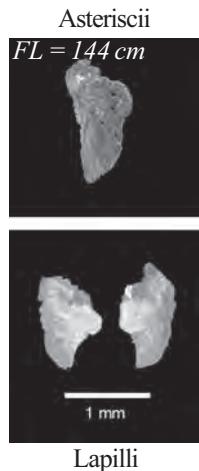
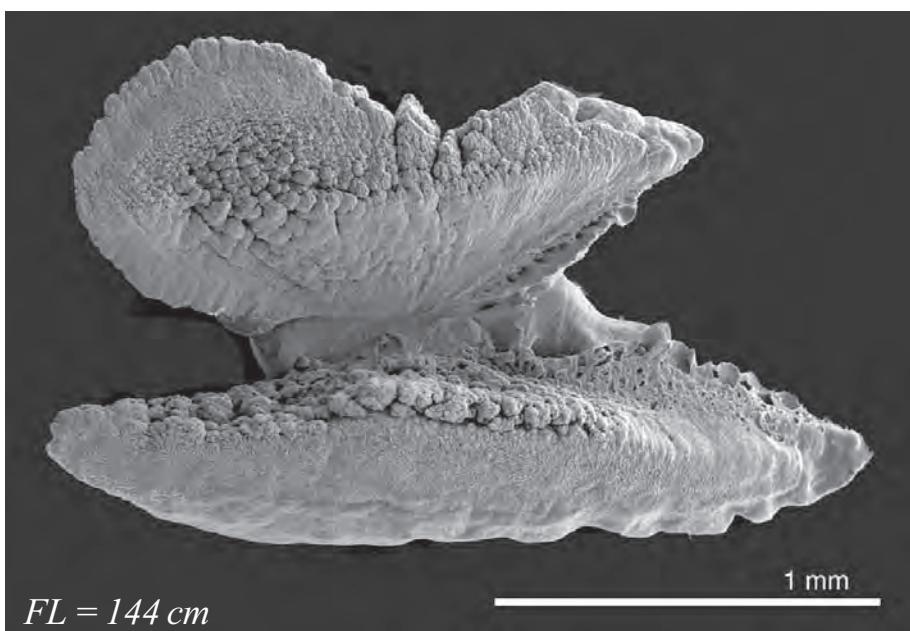
Asteriscii
& Lapilli



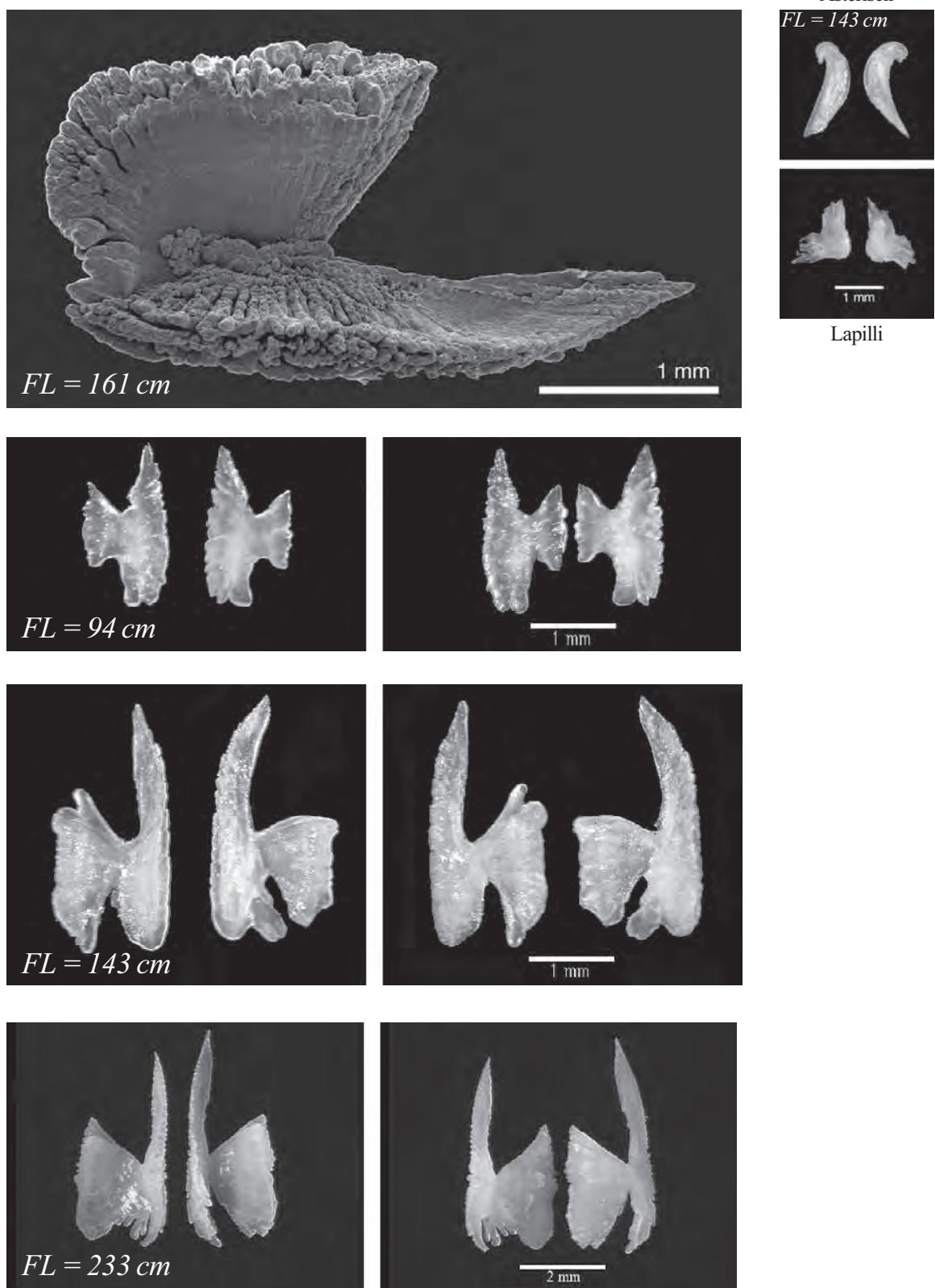
Trichiuridae: *Benthodesmus elongatus*



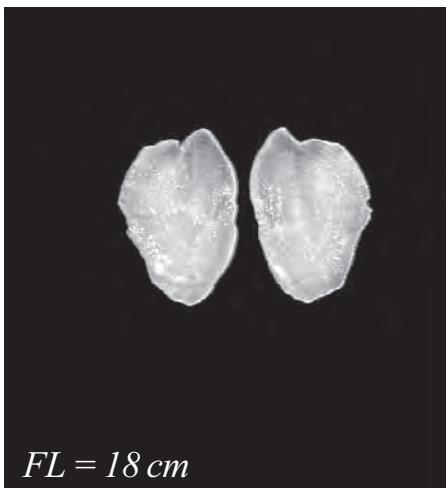
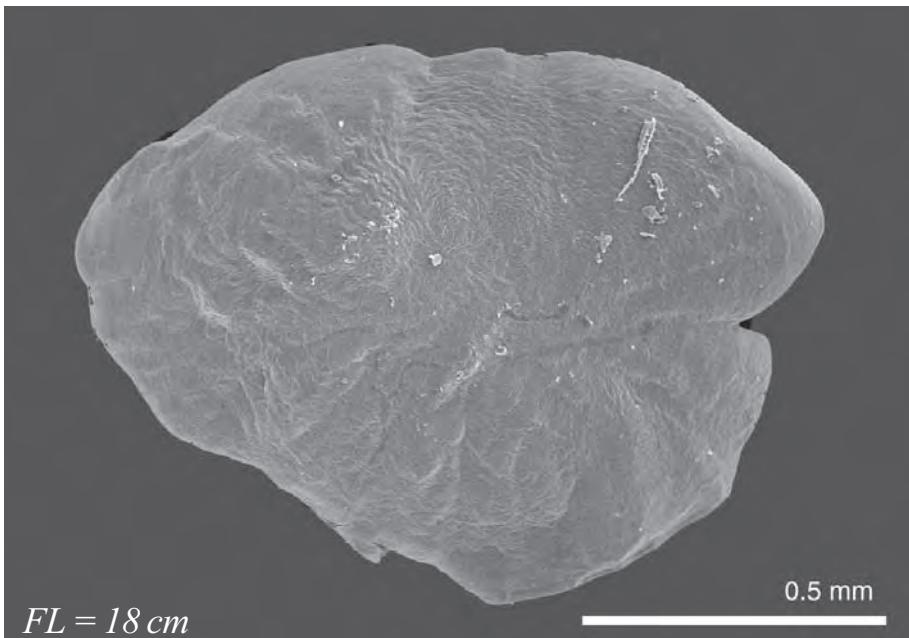
Perciformes

Xiphiidae: *Tetrapurus albidus*

Xiphiidae: *Xiphias gladius*



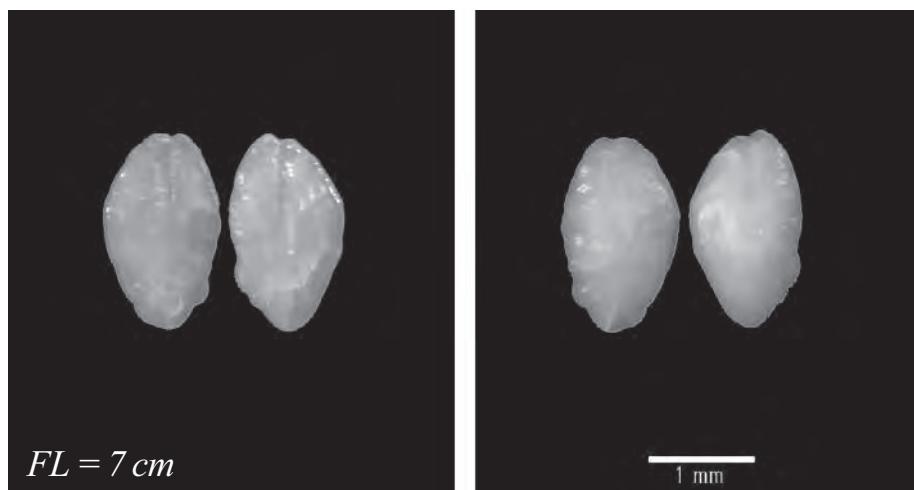
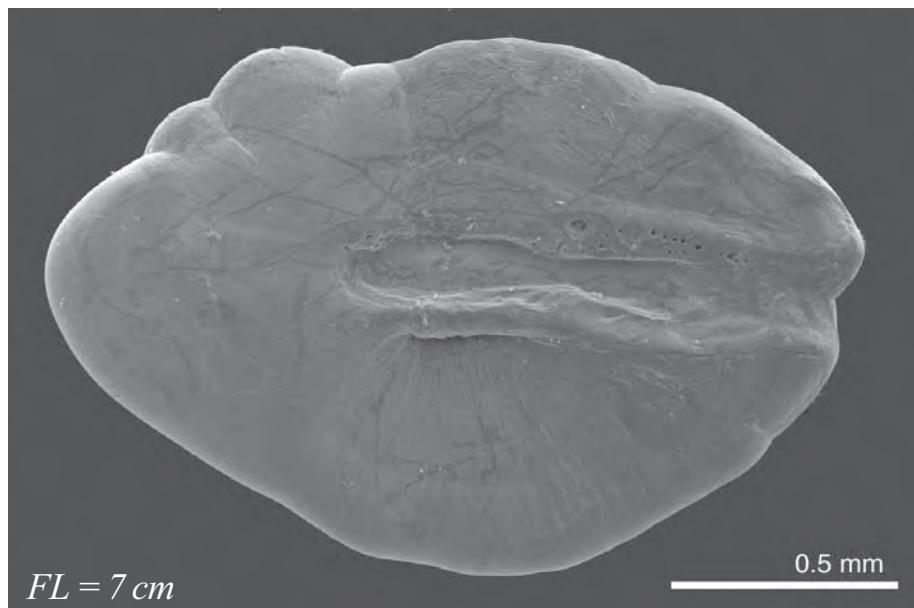
Perciformes
Zoarcidae: *Gymnelus viridis*



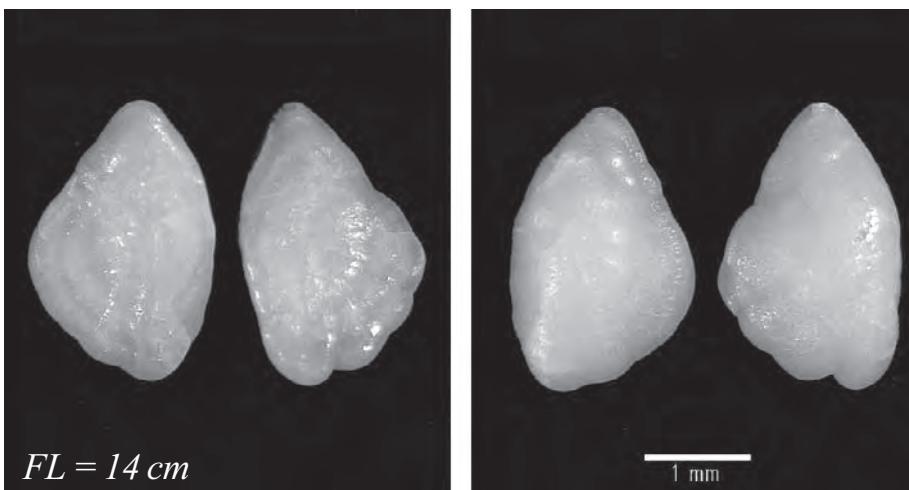
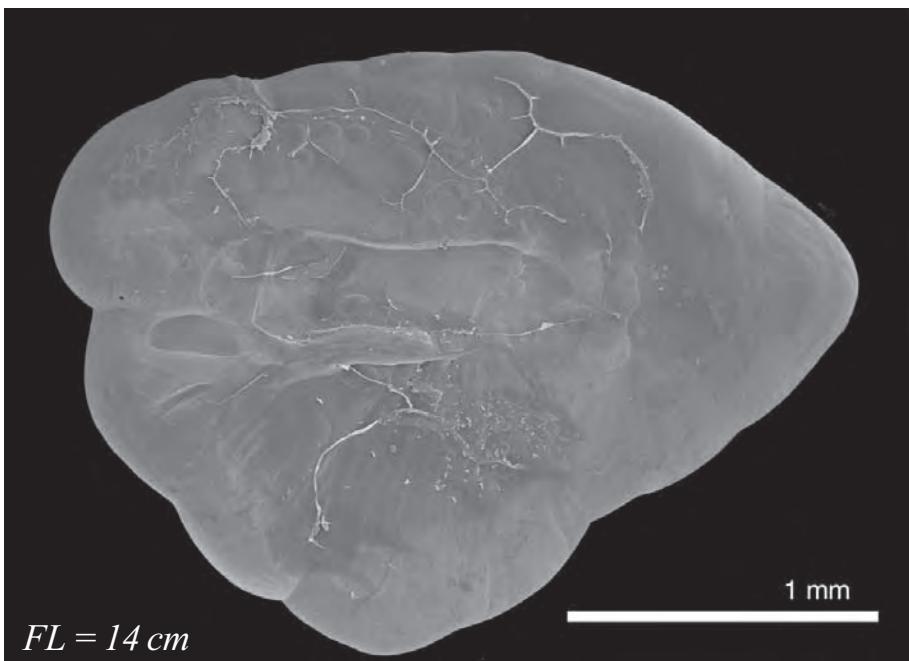
Perciformes

195

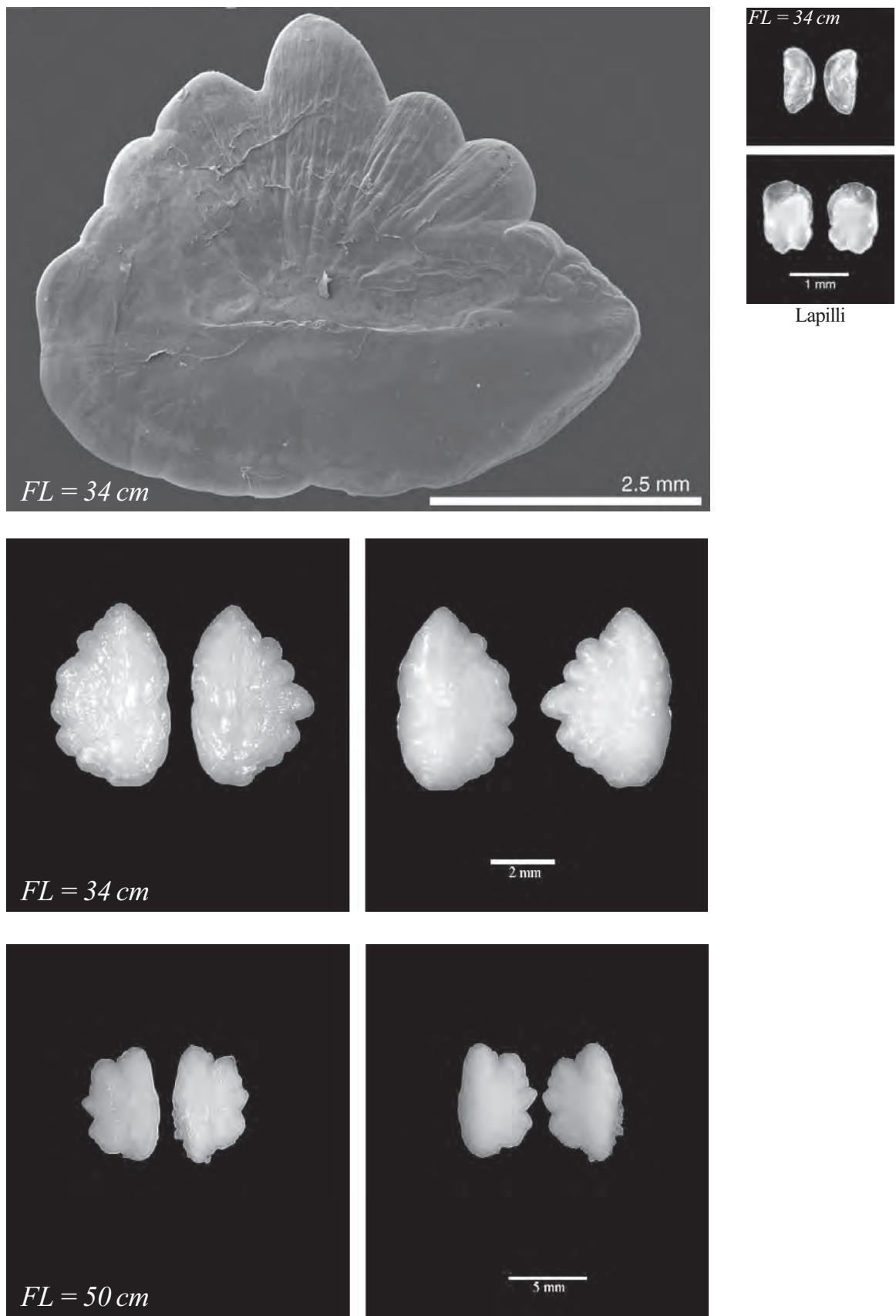
Zoarcidae: *Lycenchelys paxillus*



Perciformes
Zoarcidae: *Lycenchelys verrilli*

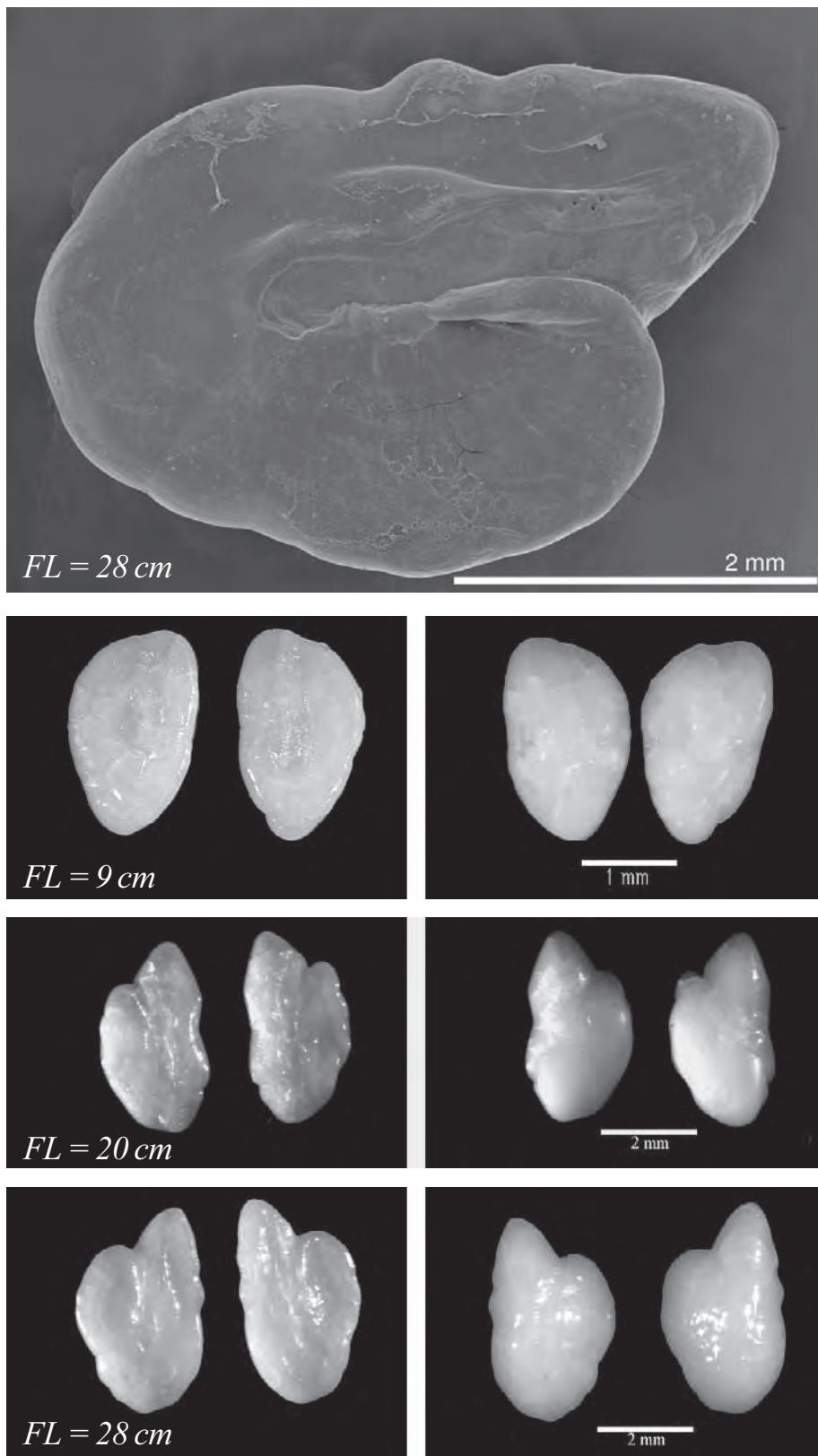


Zoarcidae: *Lycodes esmarki*



Perciformes

Zoarcidae: *Lycodes lavalaei*



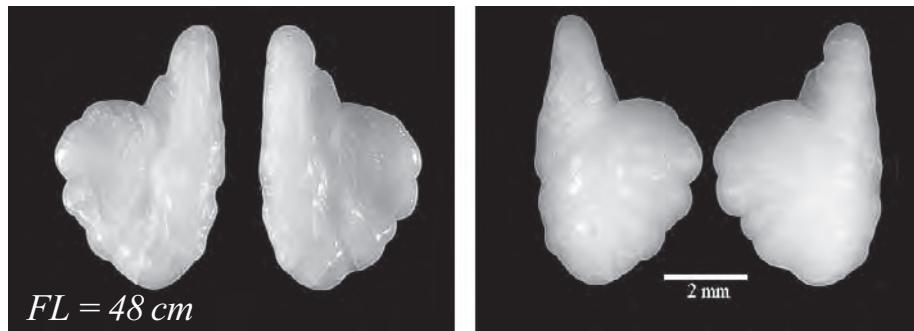
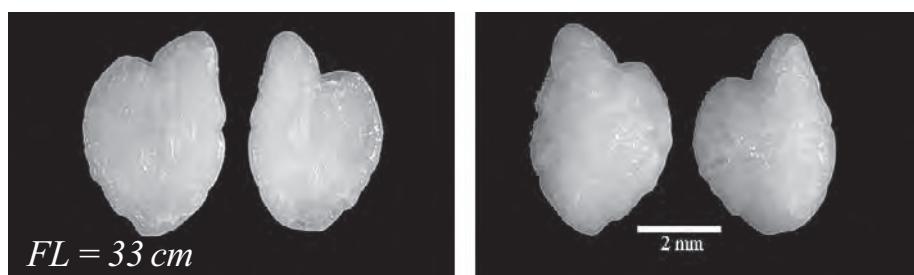
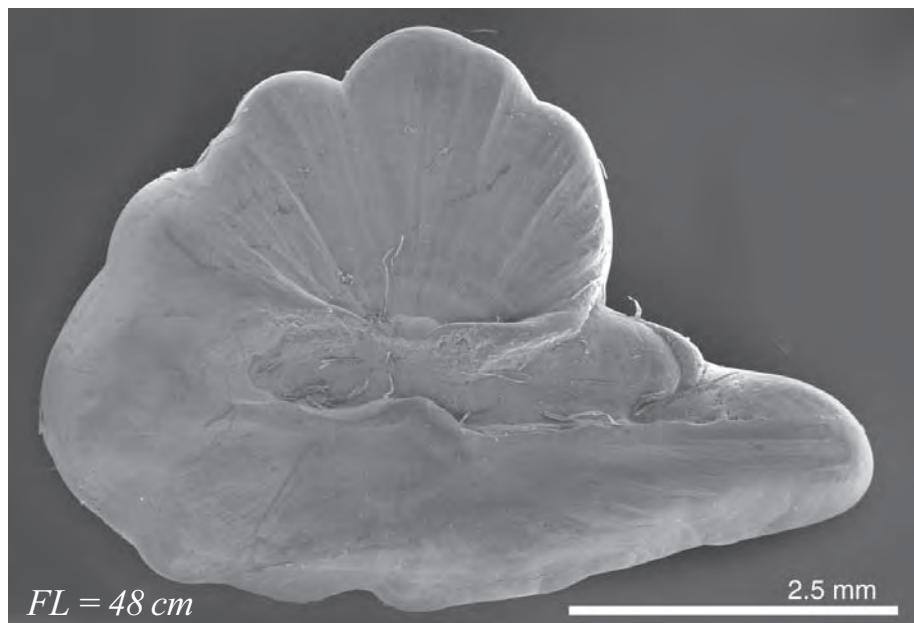
Perciformes

199

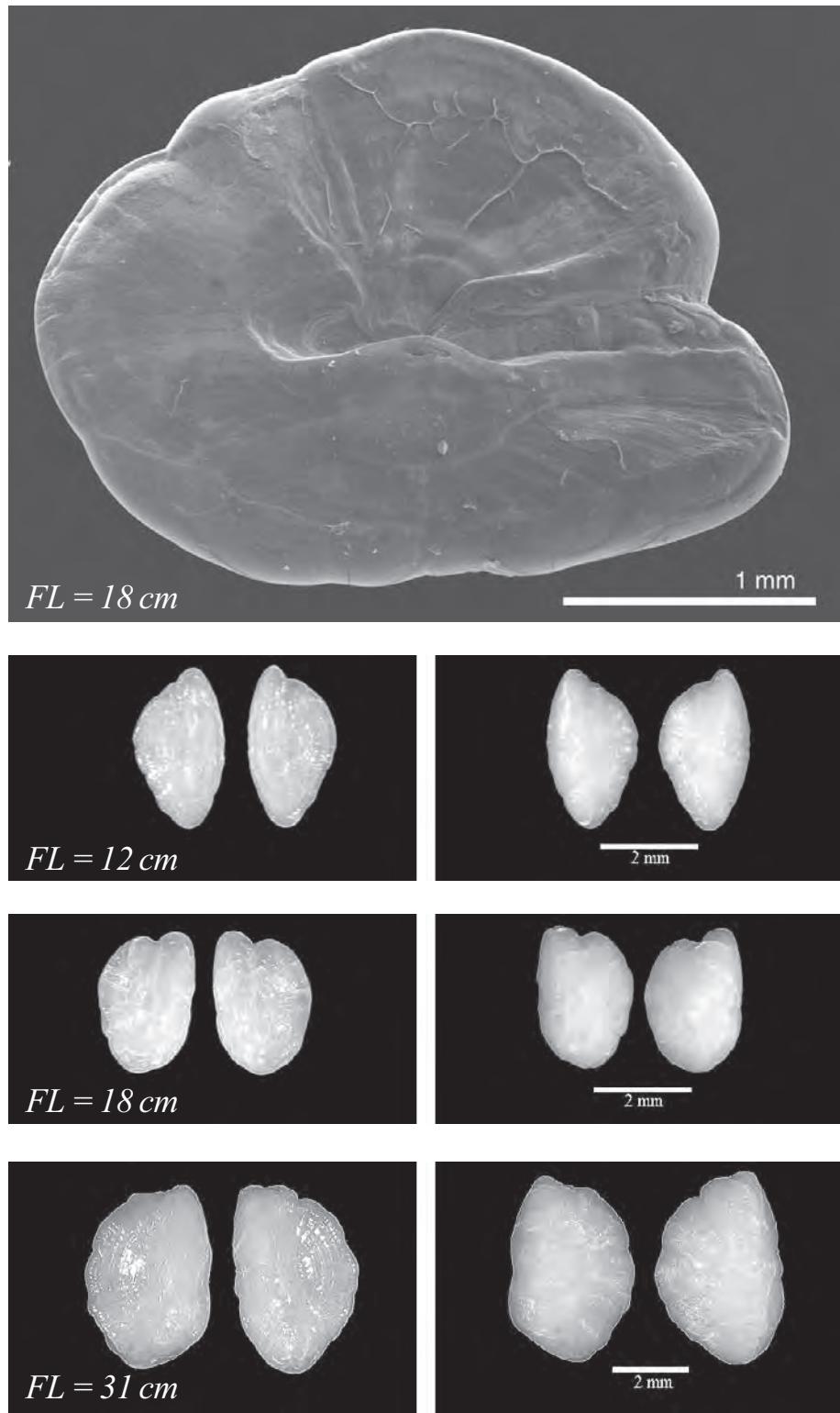
Zoarcidae: *Lycodes pallidus*



Zoarcidae: *Lycodes reticulatus*



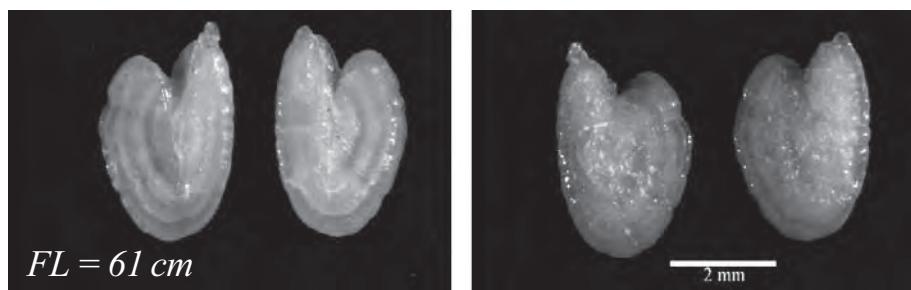
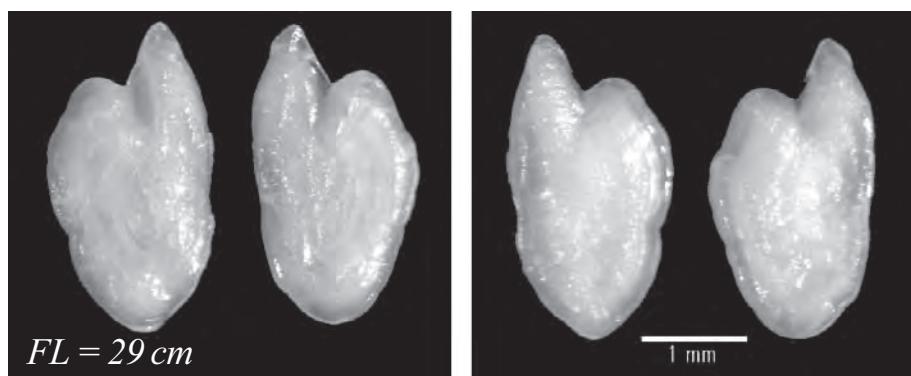
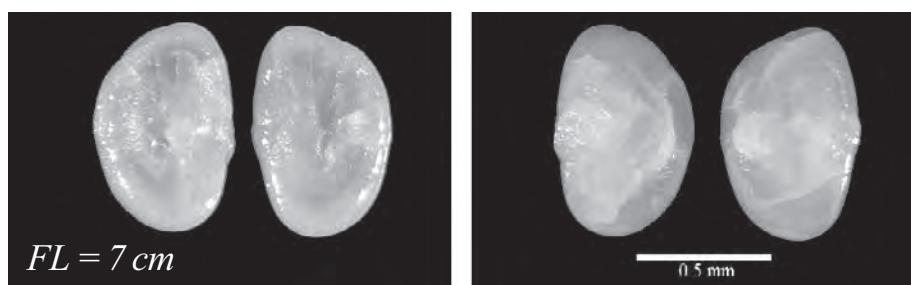
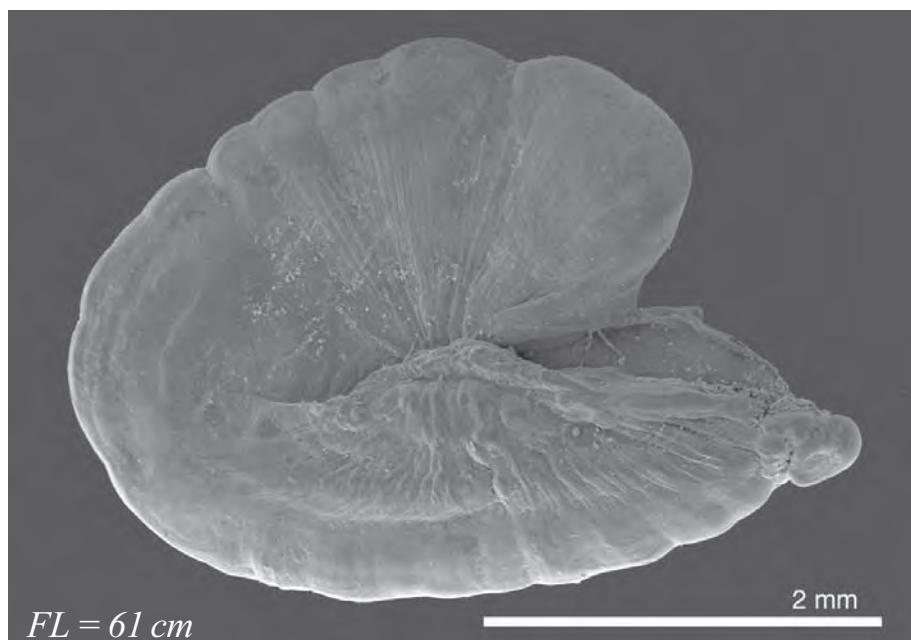
Perciformes
Zoarcidae: *Lycodes vahlii*



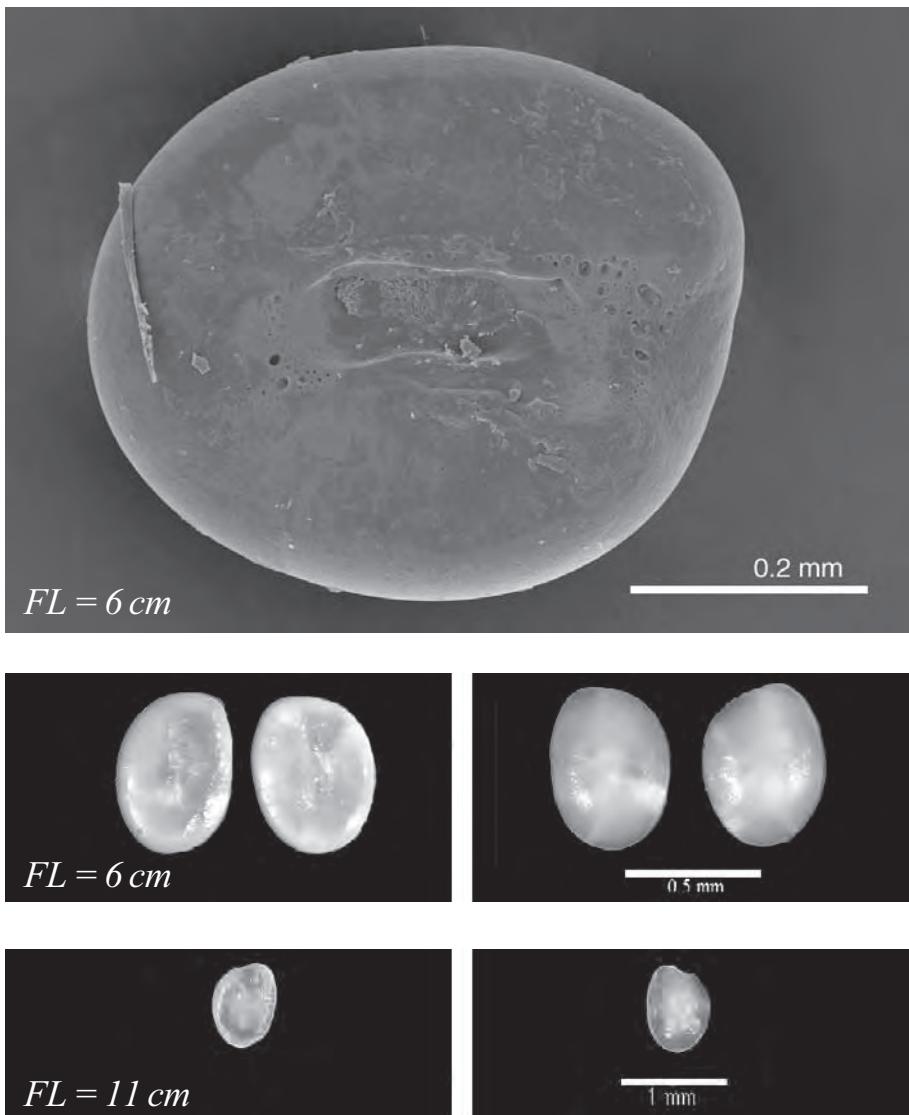
Perciformes

201

Zoarcidae: *Macrozoarces americanus*



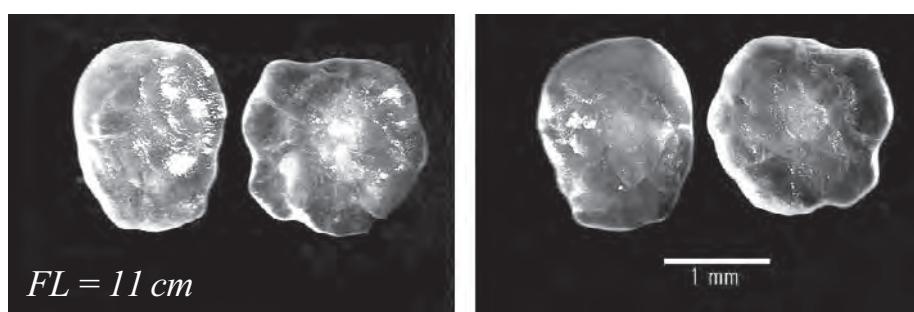
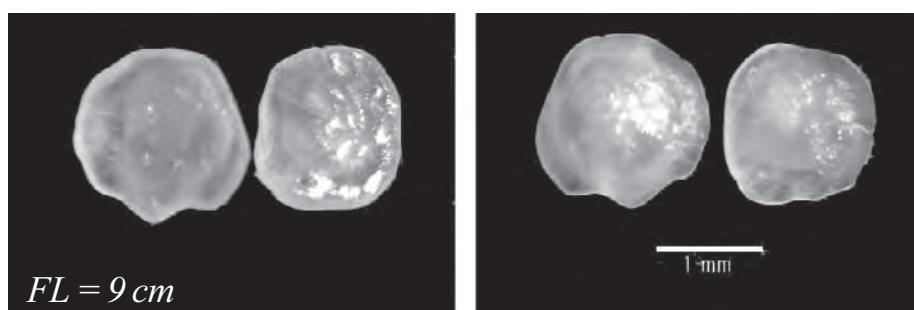
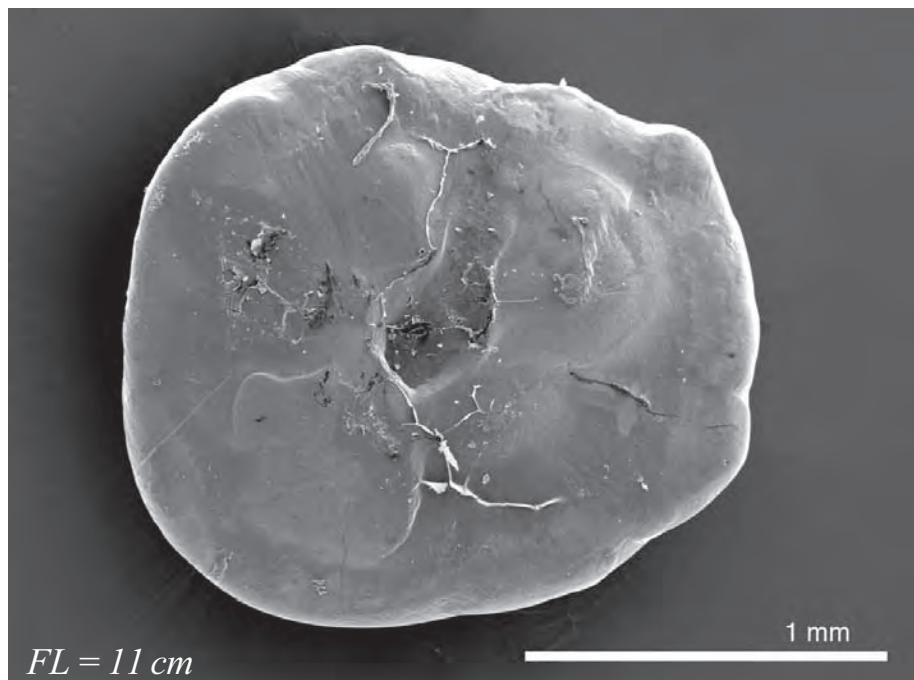
Perciformes
Zoarcidae: *Melanostigma atlanticum*



Pleuronectiformes

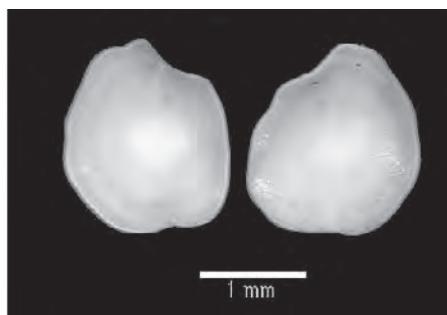
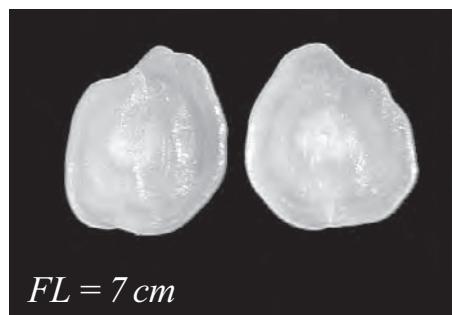
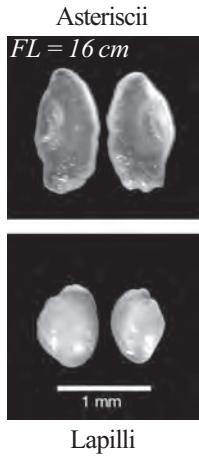
203

Cynoglossidae: *Syphurus pterospilotus*



Pleuronectiformes

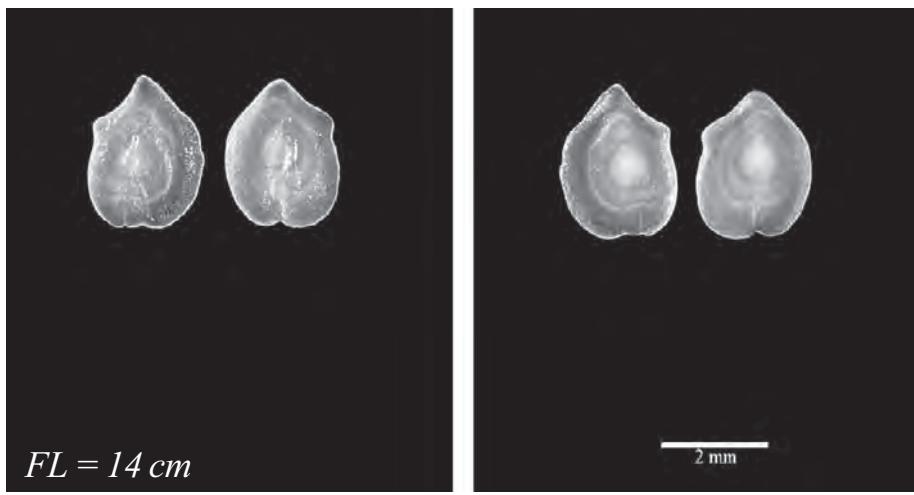
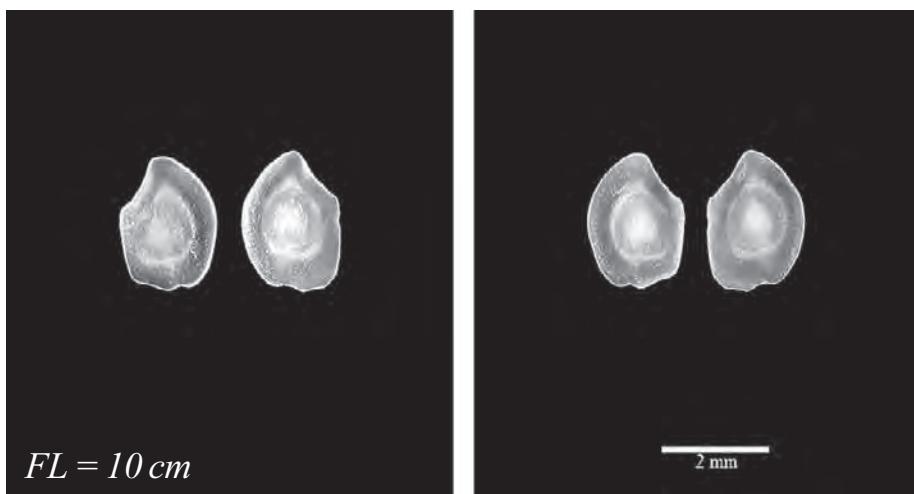
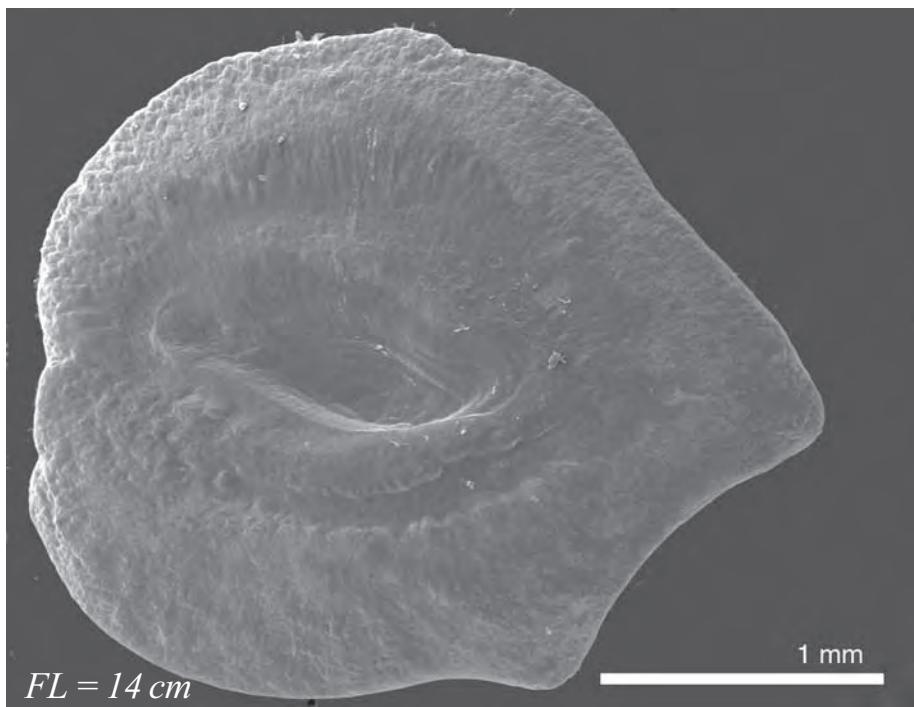
Paralichthyidae: *Citharichthys arctifrons*



Pleuronectiformes

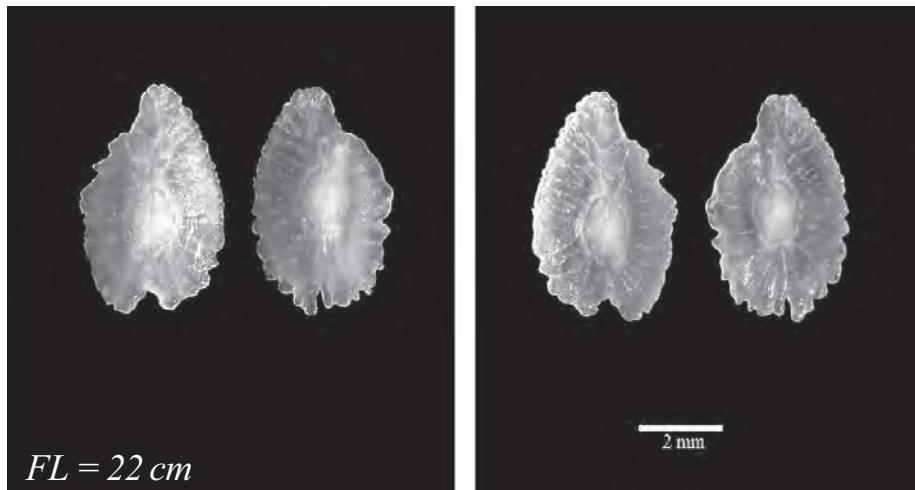
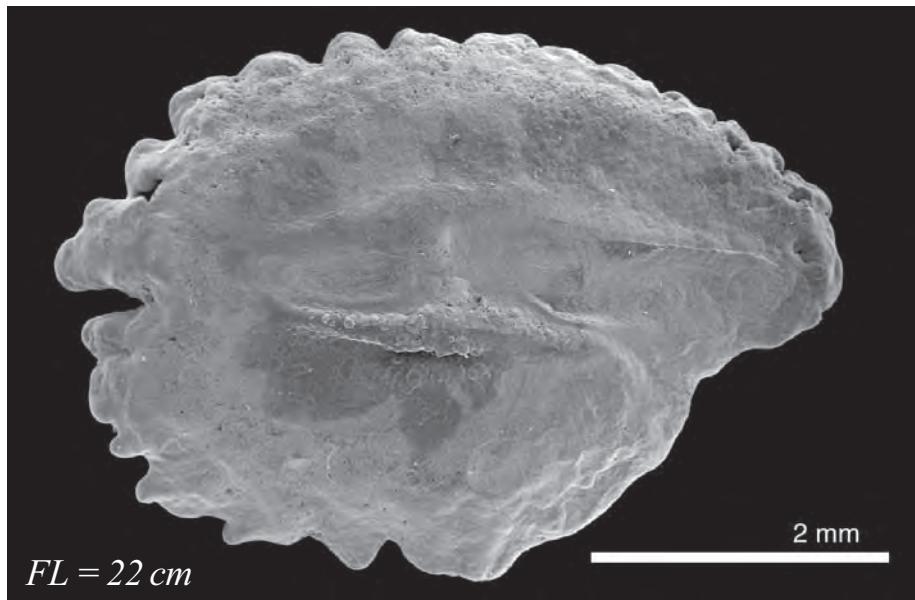
205

Paralichthyidae: *Etropus microstomus*



Pleuronectiformes

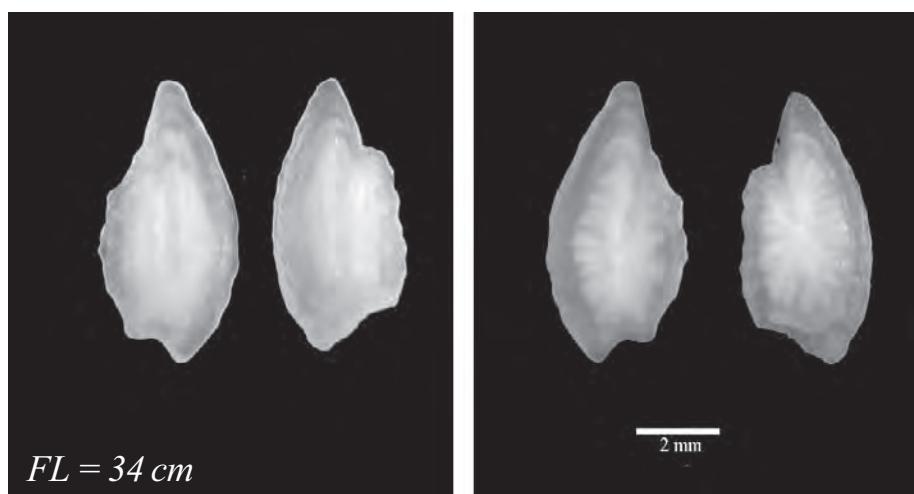
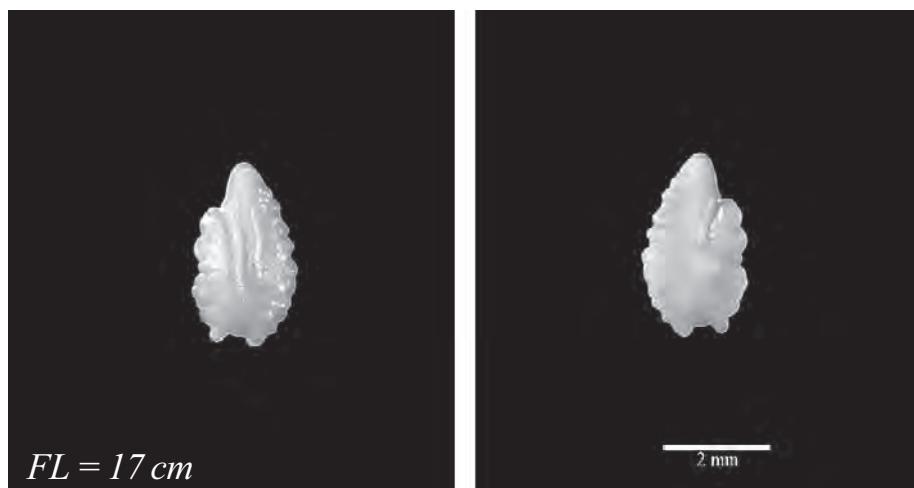
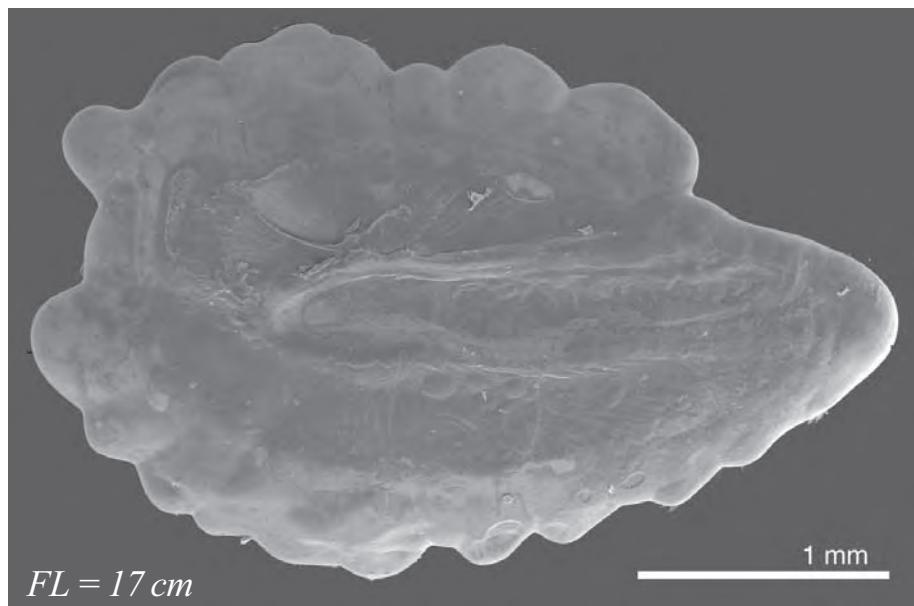
Paralichthyidae: *Paralichthys dentatus*



Pleuronectiformes

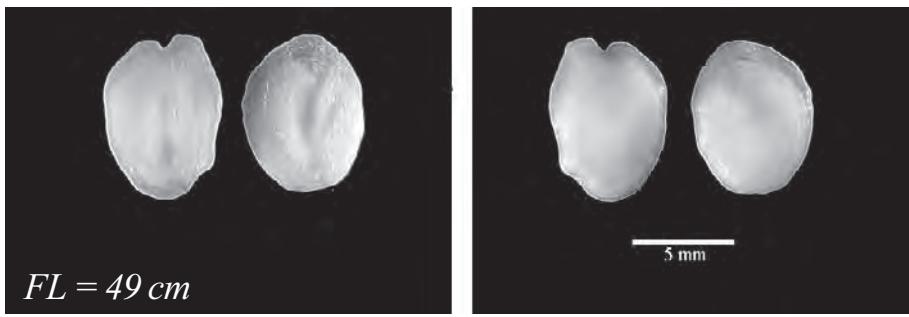
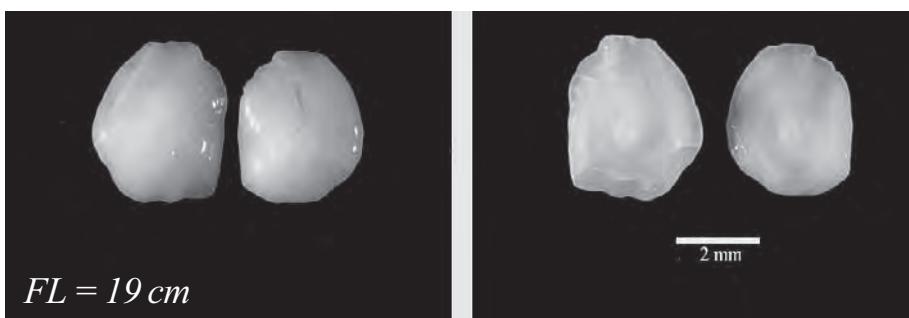
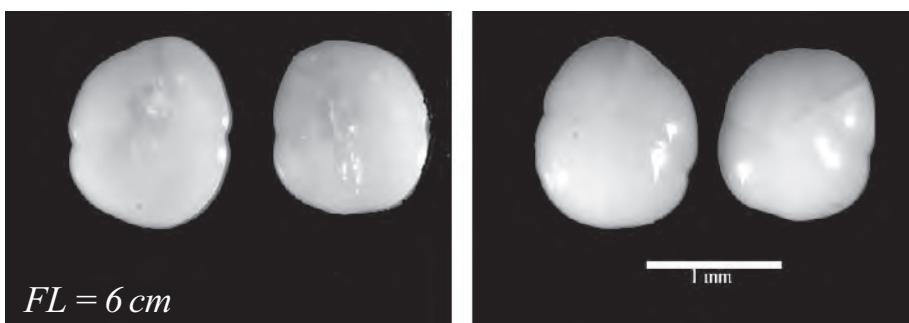
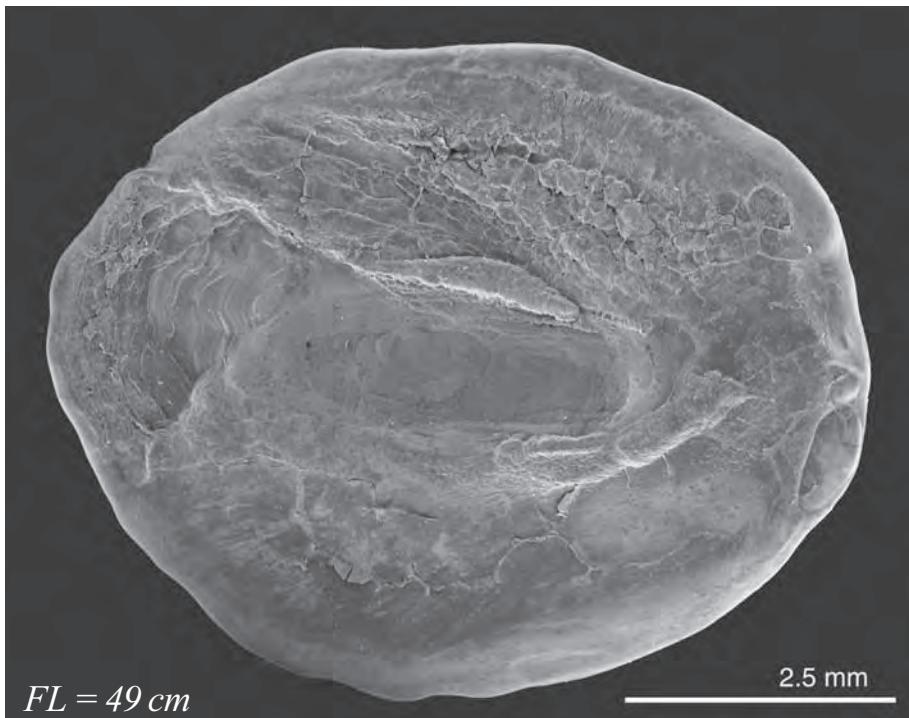
207

Paralichthyidae: *Paralichthys oblongus*



Pleuronectiformes

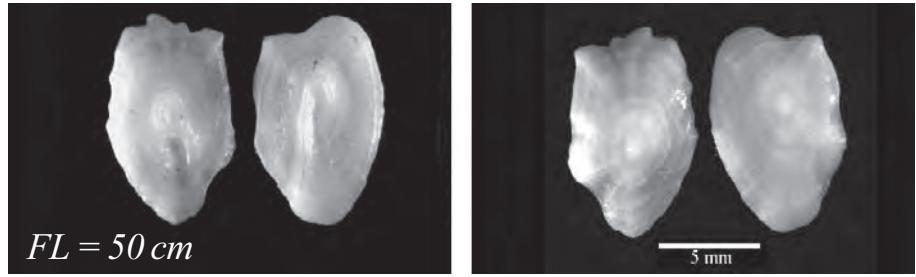
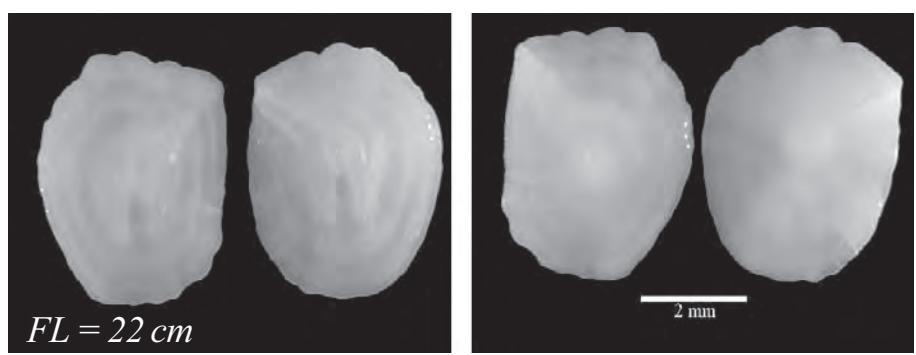
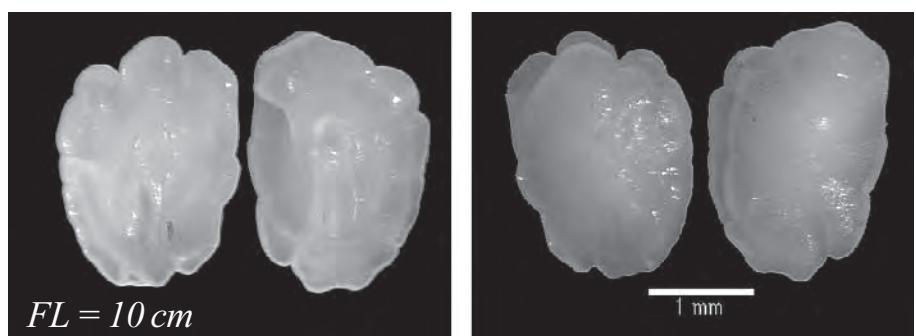
Pleuronectidae: *Glyptocephalus cynoglossus*



Pleuronectiformes

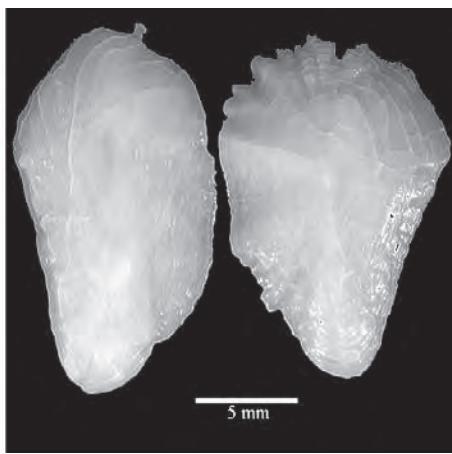
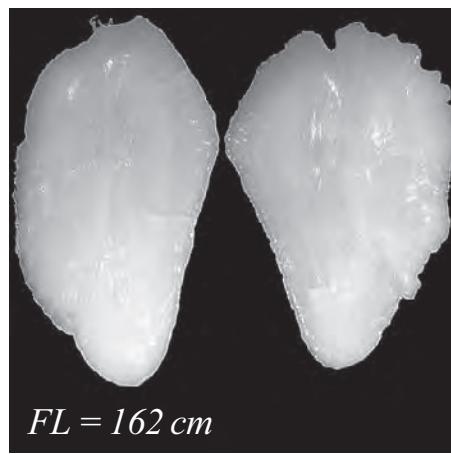
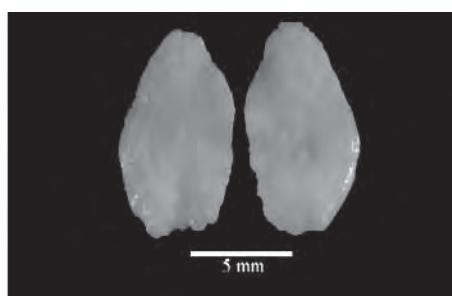
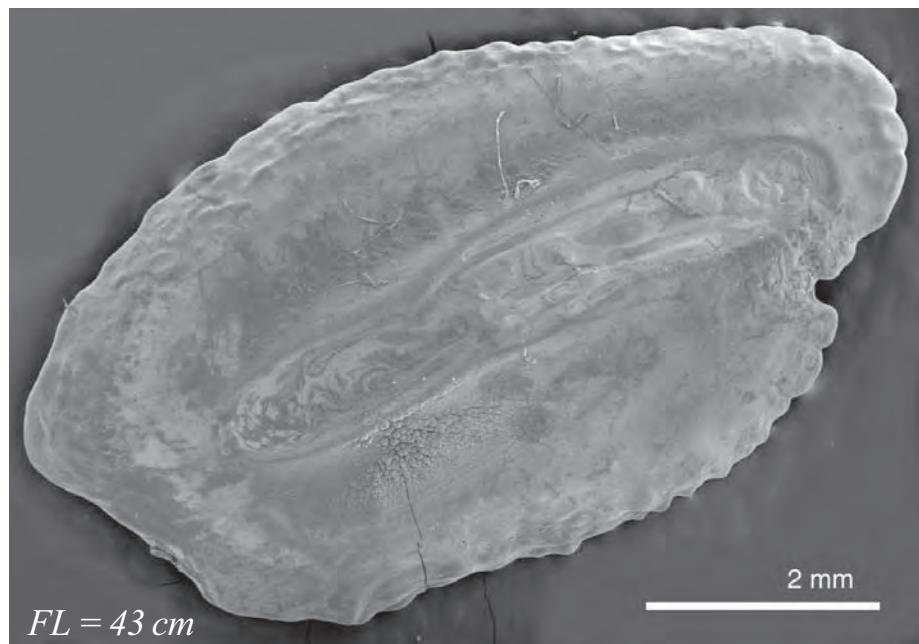
209

Pleuronectidae: *Hippoglossoides platessoides*



Pleuronectiformes

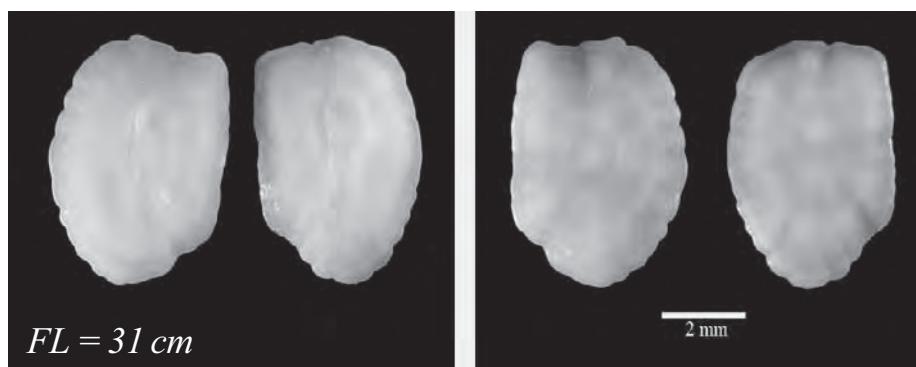
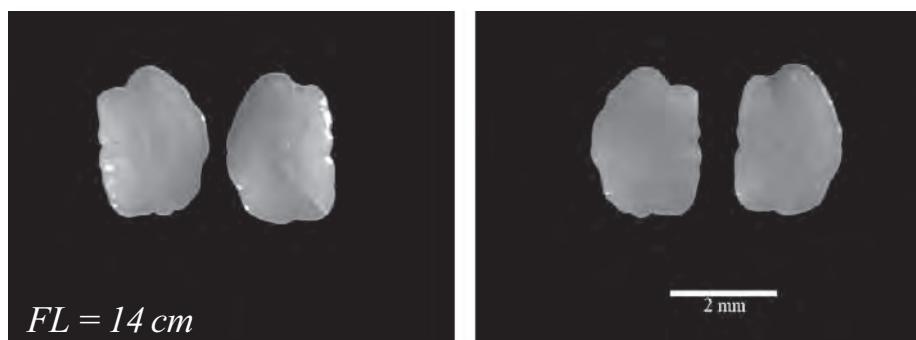
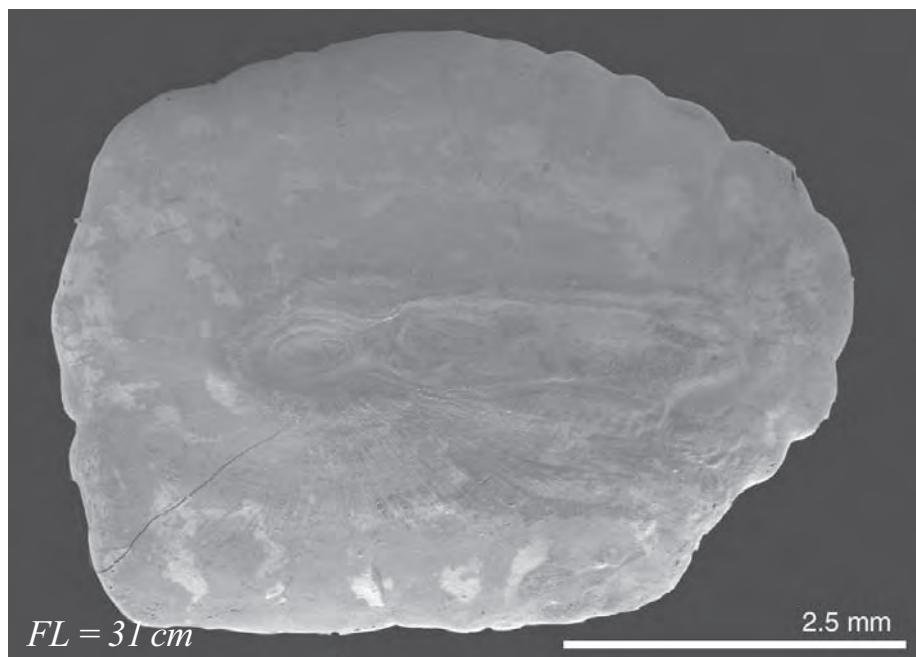
Pleuronectidae: *Hippoglossus hippoglossus*



Pleuronectiformes

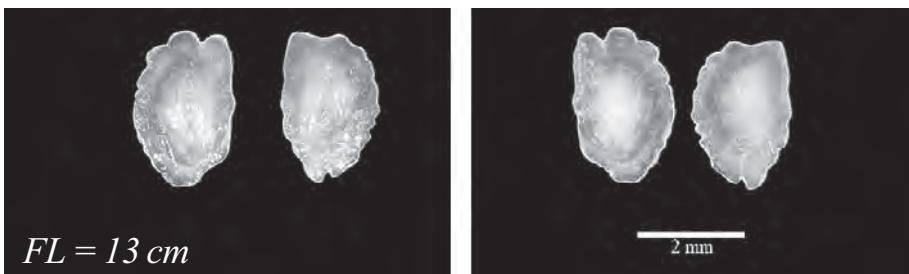
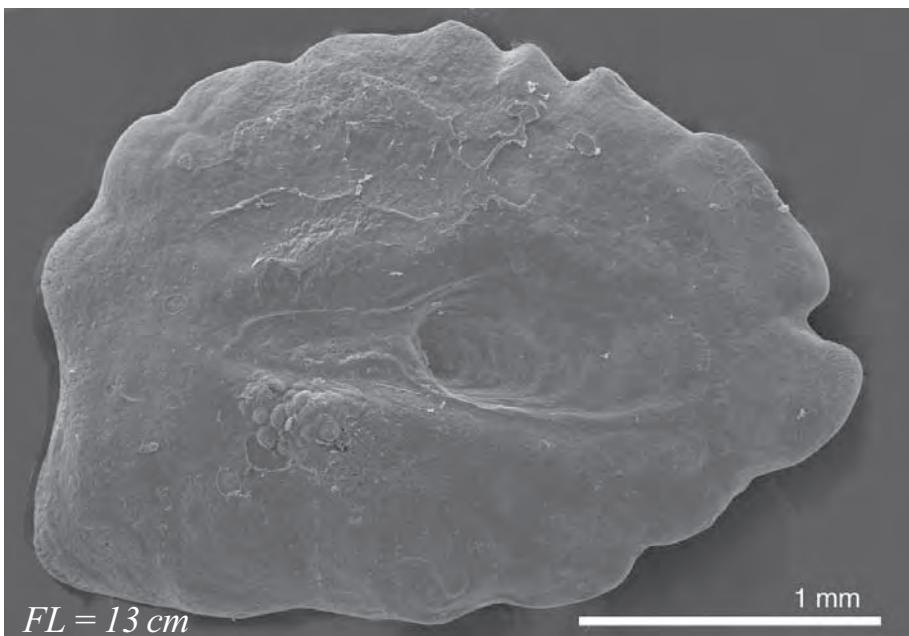
211

Pleuronectidae: *Limanda ferruginea*



Pleuronectiformes

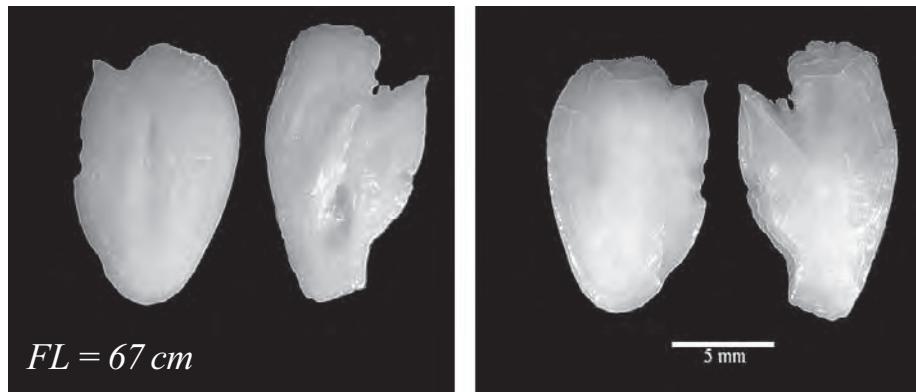
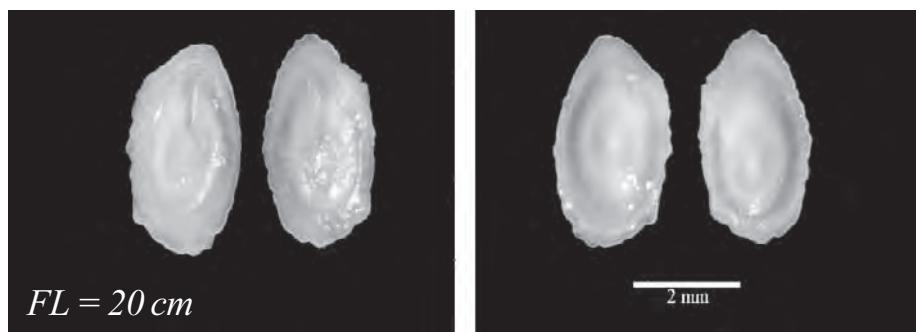
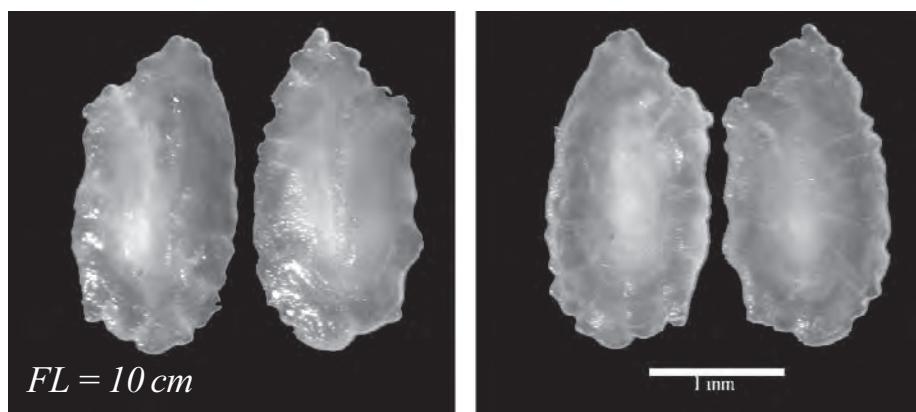
Pleuronectidae: *Liopsetta putnami*



Pleuronectiformes

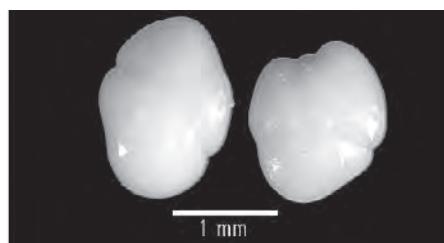
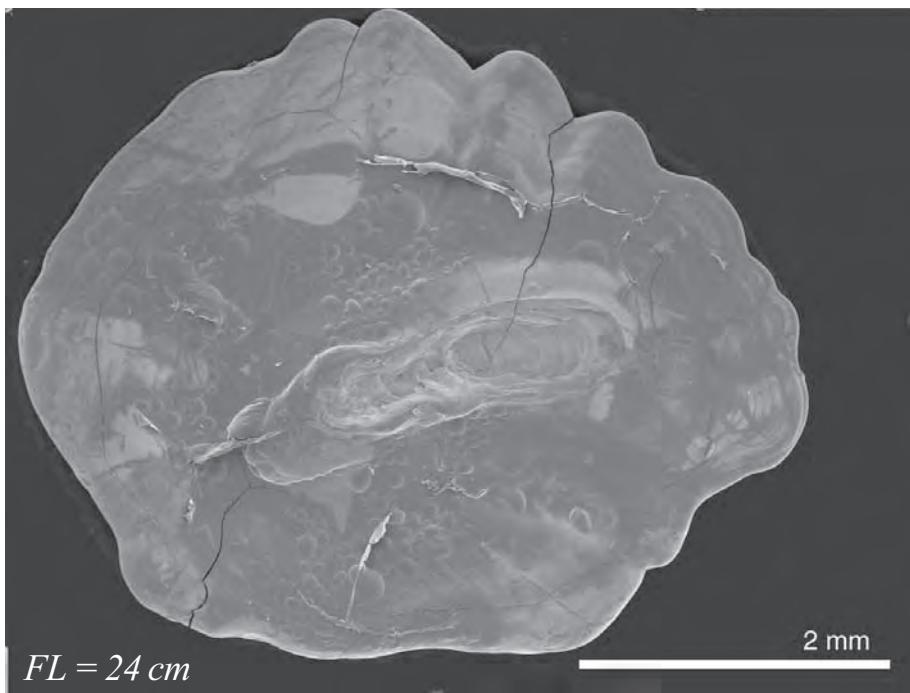
213

Pleuronectidae: *Pseudopleuronectes americanus*



Pleuronectiformes

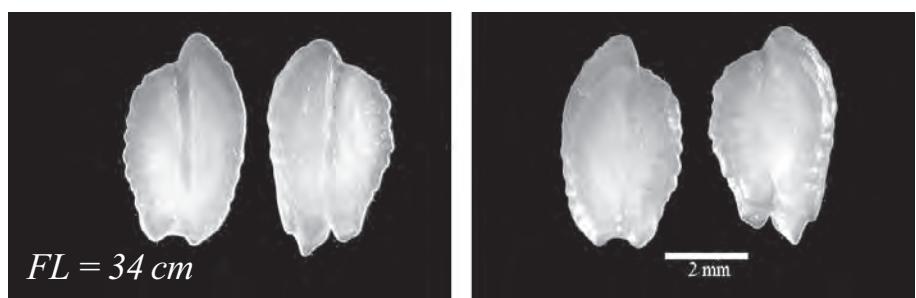
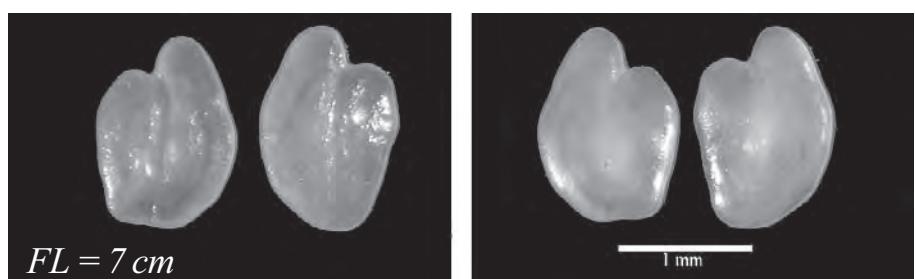
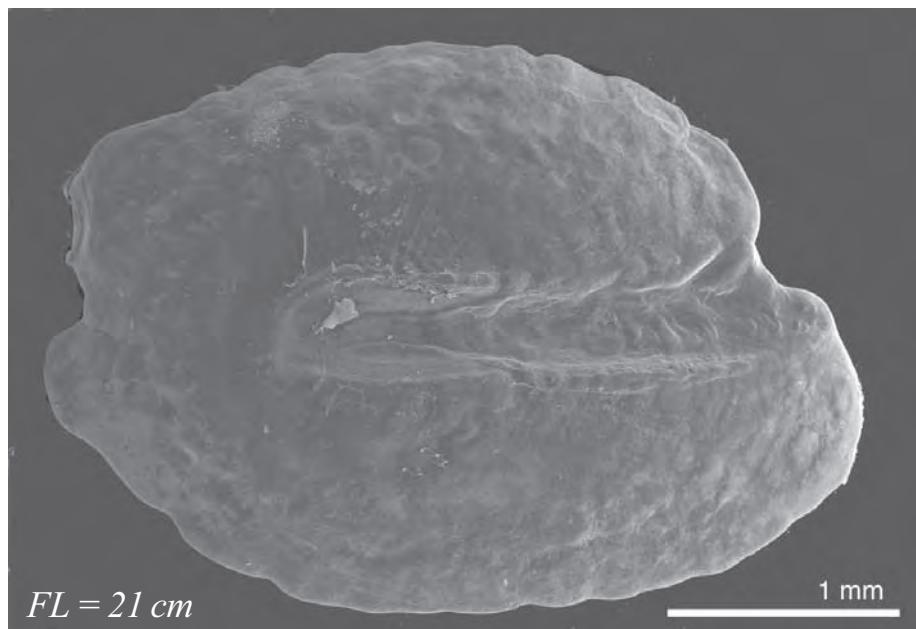
Pleuronectidae: *Reinhardtius hippoglossoides*



Pleuronectiformes

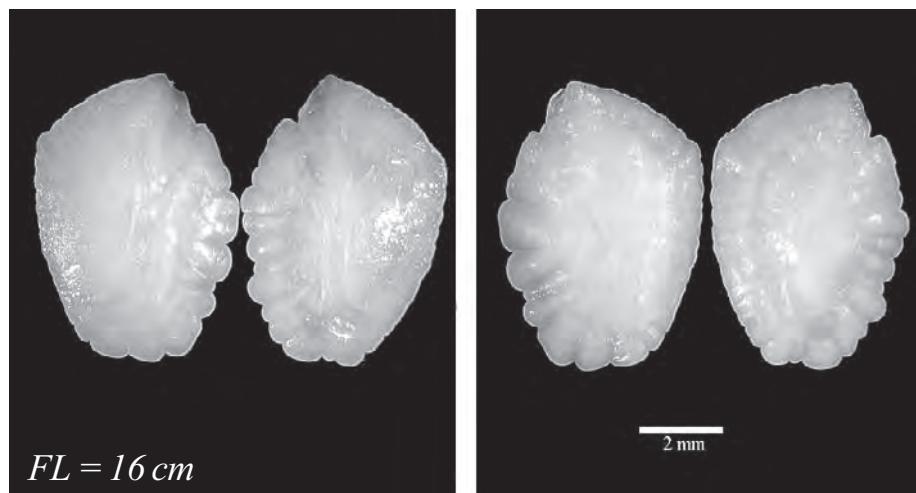
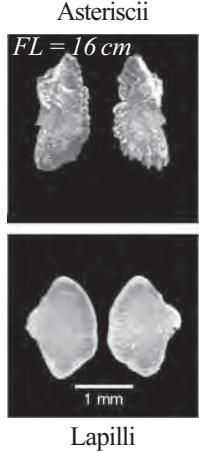
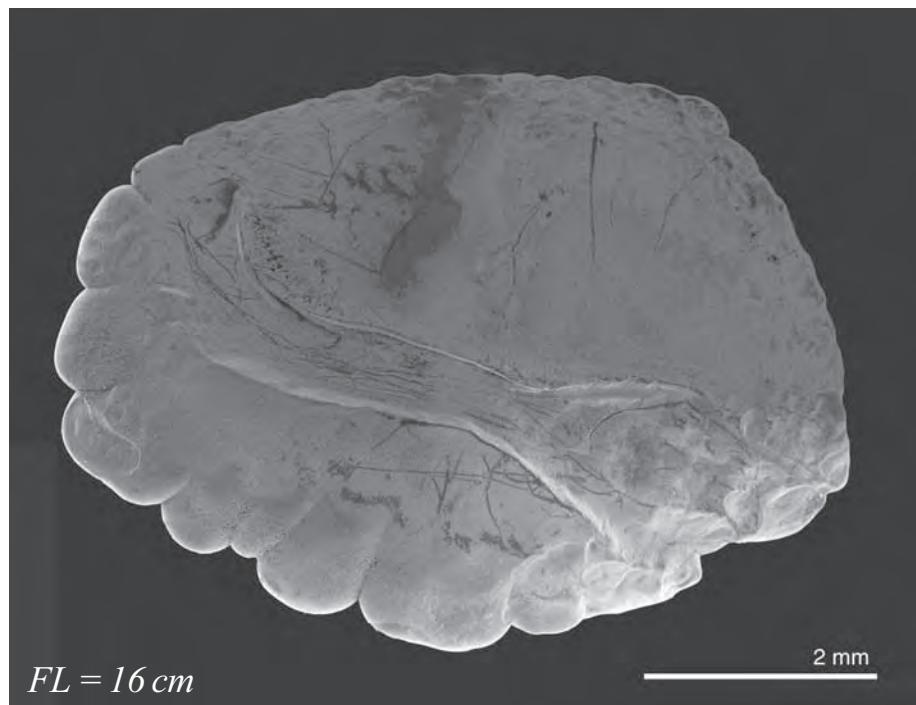
215

Scophthalmidae: *Scophthalmus aquosus*



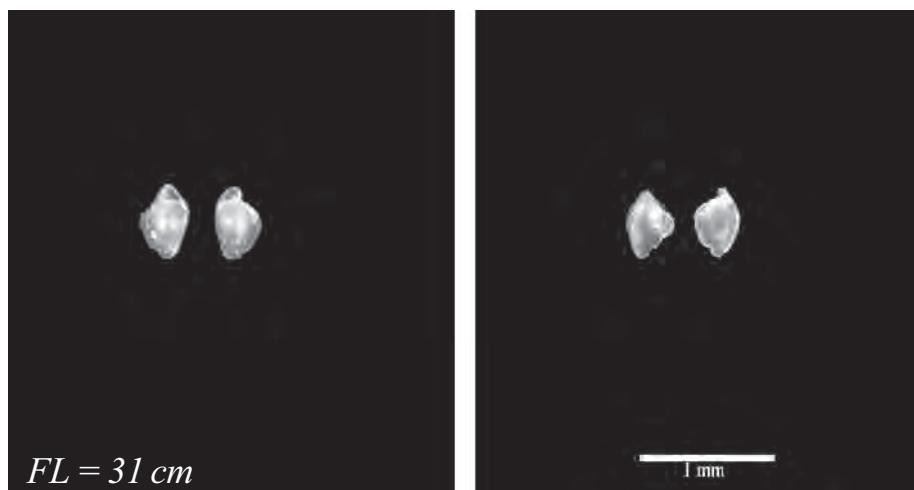
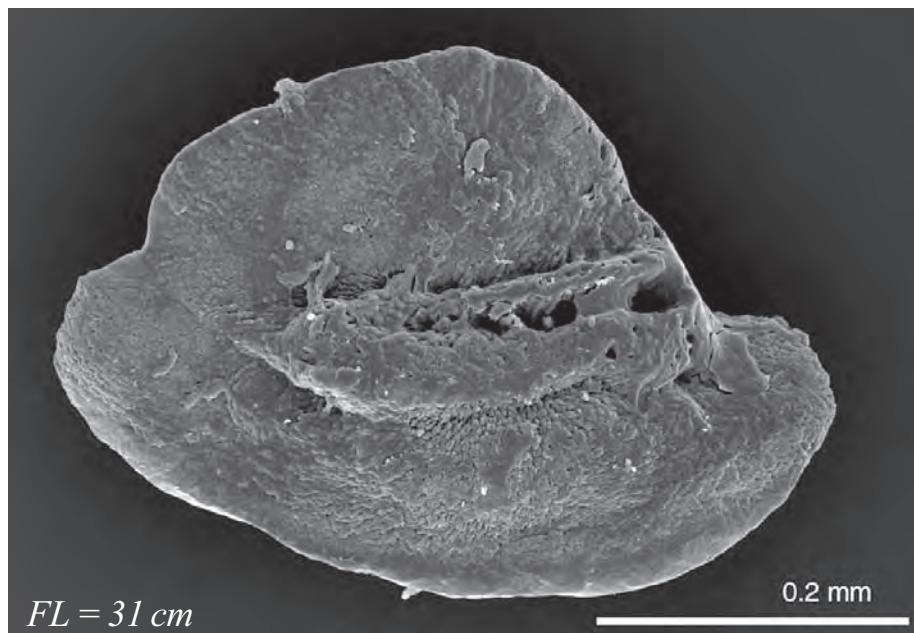
Polymixiiformes

Polymixiidae: *Polymixia lowei*



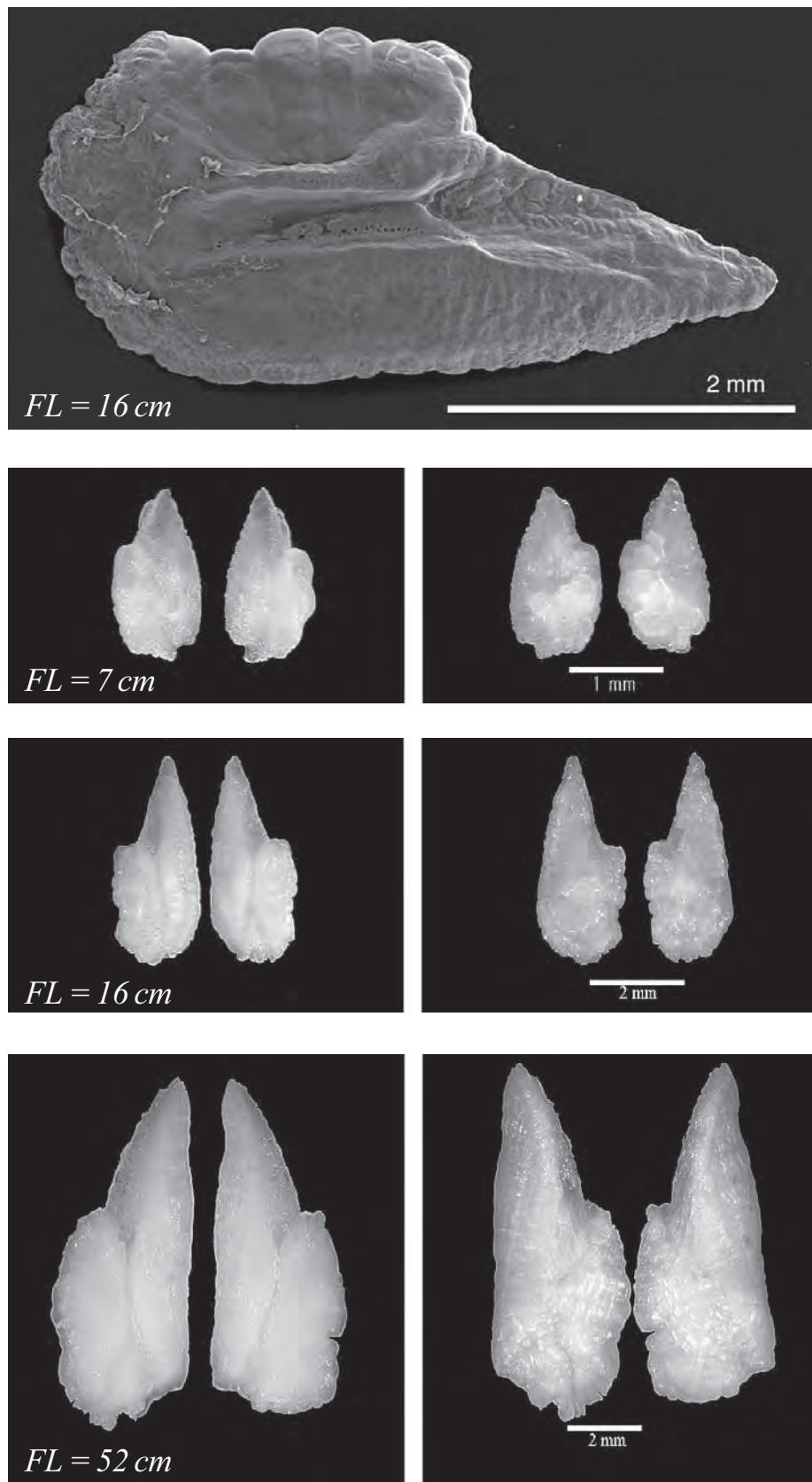
Saccopharyngiformes
Eurypharyngidae: *Eurypharynx pelecanoides*

217



Salmoniformes

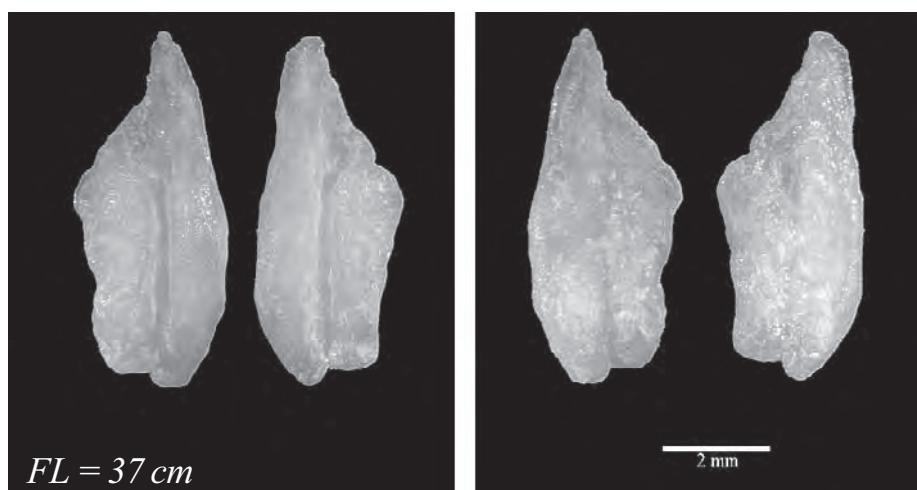
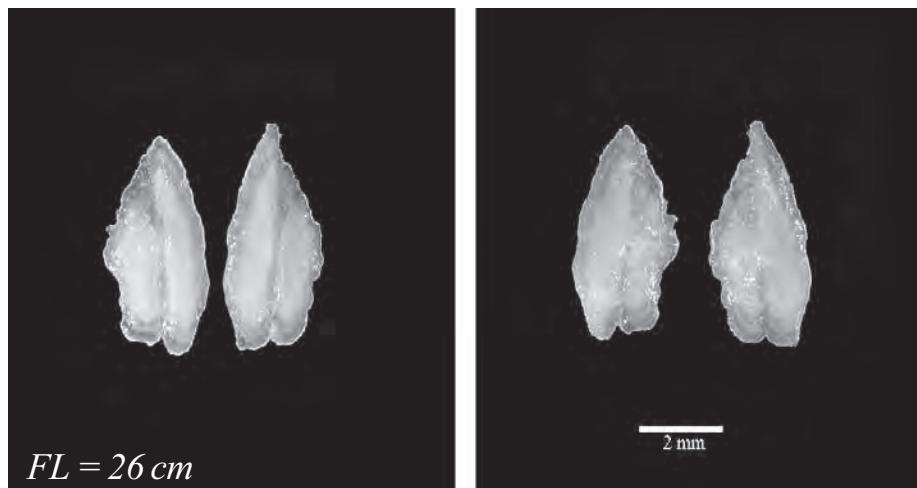
Salmonidae: *Coregonus clupeaformis*



Salmoniformes

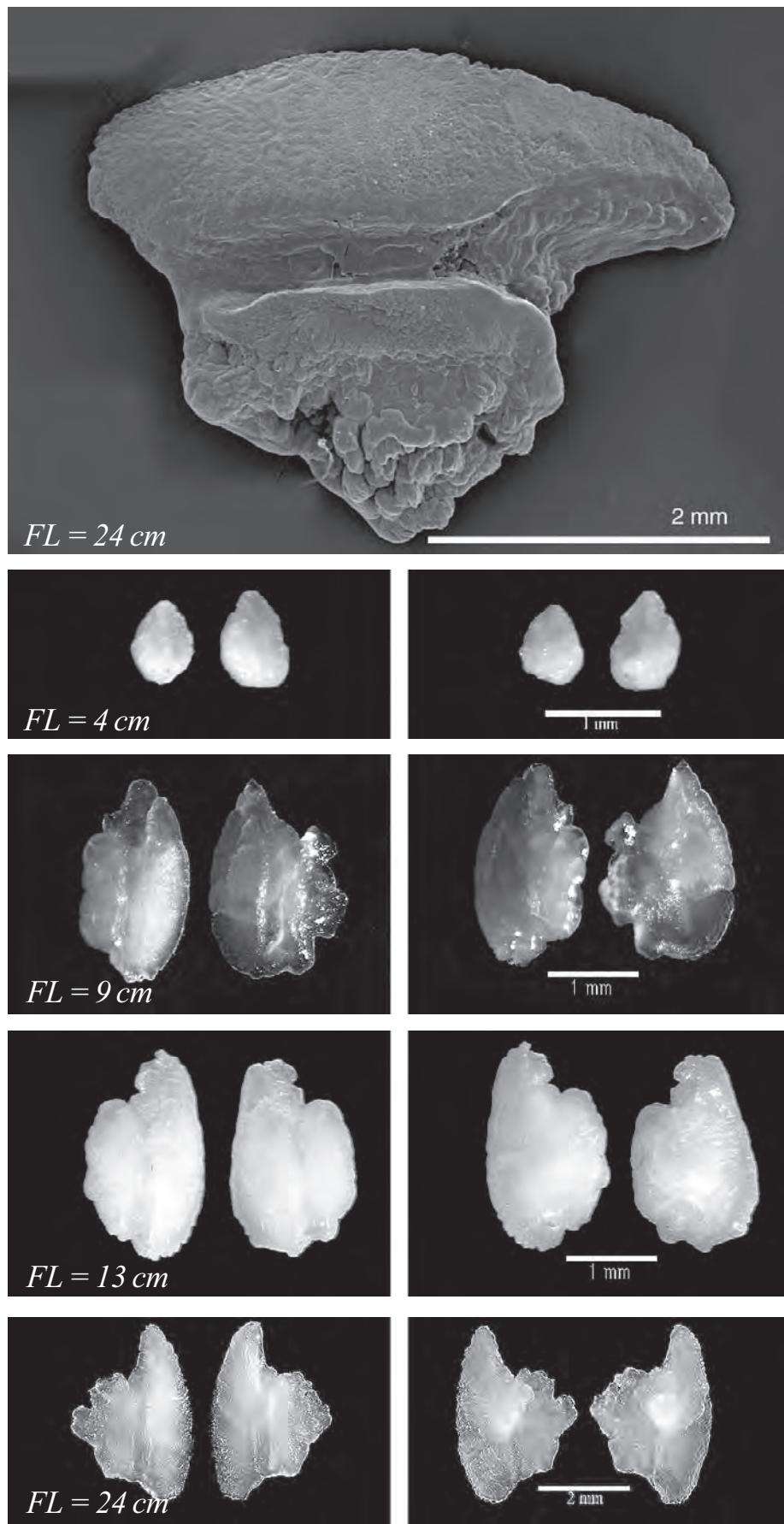
219

Salmonidae: *Coregonus huntsmani*



Salmoniformes

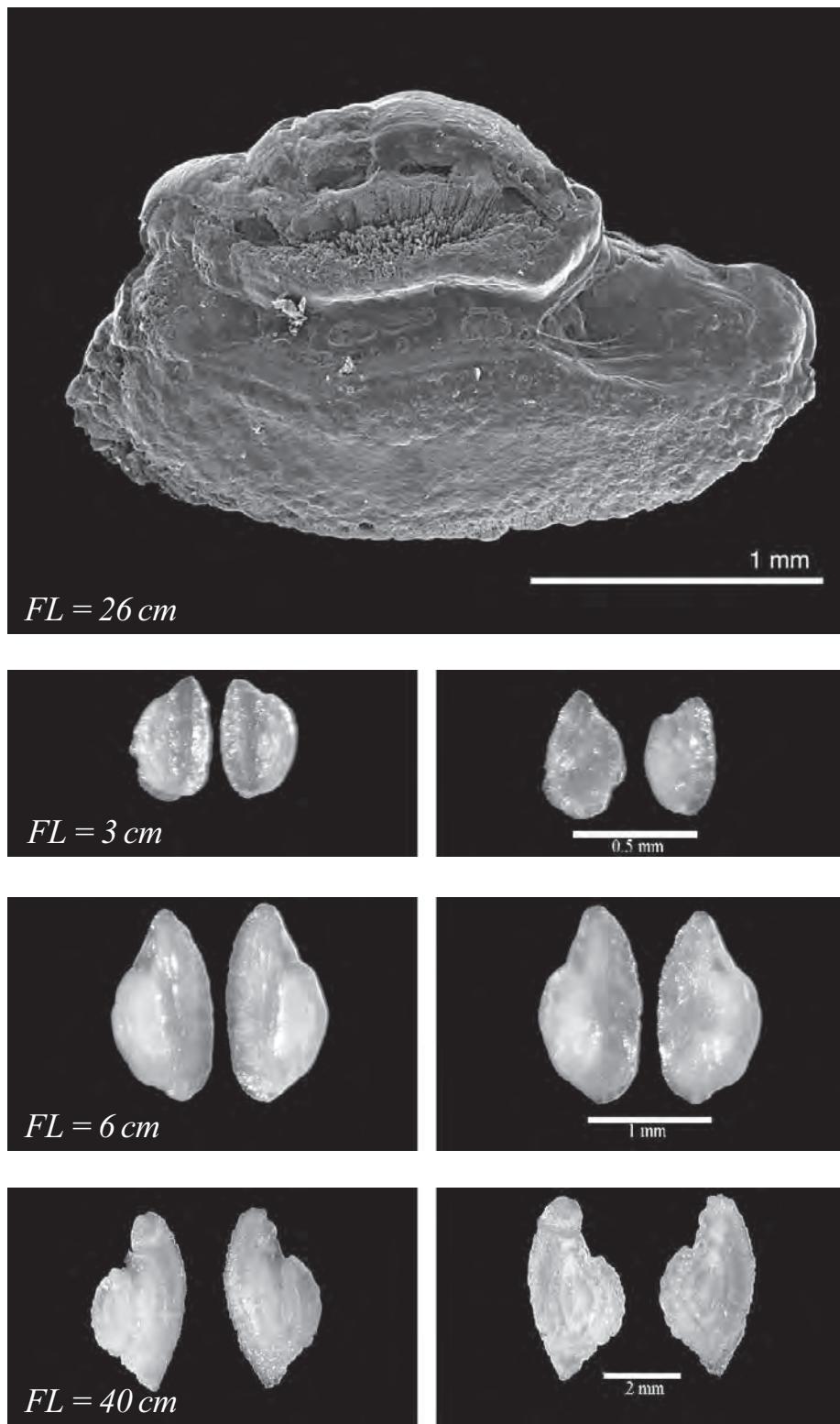
Salmonidae: *Oncorhynchus mykiss*



Salmoniformes

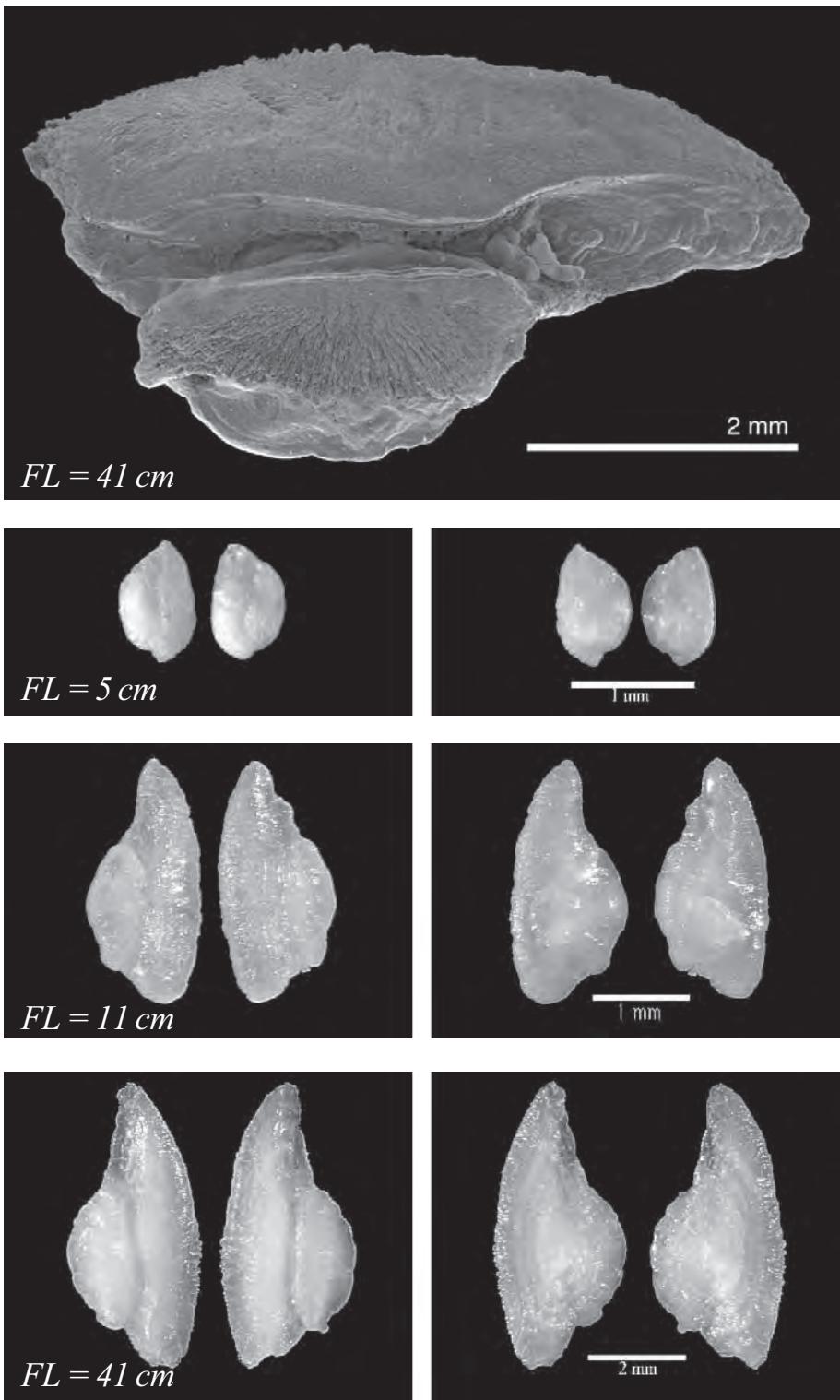
Salmonidae: *Salmo salar*

221

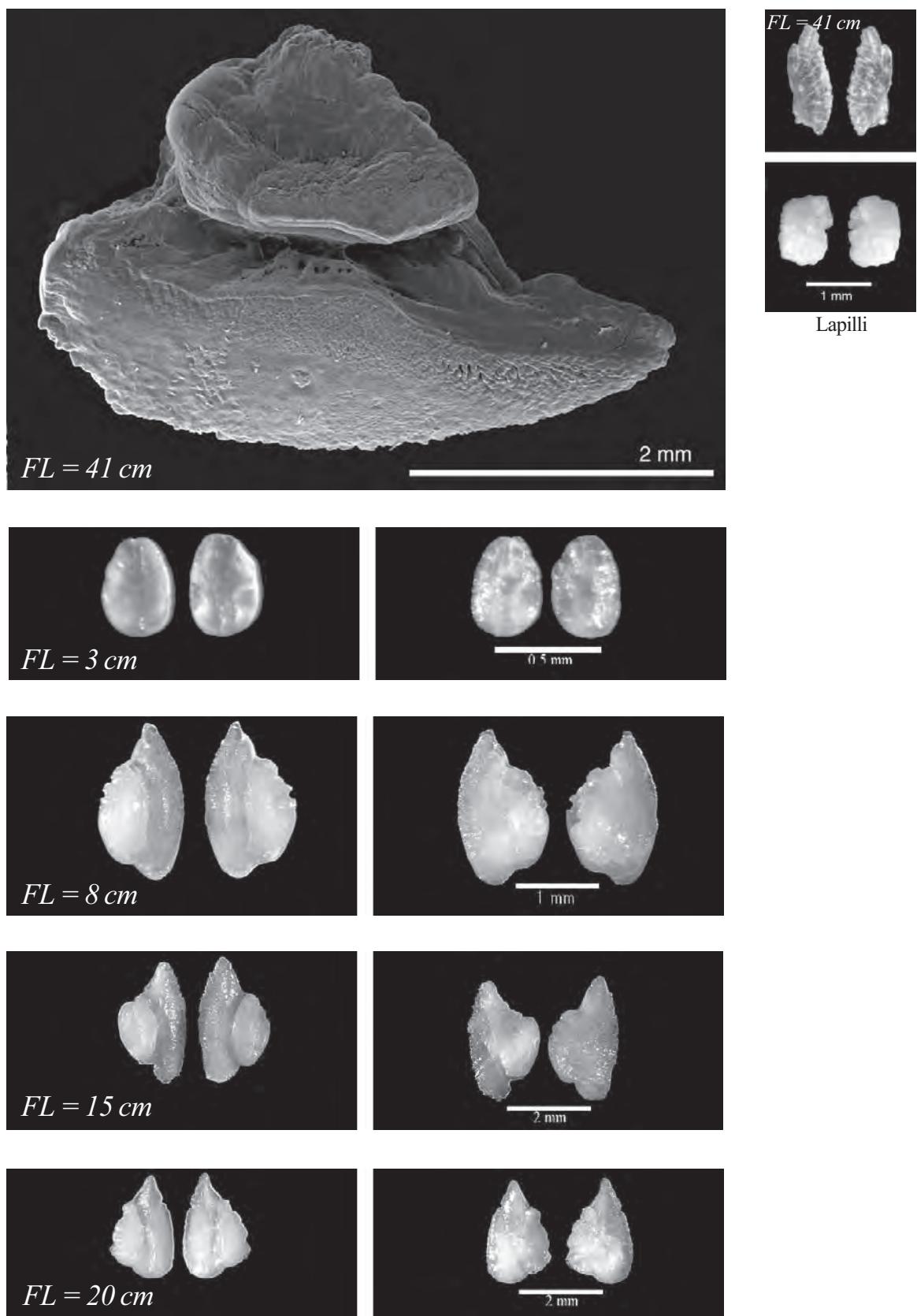


Salmoniformes

Salmonidae: *Salmo trutta*

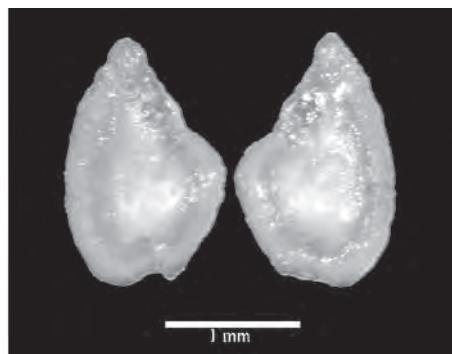
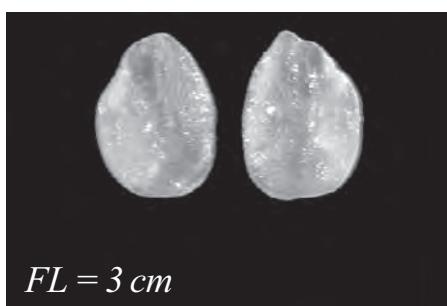
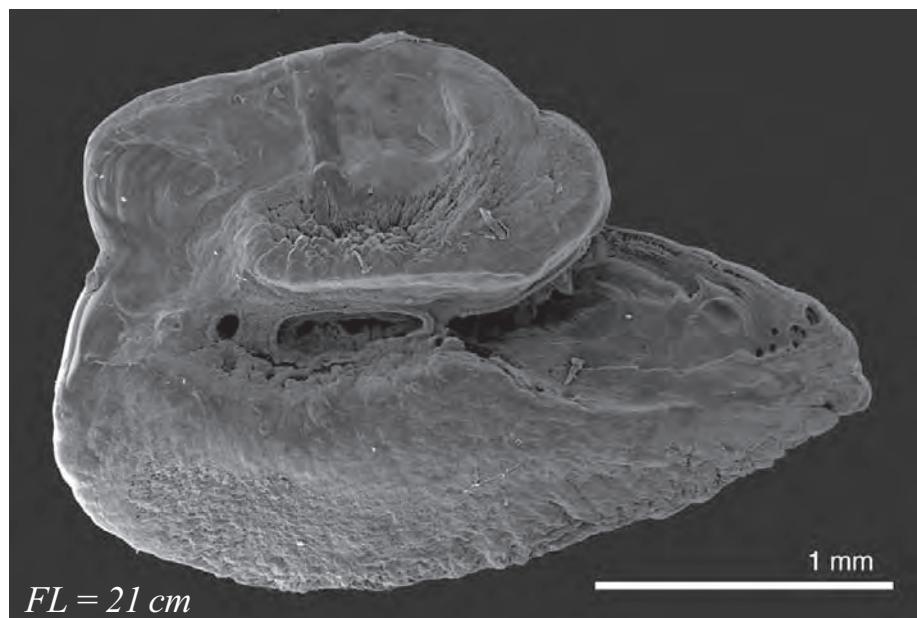


Salmonidae: *Salvelinus alpinus*



Salmoniformes

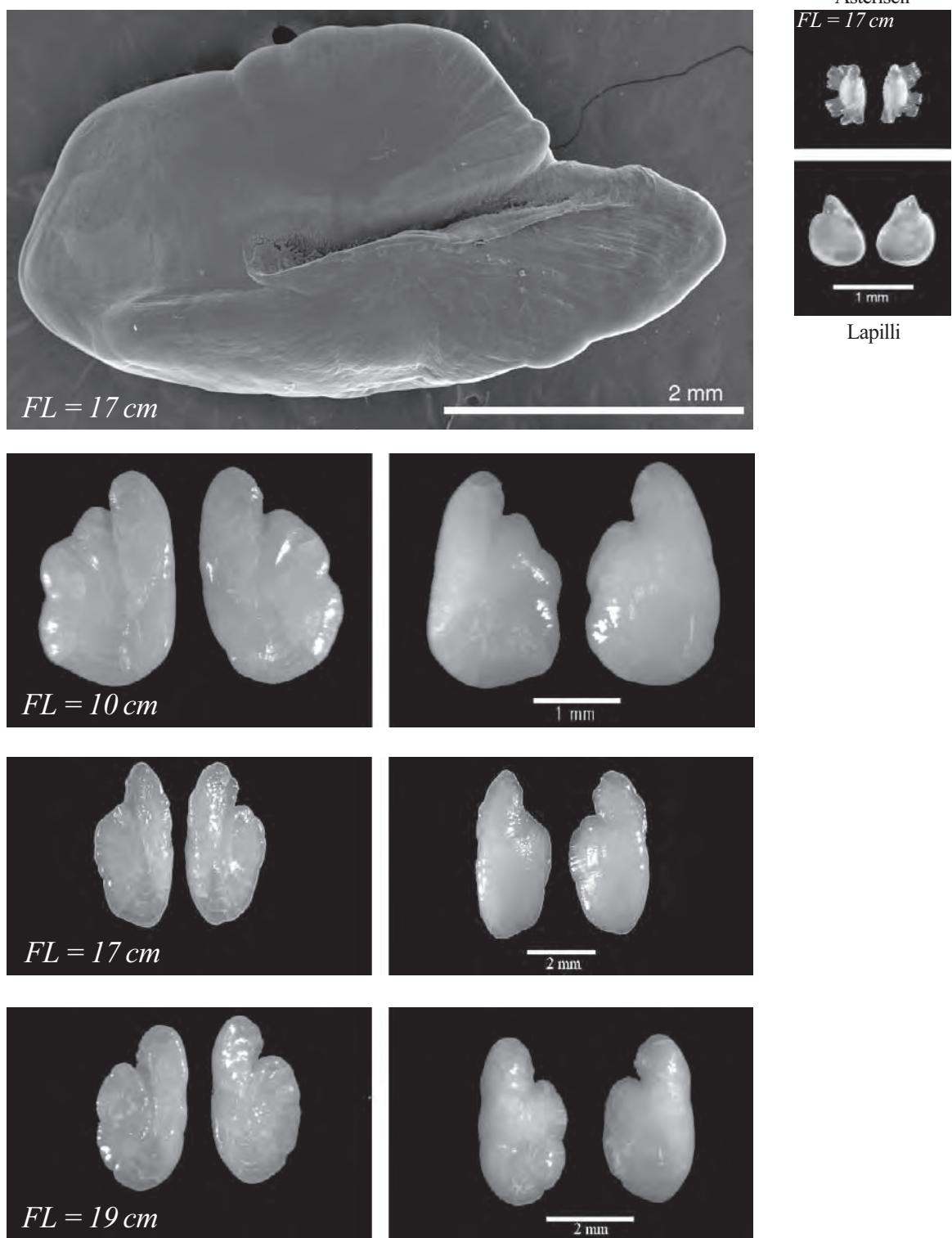
Salmonidae: *Salvelinus fontinalis*



Scorpaeniformes

225

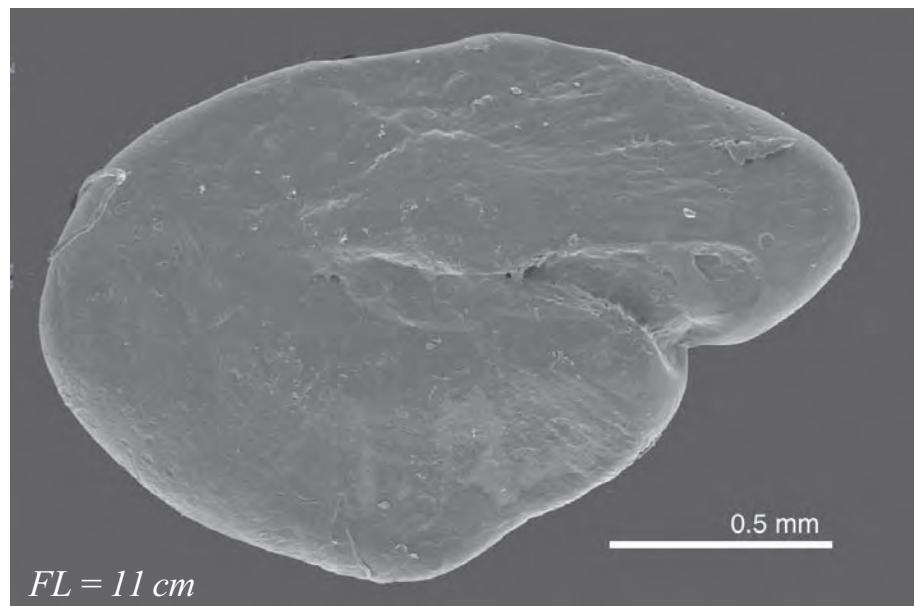
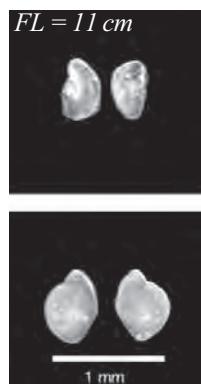
Agonidae: *Agonus decagonus*



Scorpaeniformes

Agonidae: *Aspidophoroides monopterygius*

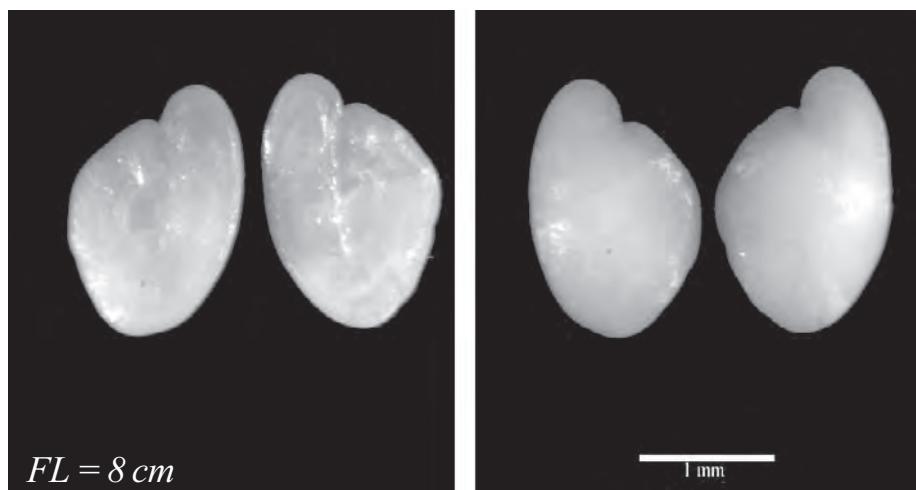
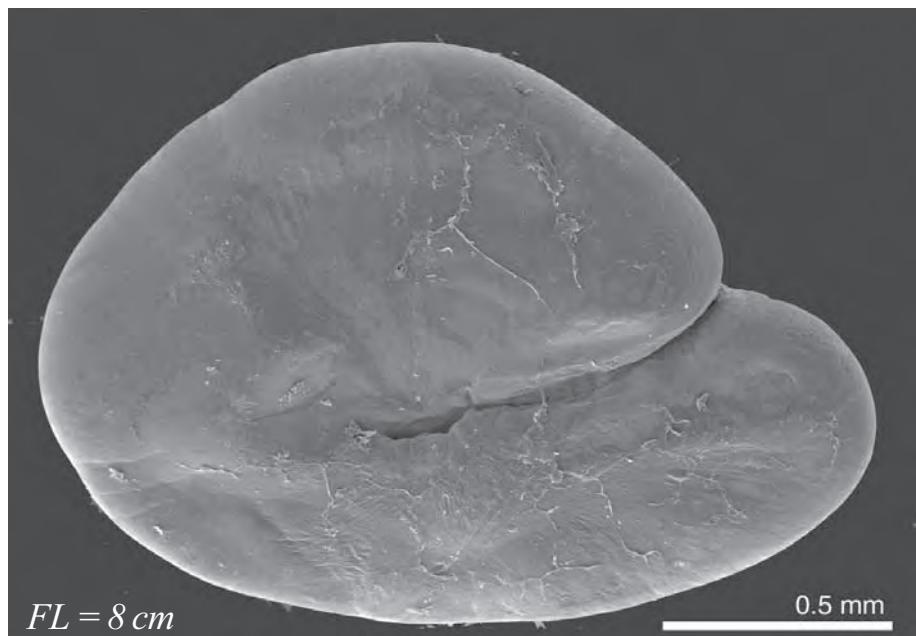
Asterisci



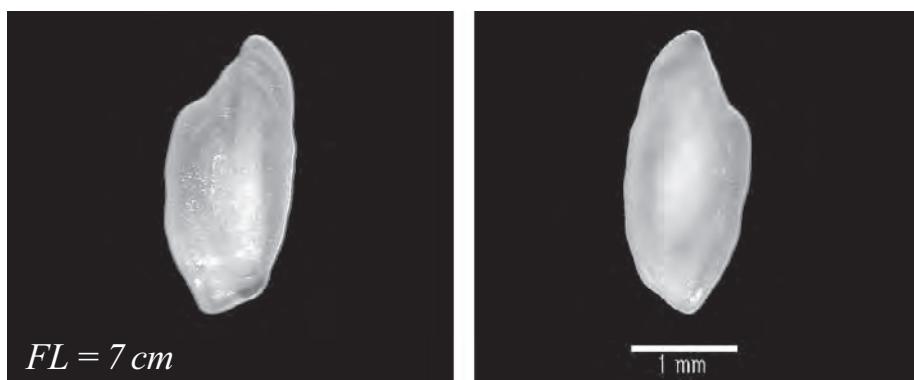
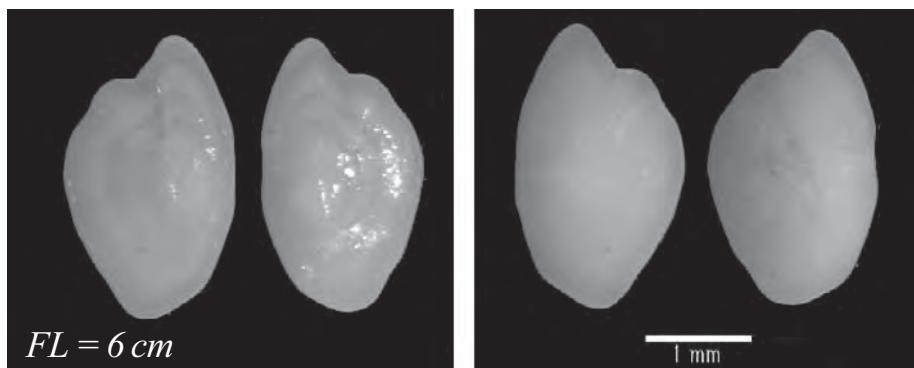
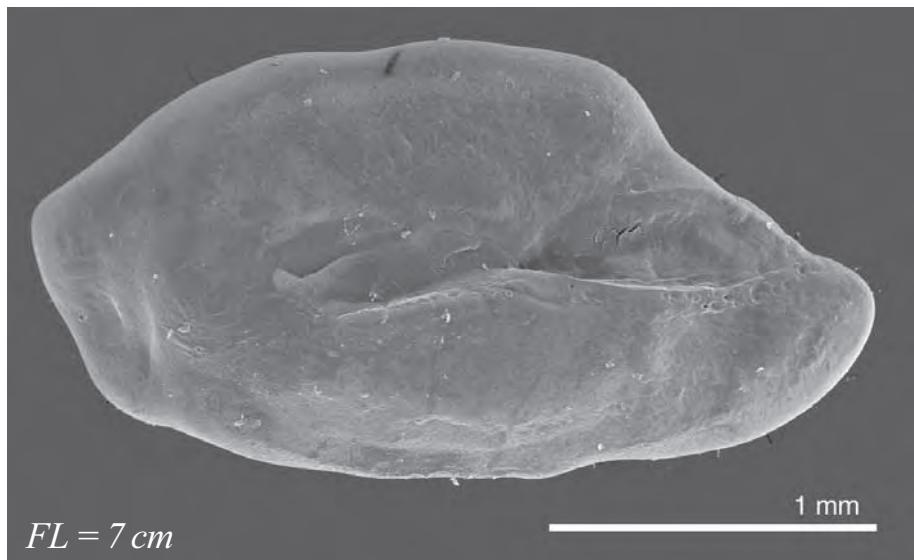
Scorpaeniformes

227

Agonidae: *Aspidophoroides olriki*



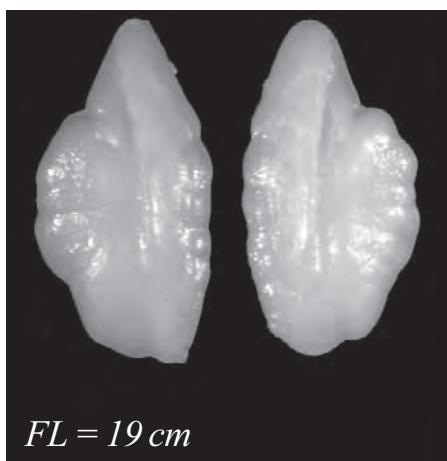
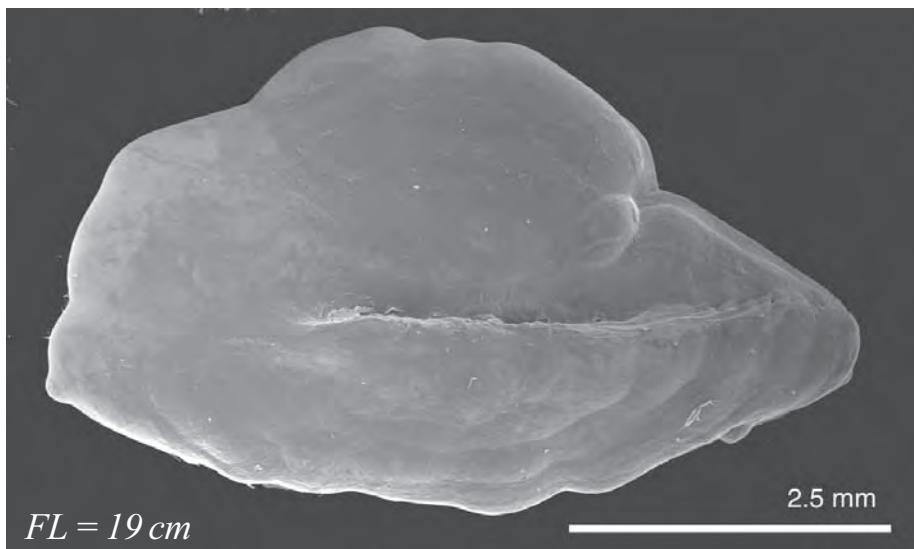
Scorpaeniformes
Cottidae: *Artemiellus atlanticus*



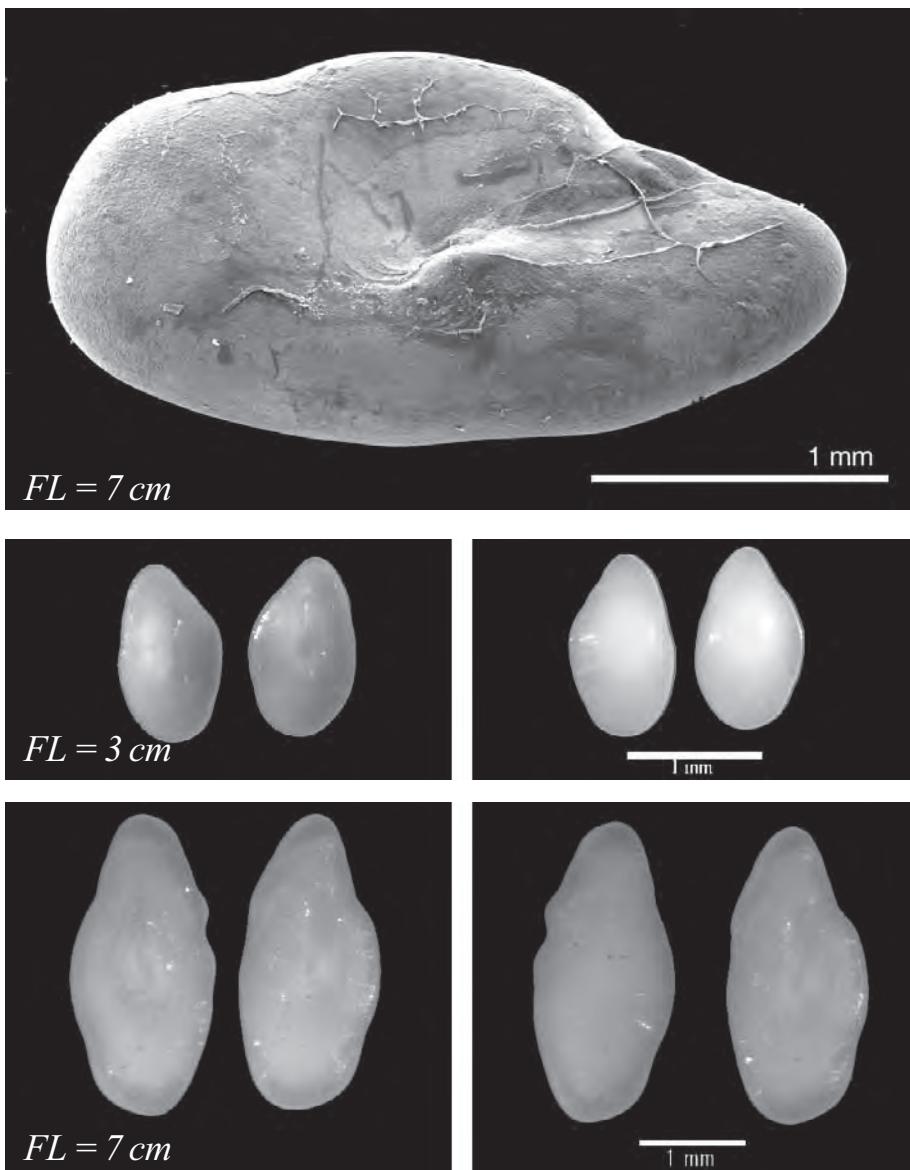
Scorpaeniformes

229

Cottidae: *Gymnocanthus tricuspis*



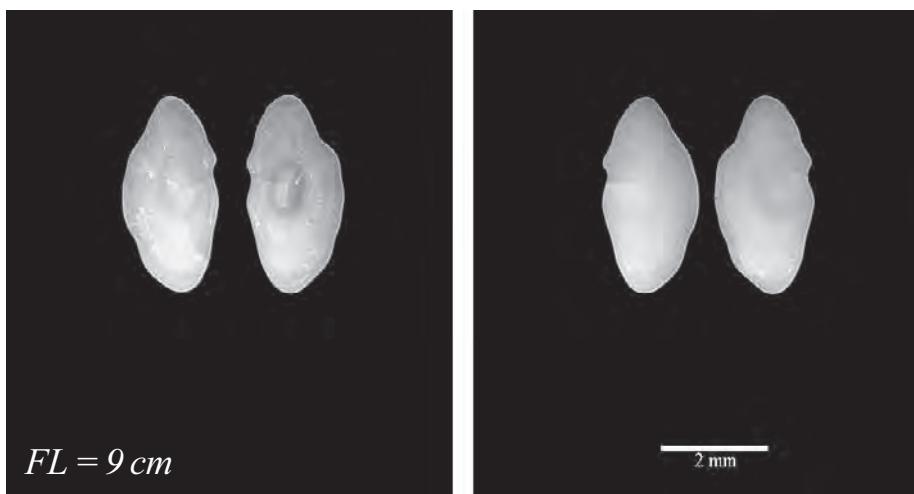
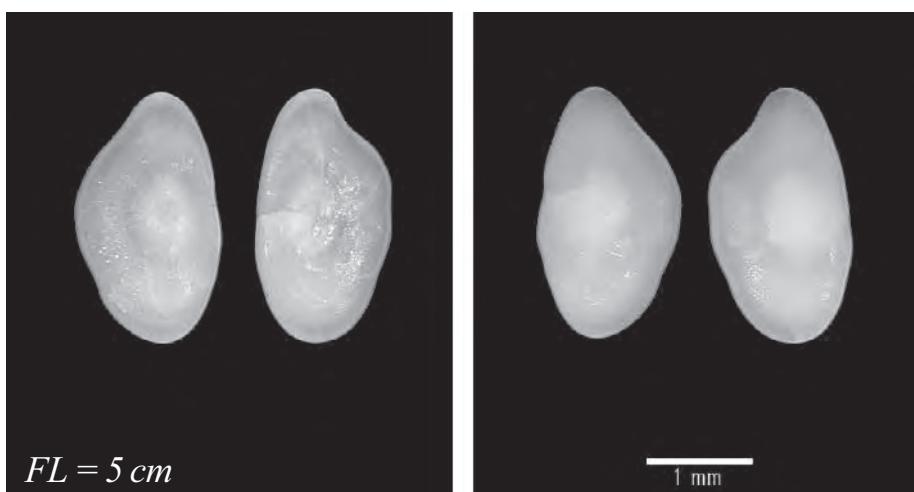
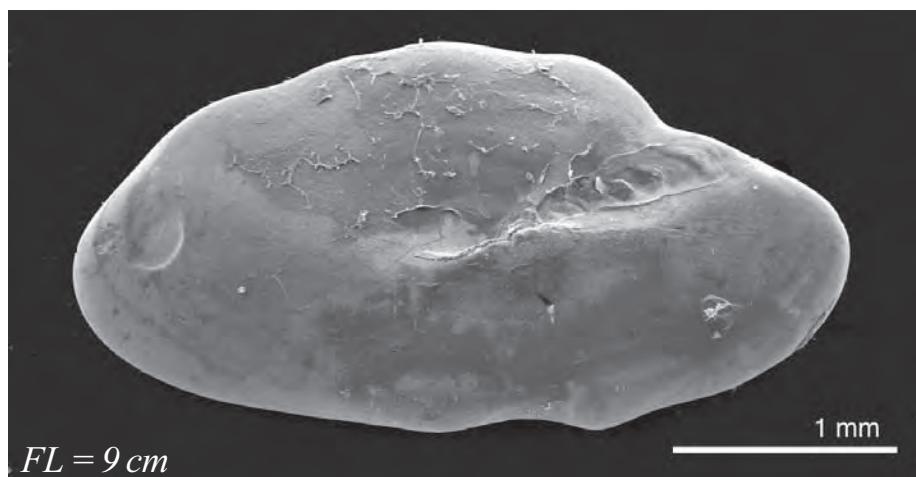
Scorpaeniformes
Cottidae: *Icelus bicornis*



Scorpaeniformes

Cottidae: *Icelus spatula*

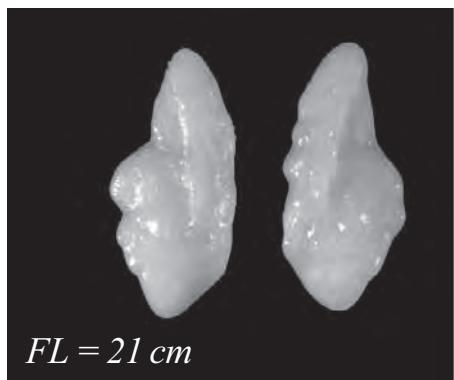
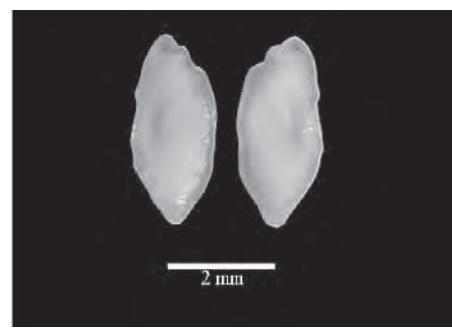
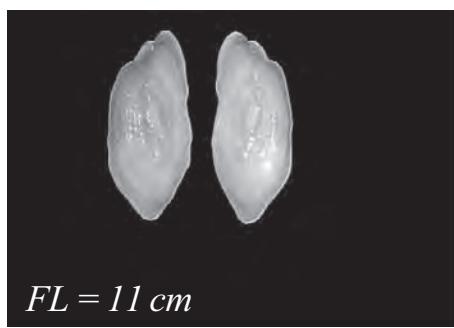
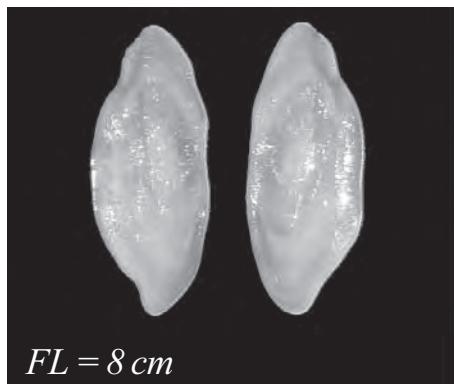
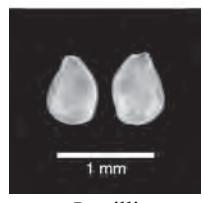
231



Scorpaeniformes

Cottidae: *Myoxocephalus aenaeus*

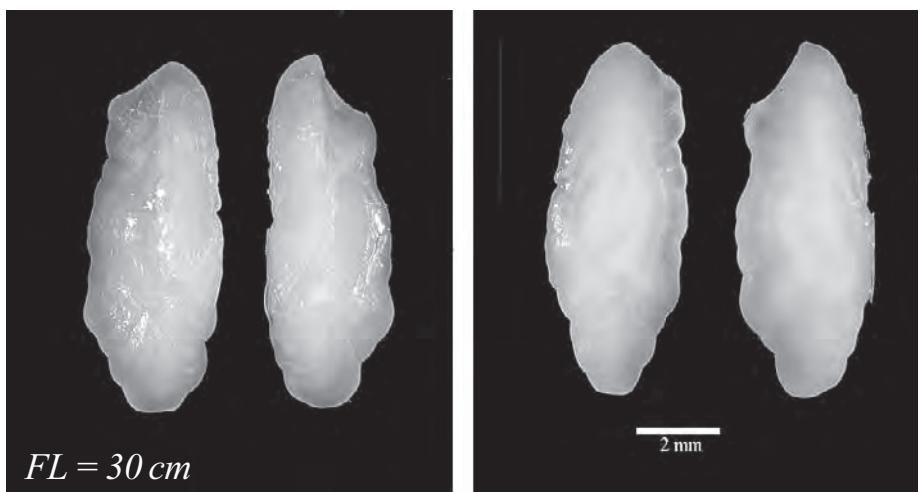
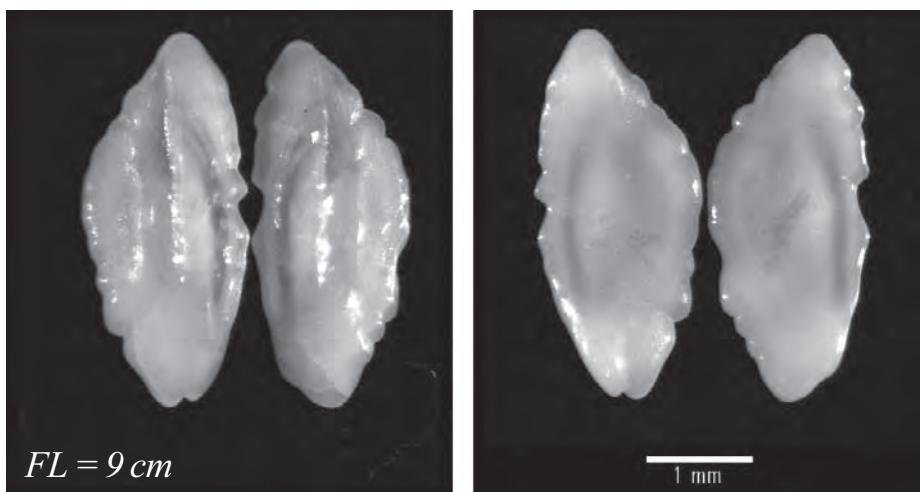
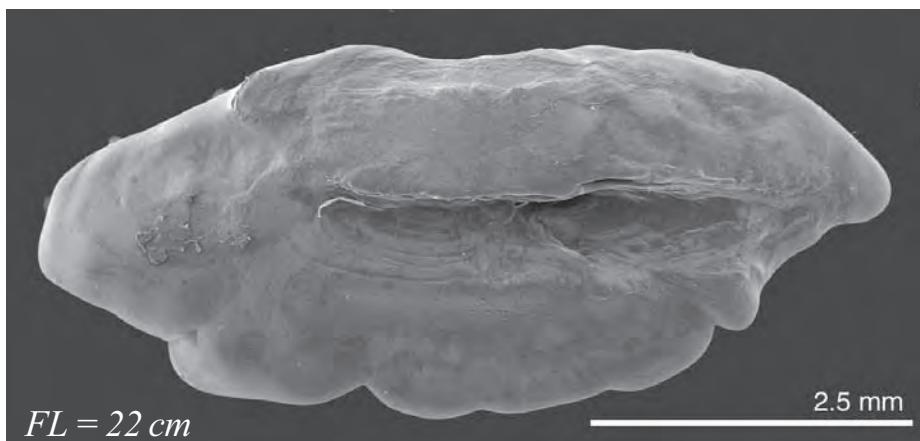
Asterisci



Scorpaeniformes

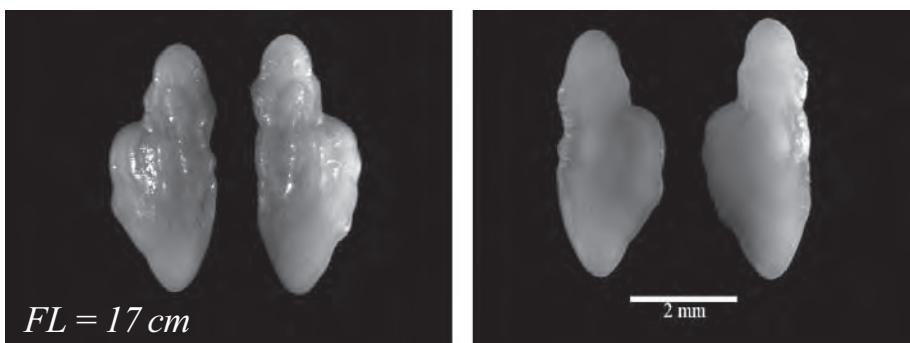
233

Cottidae: *Myoxocephalus octodecemspinosis*



Scorpaeniformes

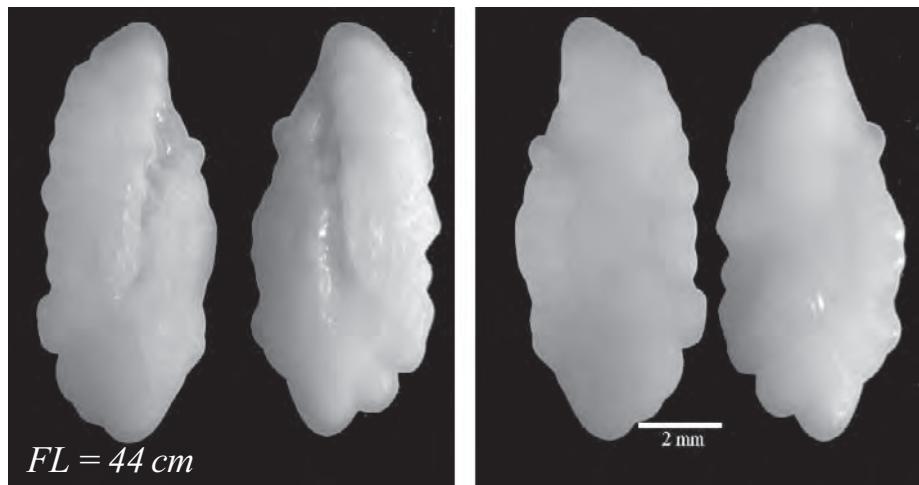
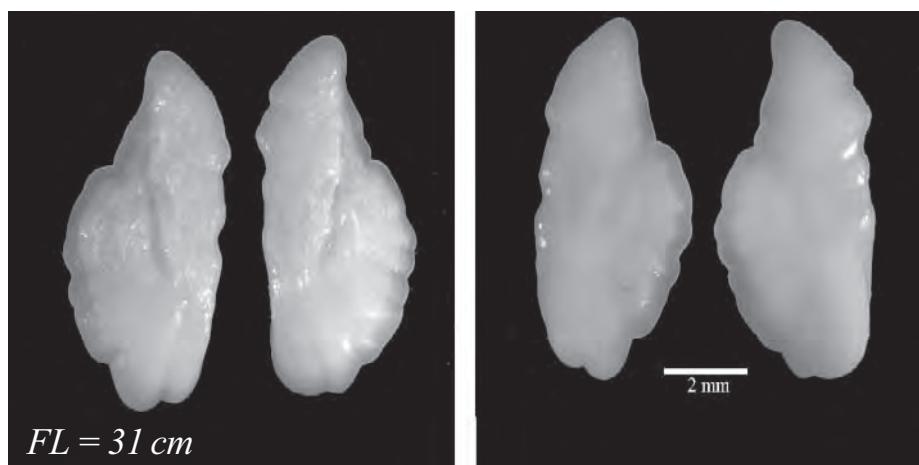
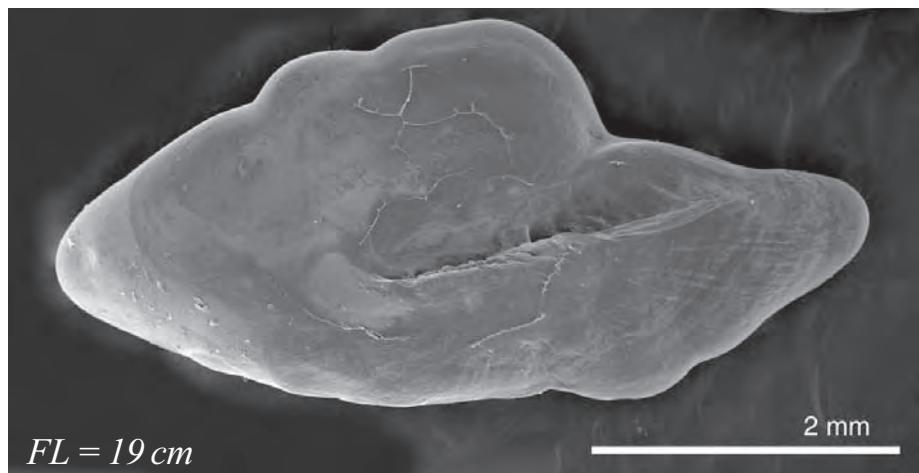
Cottidae: *Myoxocephalus scorpioides*



Scorpaeniformes

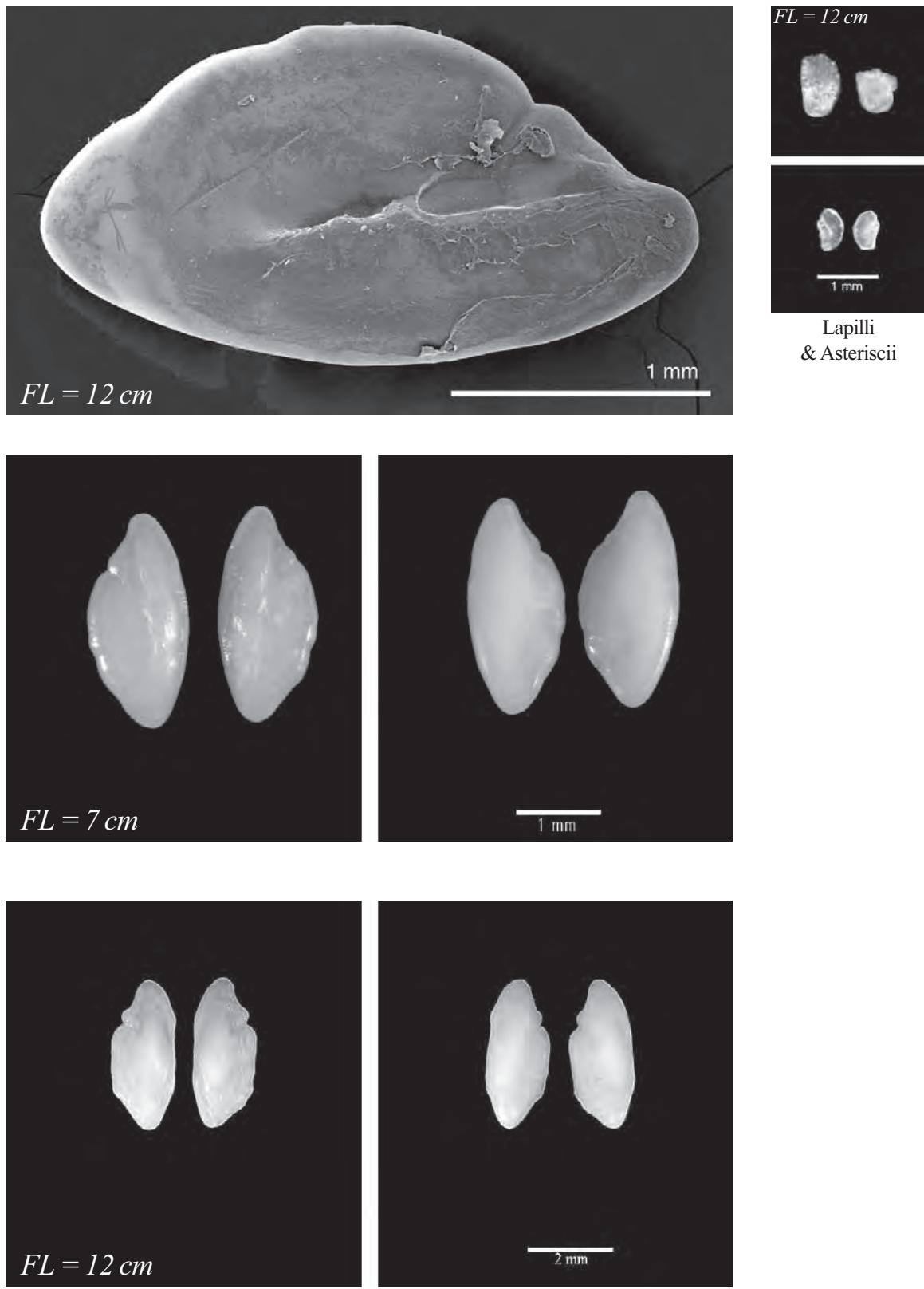
235

Cottidae: *Myoxocephalus scorpius*



Scorpaeniformes

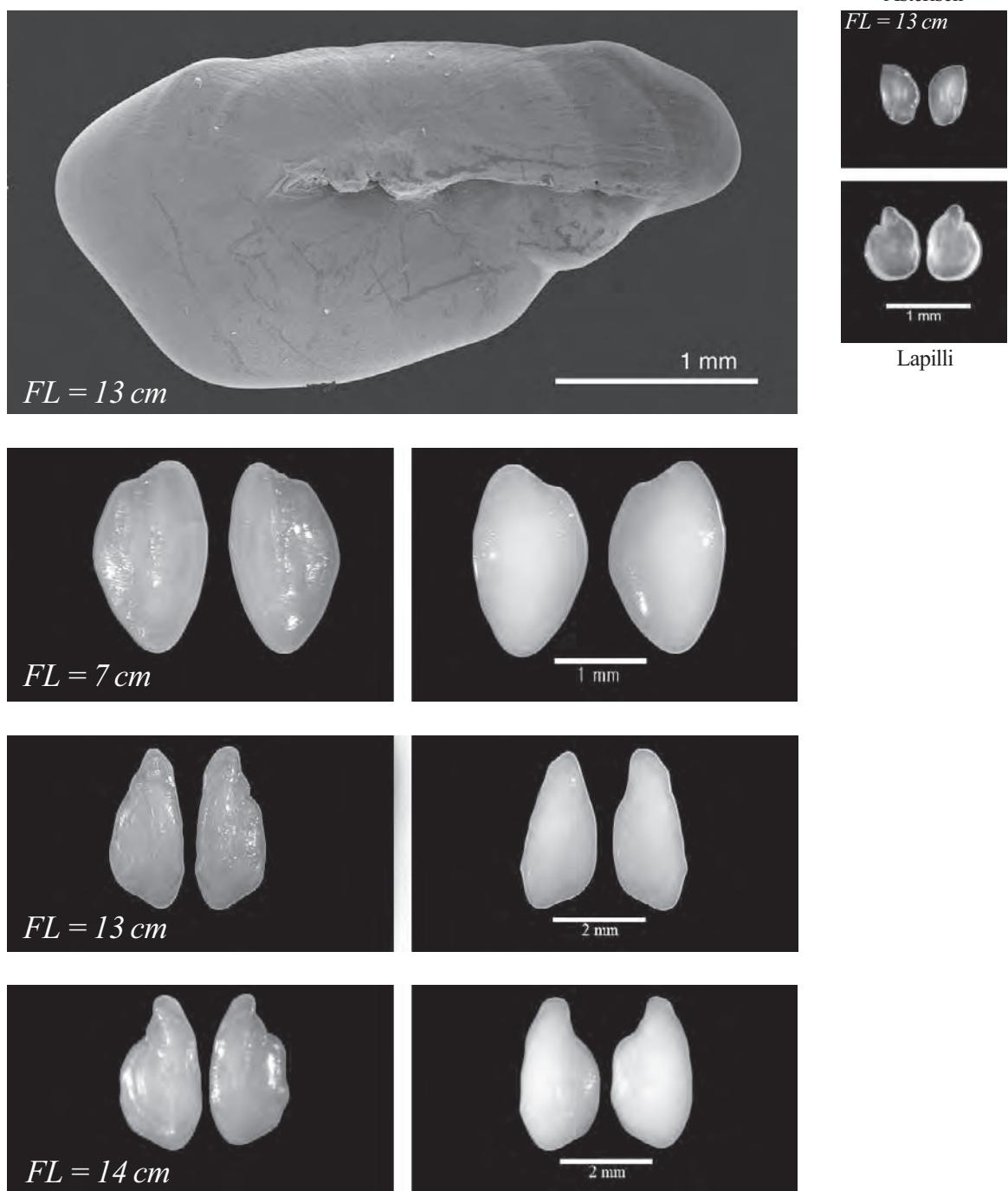
Cottidae: *Triglops murrayi*



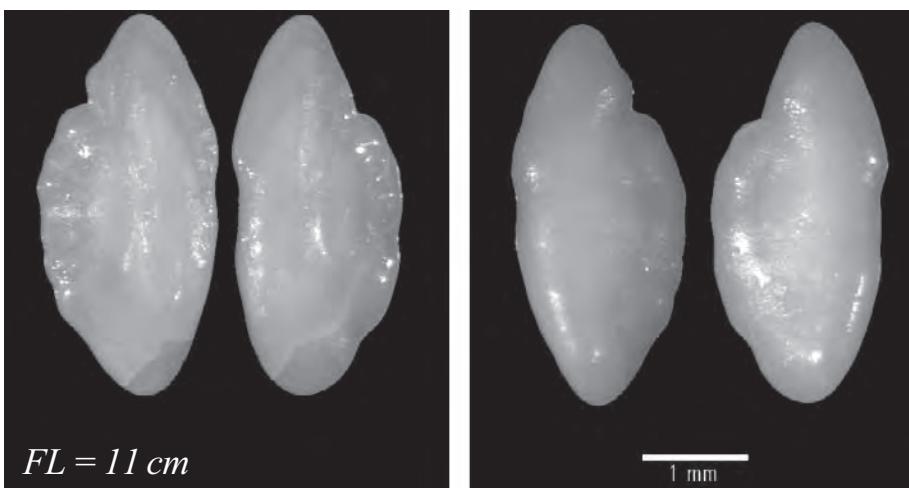
Scorpaeniformes

Cottidae: *Triglops nybelini*

237



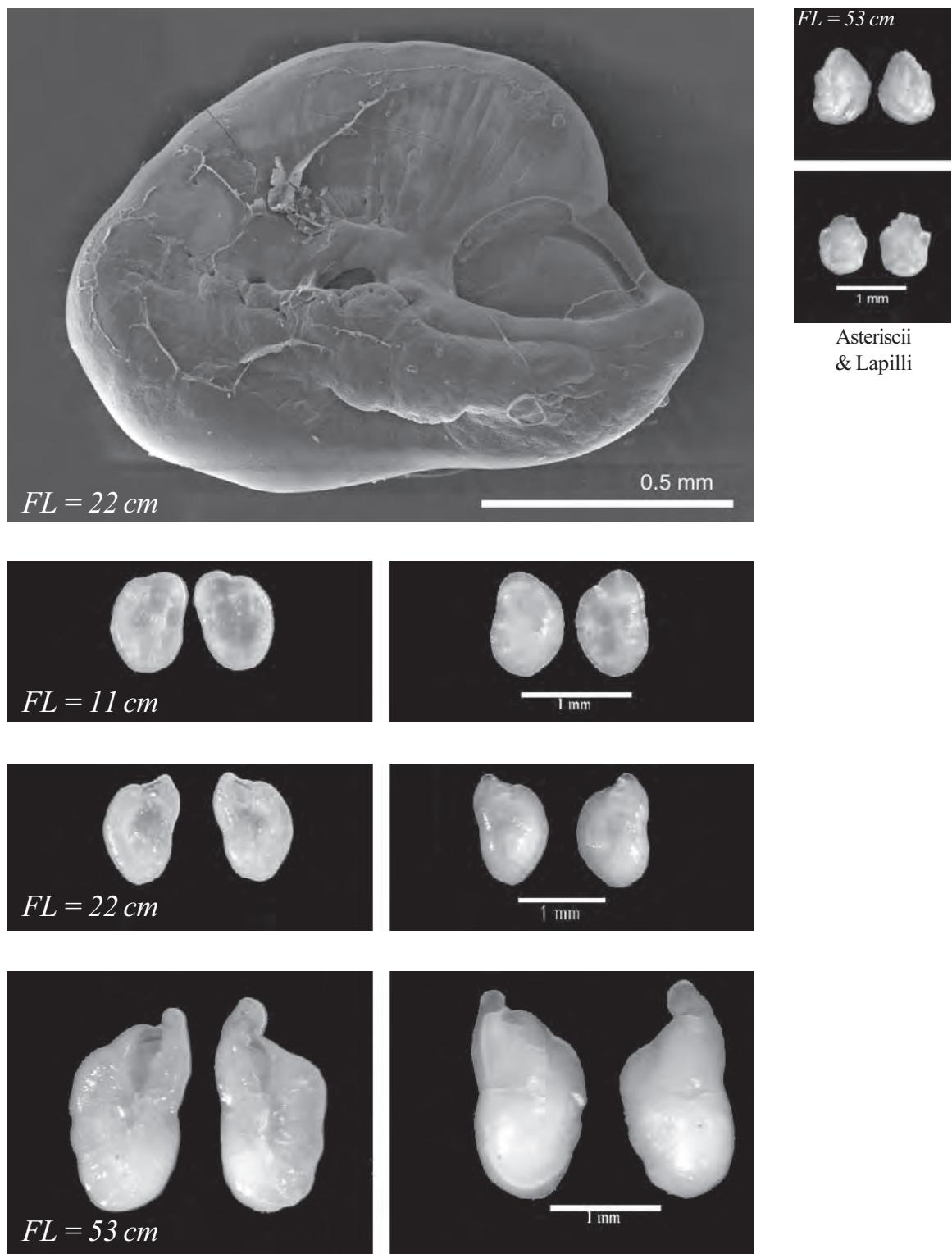
Scorpaeniformes
Cottidae: *Triglops pingeli*



Scorpaeniformes

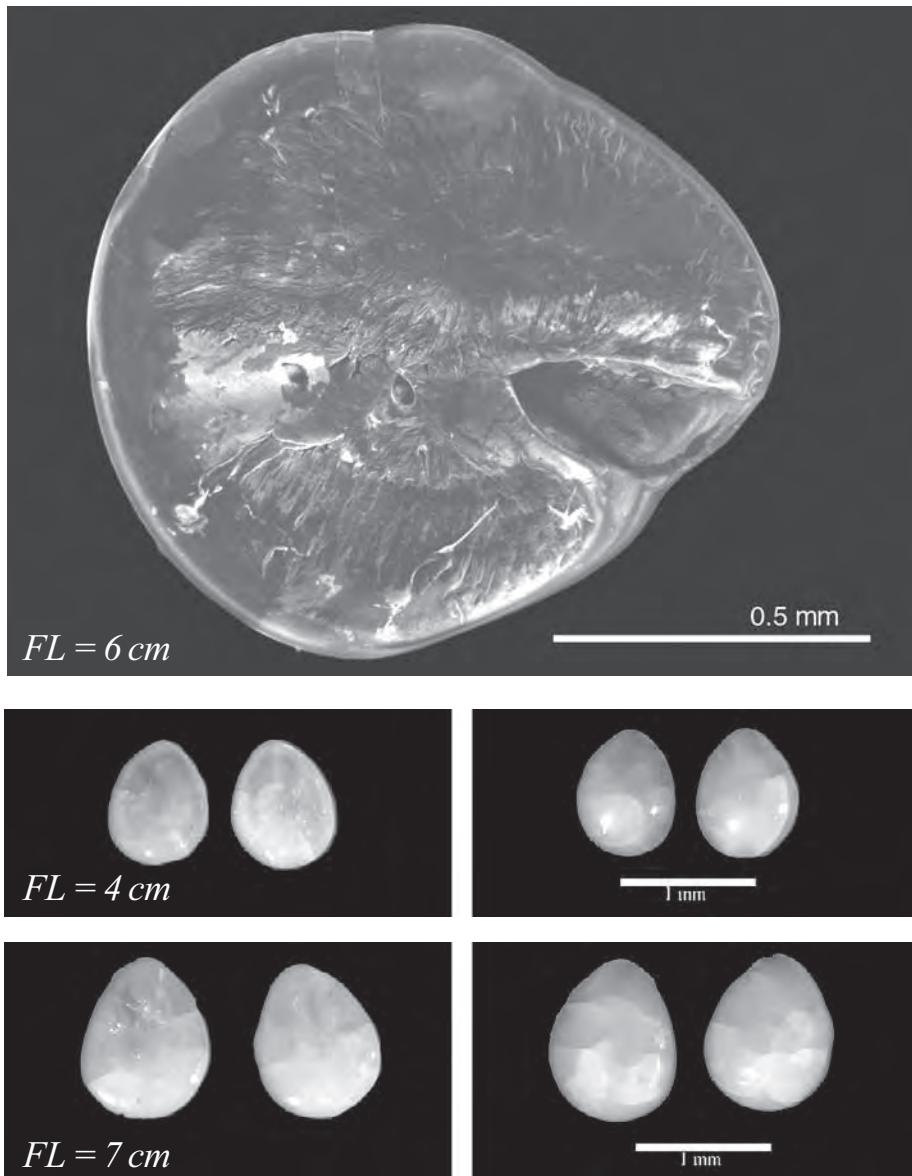
239

Cyclopteridae: *Cyclopterus lumpus*



Scorpaeniformes

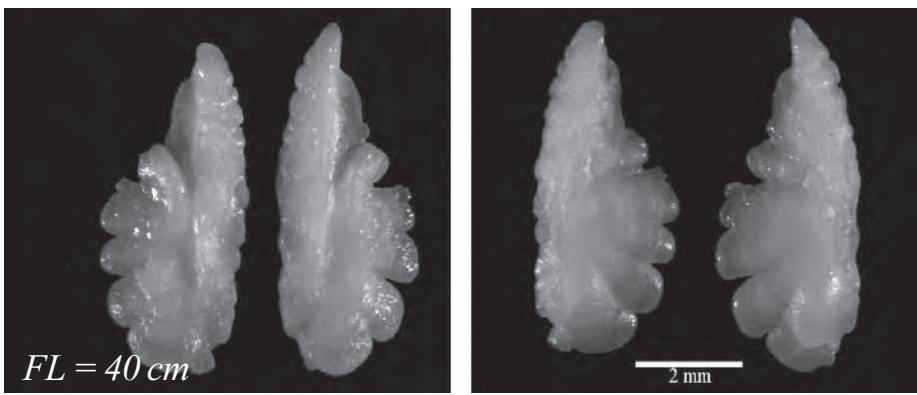
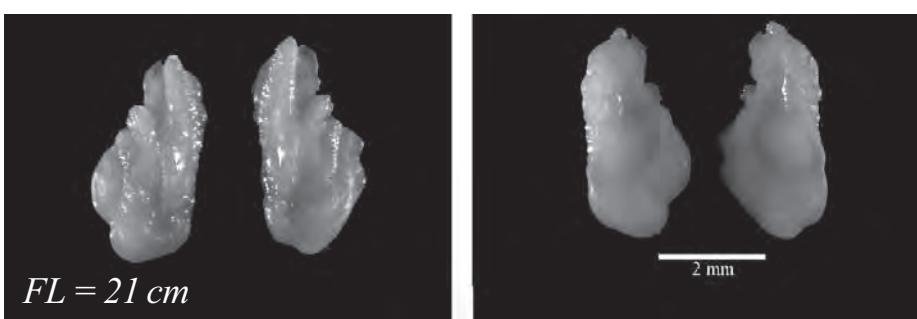
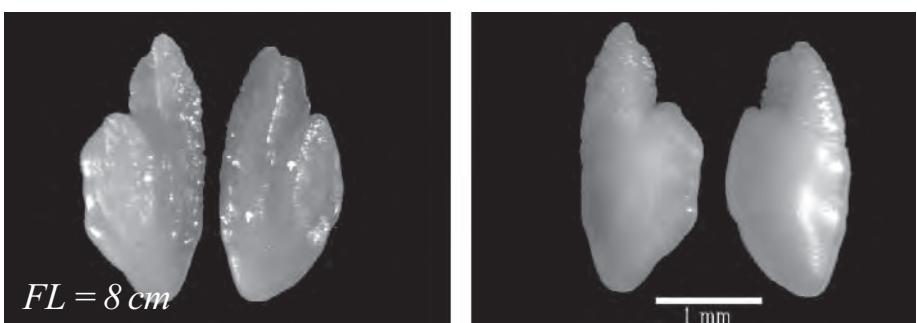
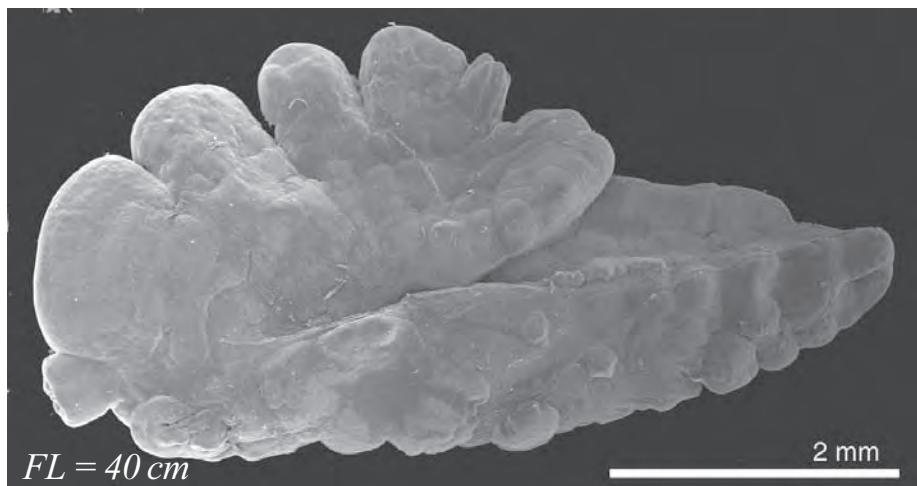
Cyclopteridae: *Eumicrotremus spinosus*



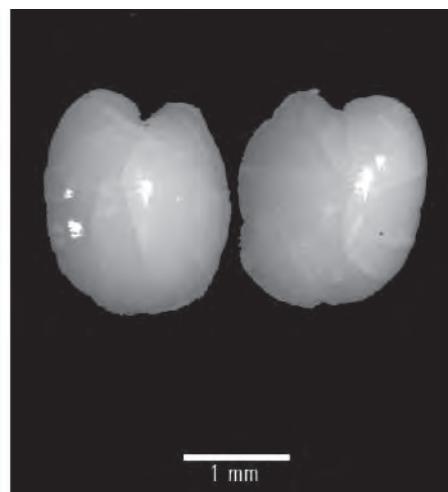
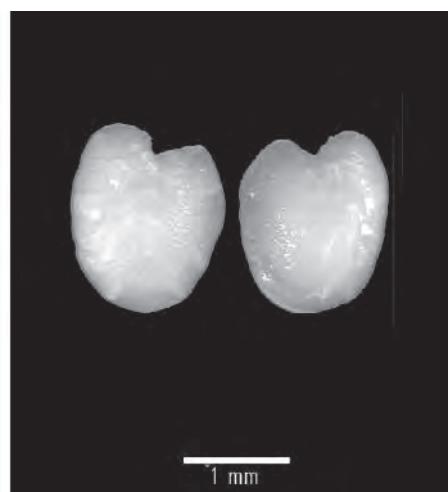
Scorpaeniformes

241

Hemitripteridae: *Hemitripterus americanus*

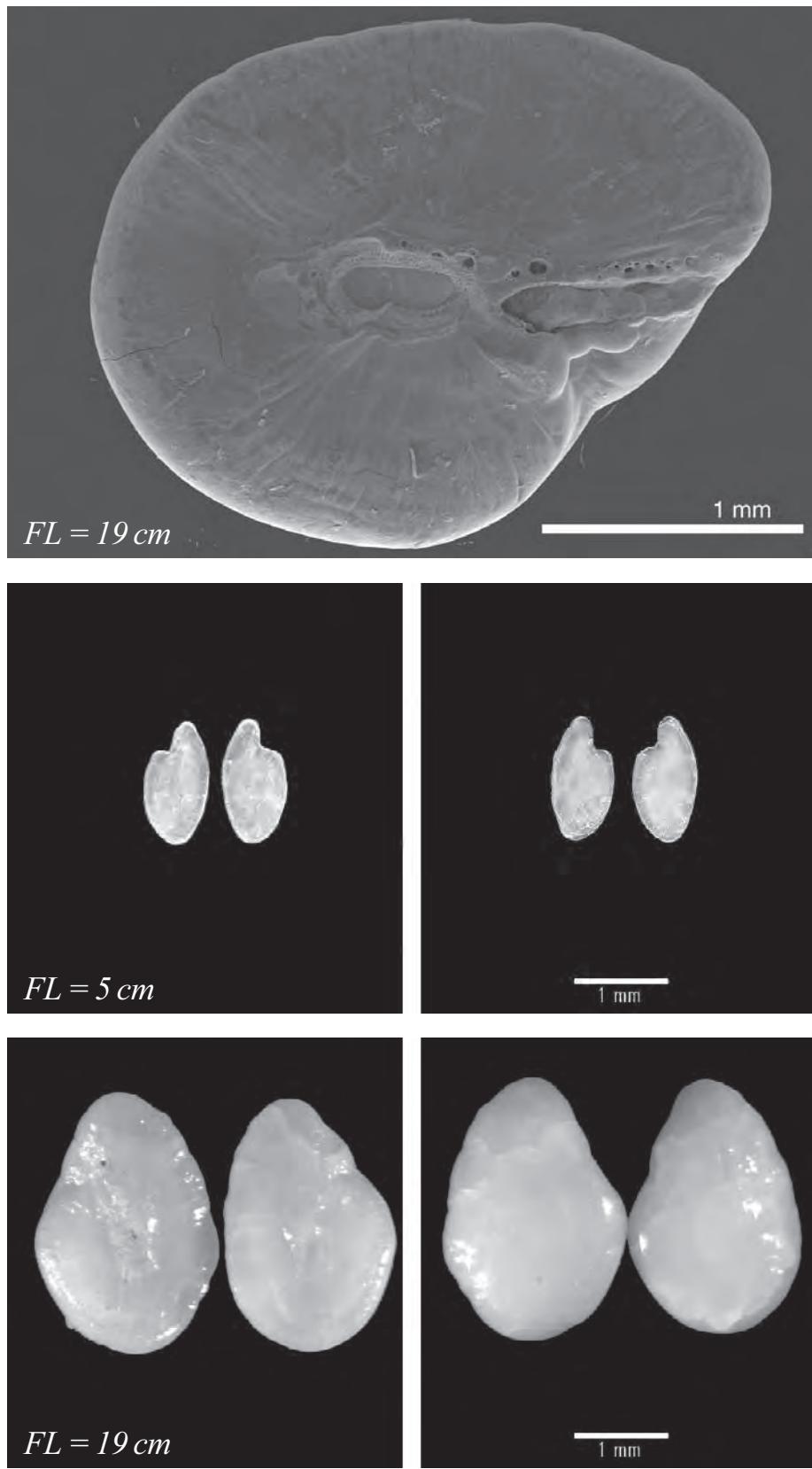


Scorpaeniformes

Liparidae: *Careproctus reinhardti*

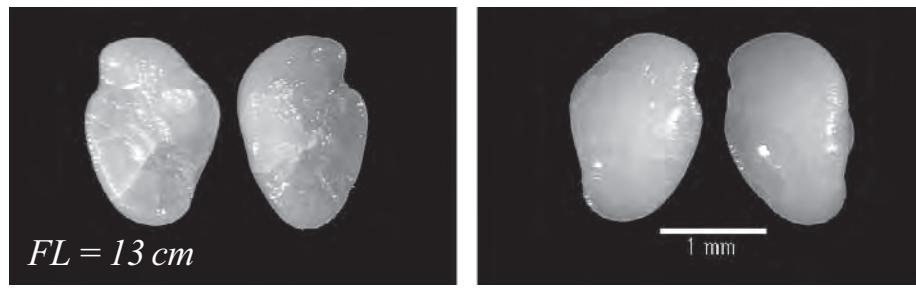
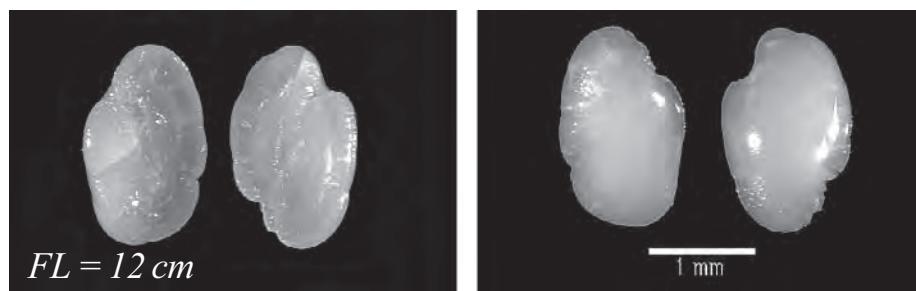
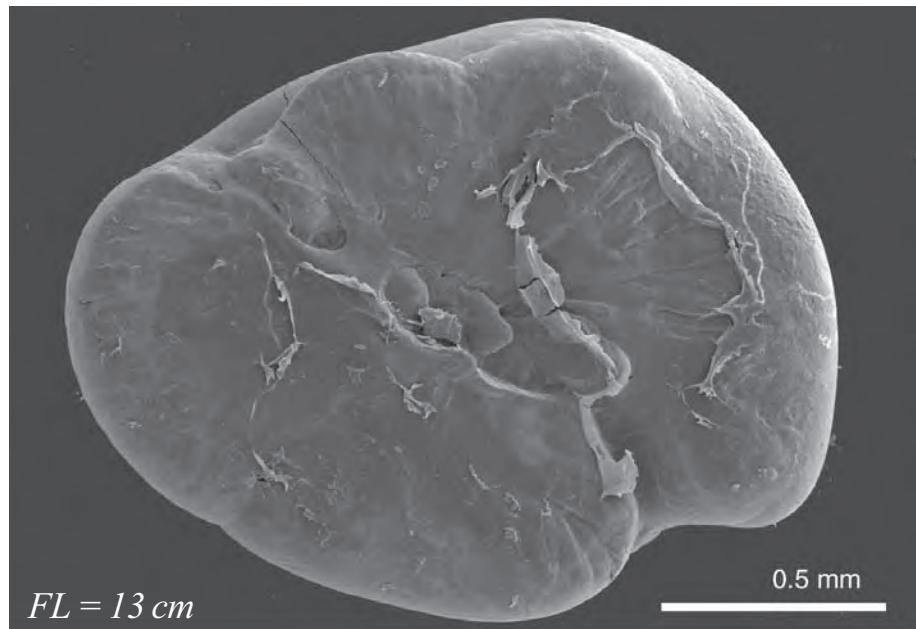
Scorpaeniformes
Liparidae: *Liparis atlanticus*

243



Scorpaeniformes

Liparidae: *Liparis fabricii*

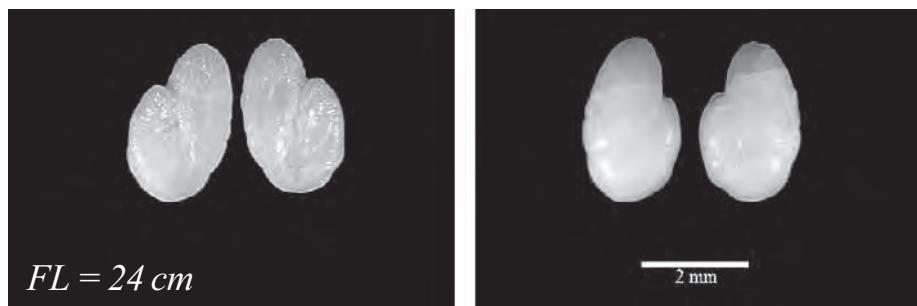
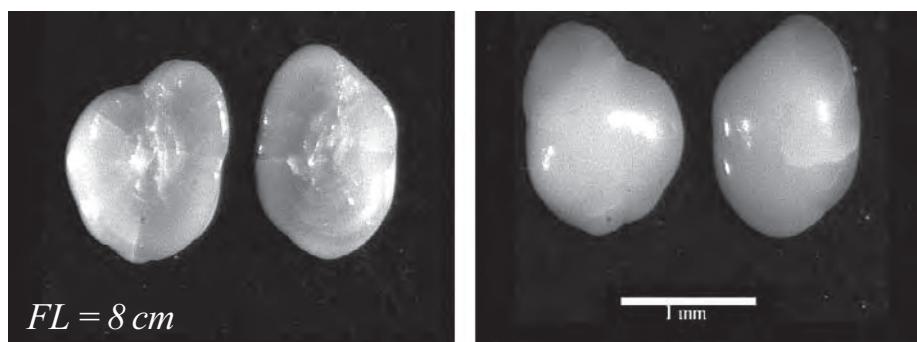
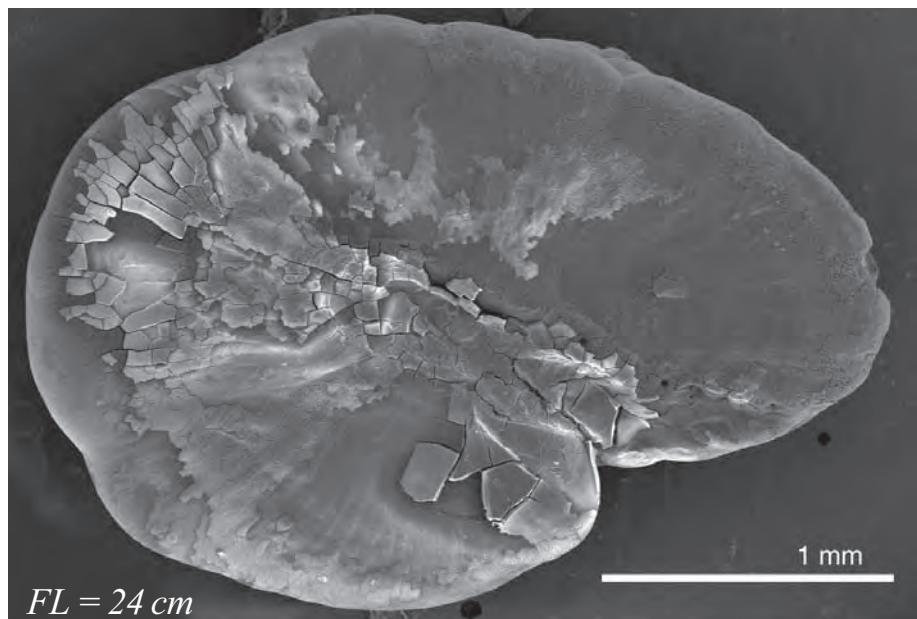
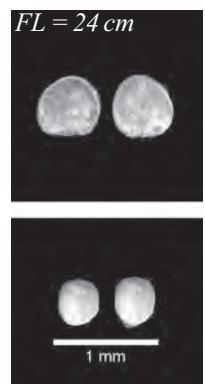


Scorpaeniformes

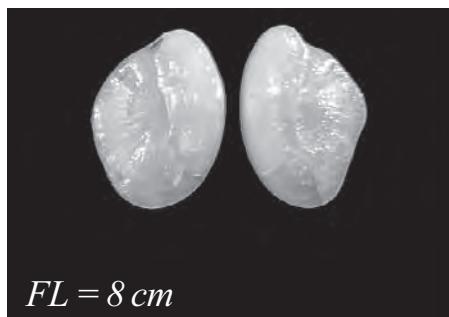
Liparidae: *Liparis gibbus*

245

Asterisci

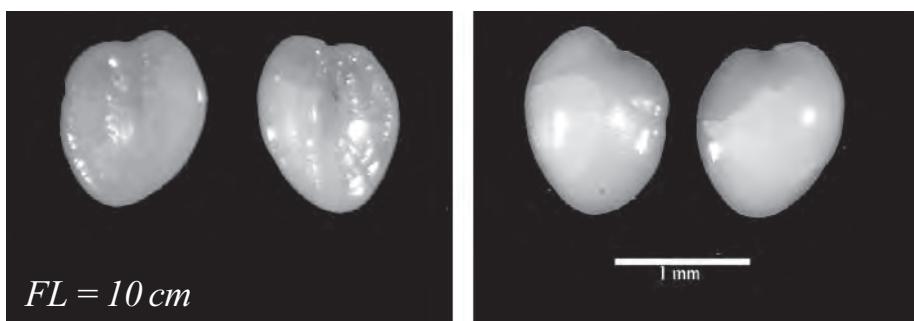
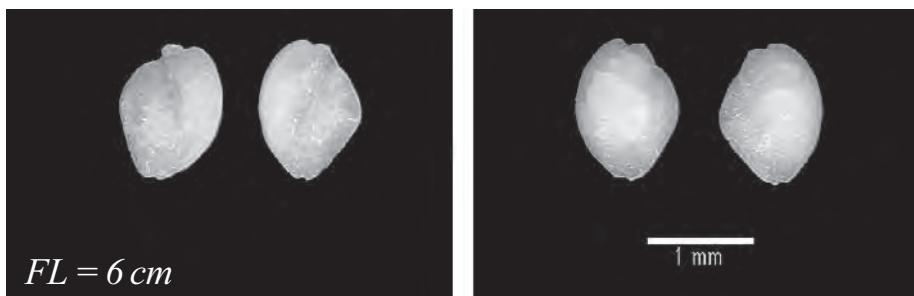


Scorpaeniformes
Liparidae: *Paraliparis calidus*



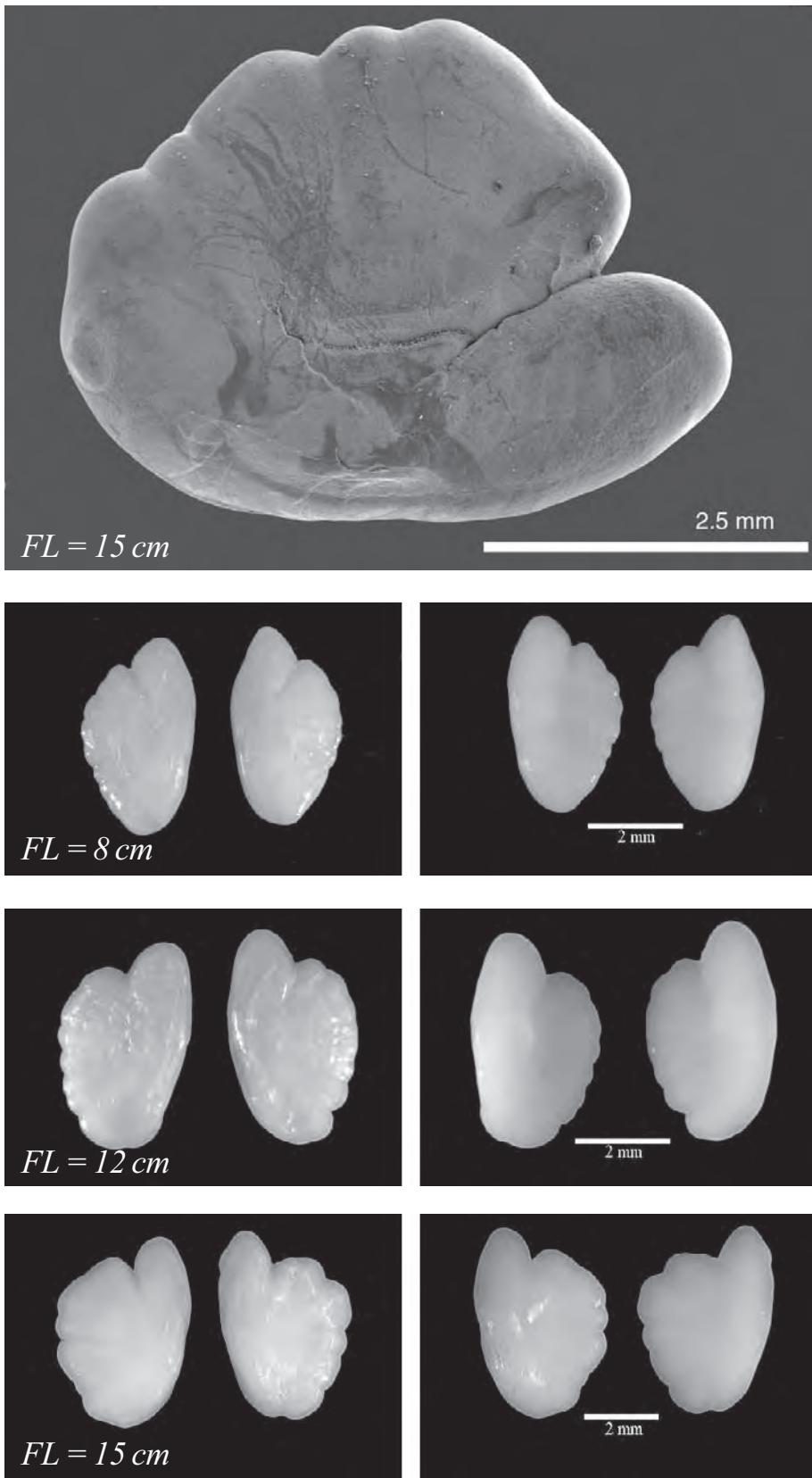
Scorpaeniformes
Liparidae: *Paraliparis copei*

247



Scorpaeniformes

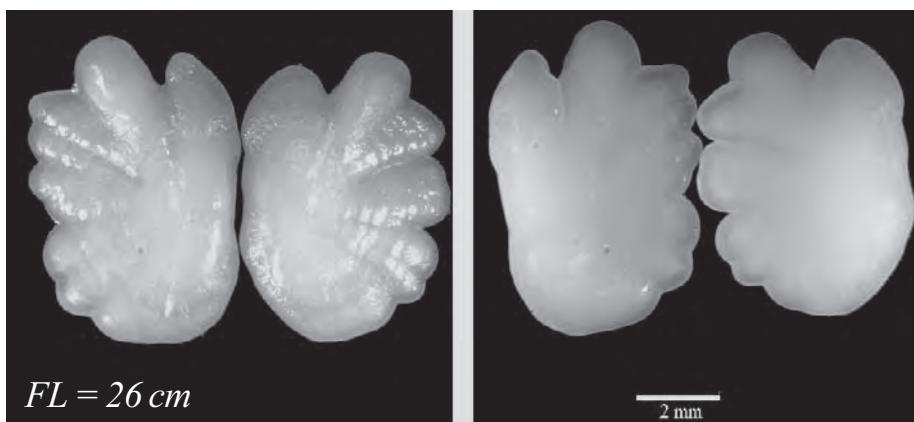
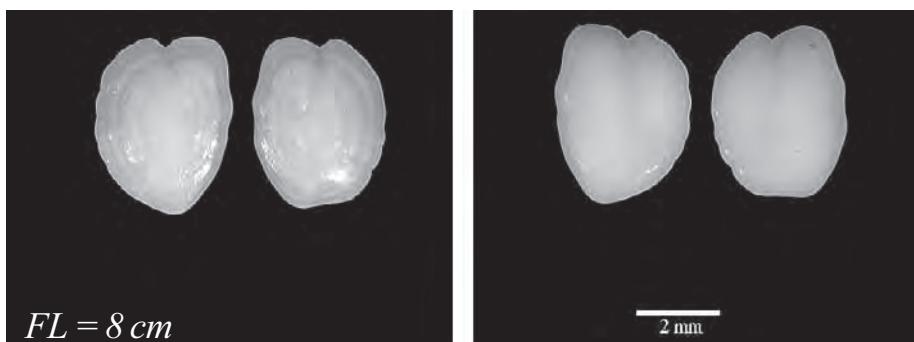
Psychrolutidae: *Cottunculus microps*



Scorpaeniformes

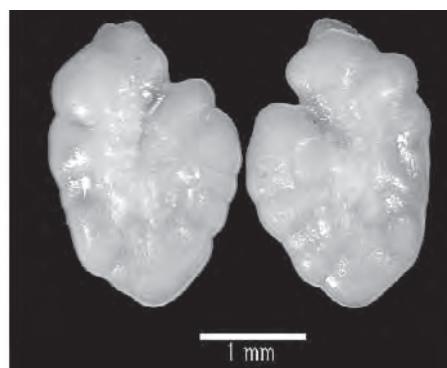
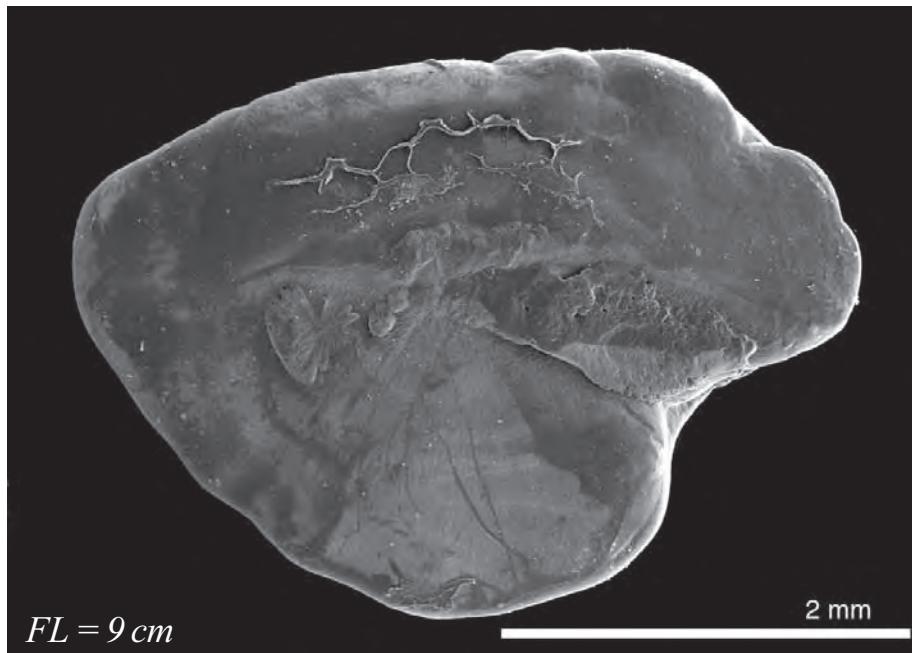
249

Psychrolutidae: *Cottunculus thomsonii*



Scorpaeniformes

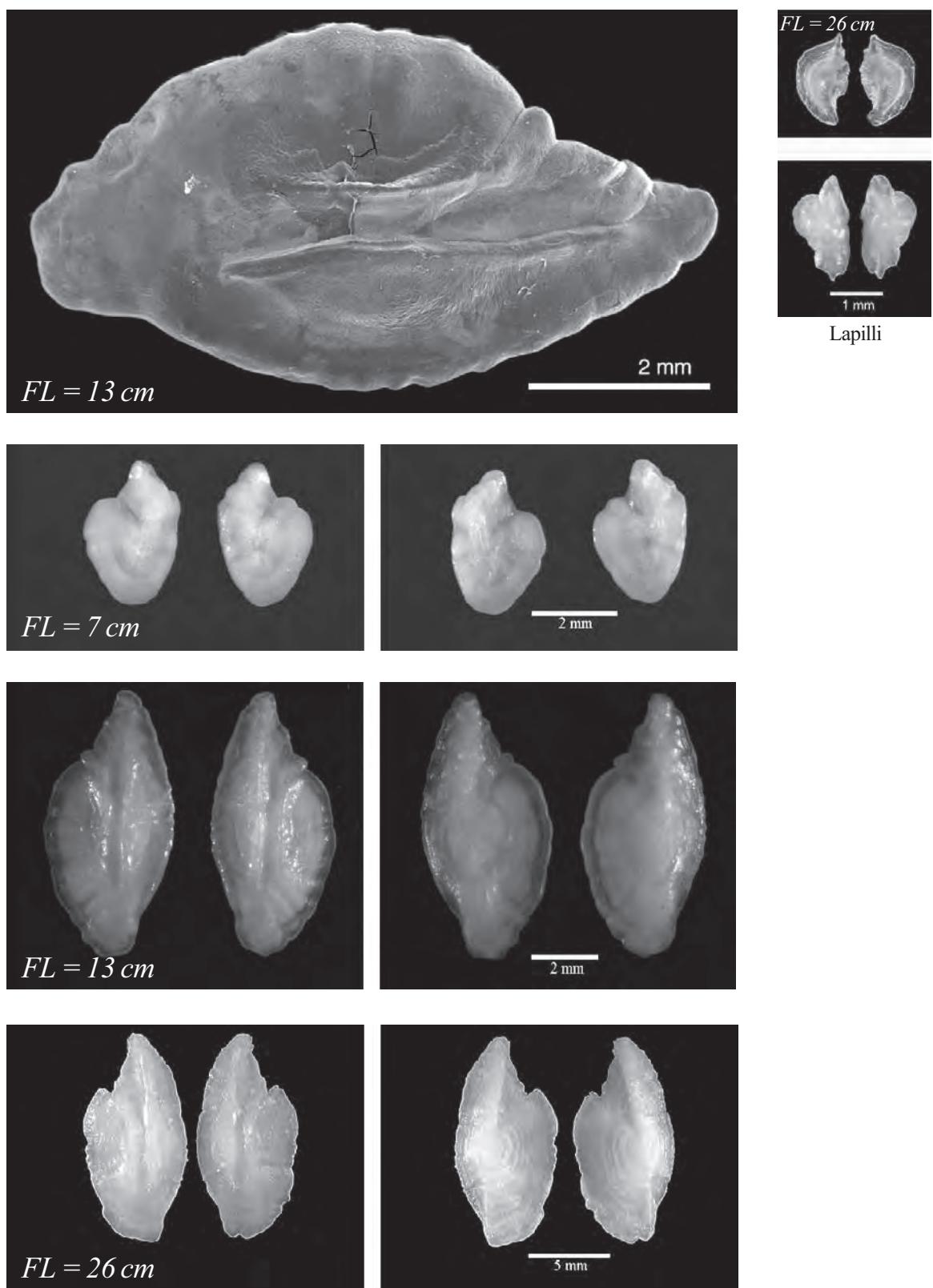
Scorpaenidae: *Etreposebastes imus*



Scorpaeniformes

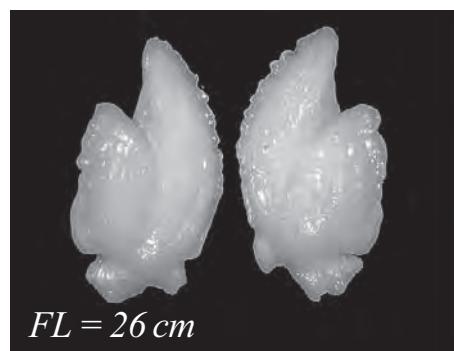
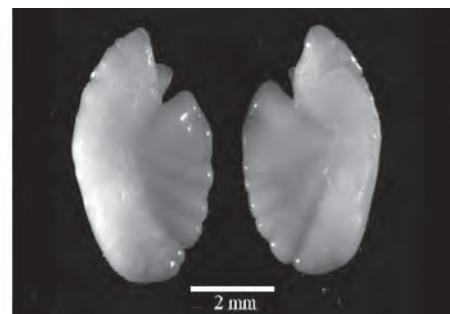
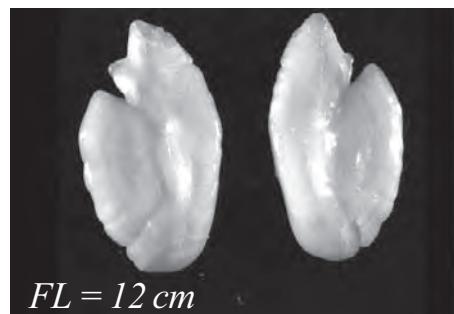
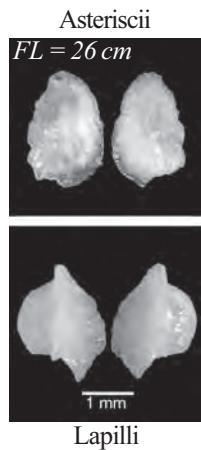
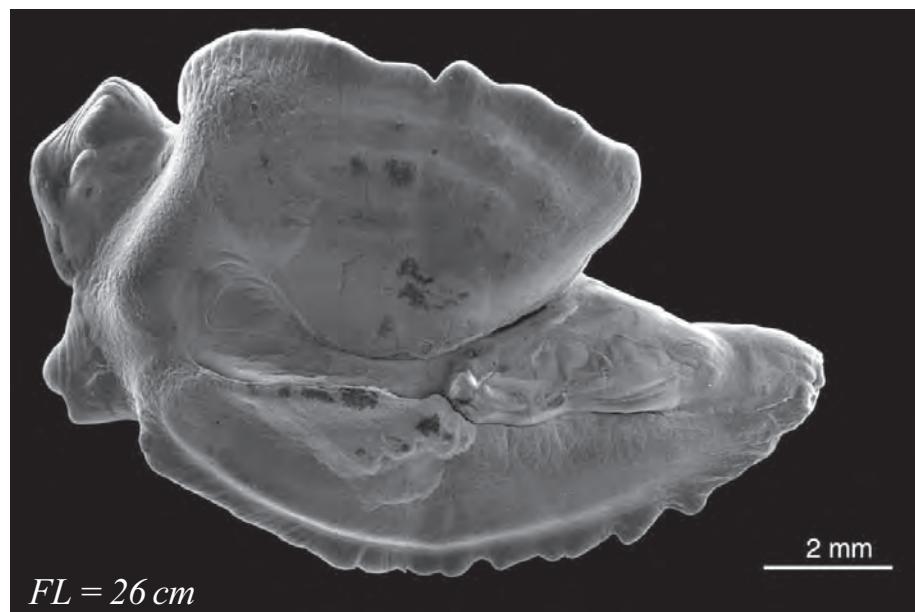
251

Scorpaenidae: *Helicolenus dactylopterus*



Scorpaeniformes

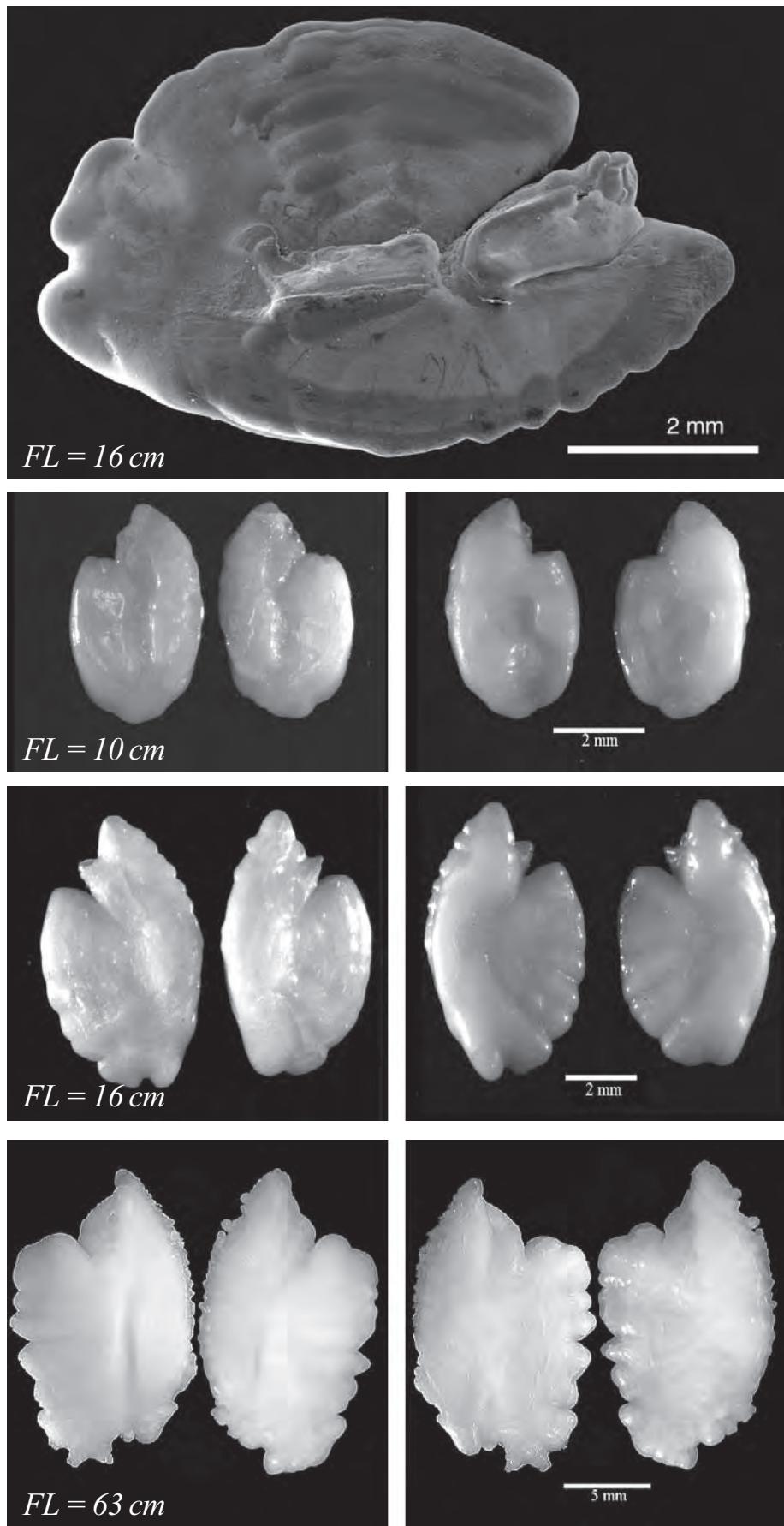
Scorpaenidae: *Sebastes fasciatus*



Scorpaeniformes

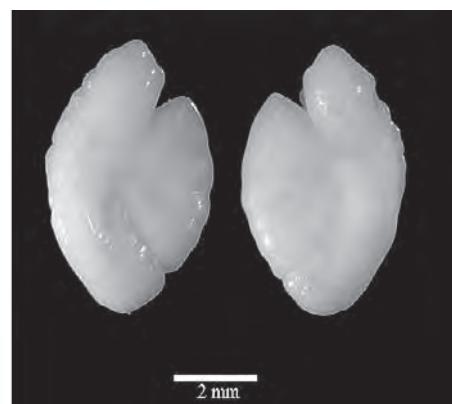
Scorpaenidae: *Sebastes marinus*

253

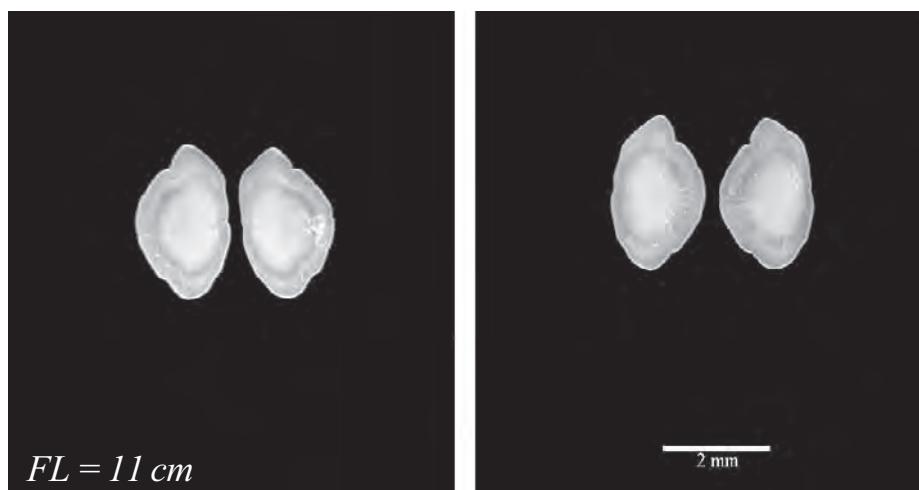
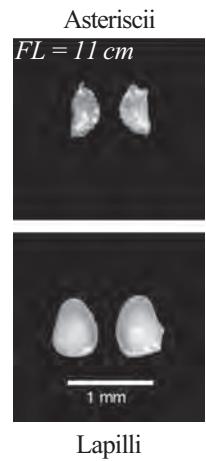
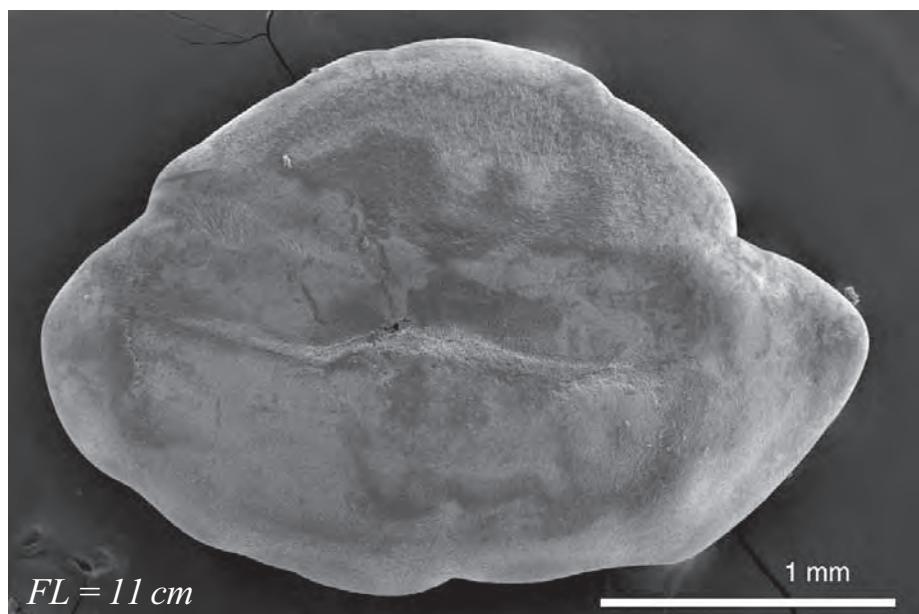


Scorpaeniformes

Scorpaenidae: *Sebastes mentella*

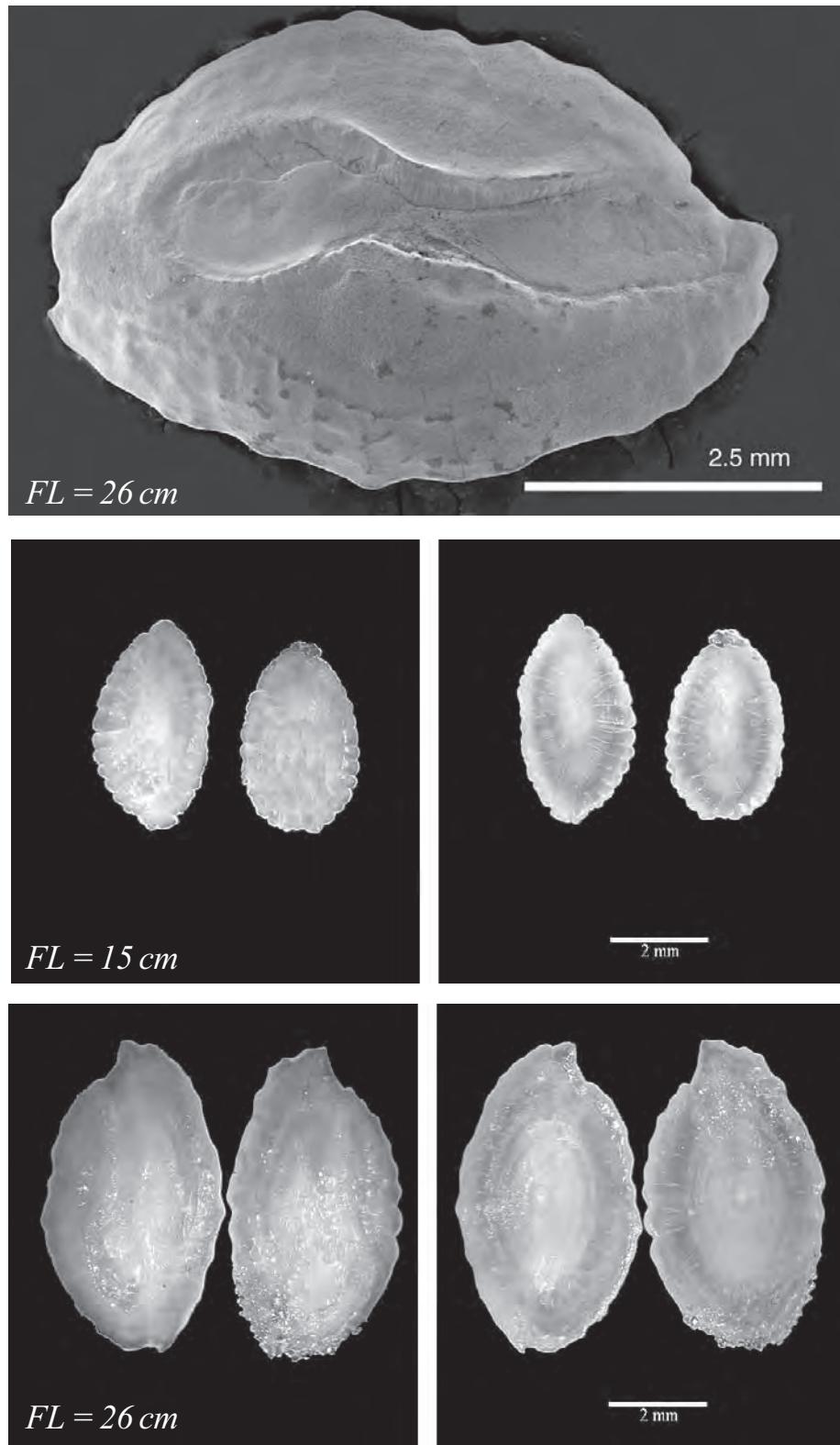


Triglidae: *Peristedion miniatum*



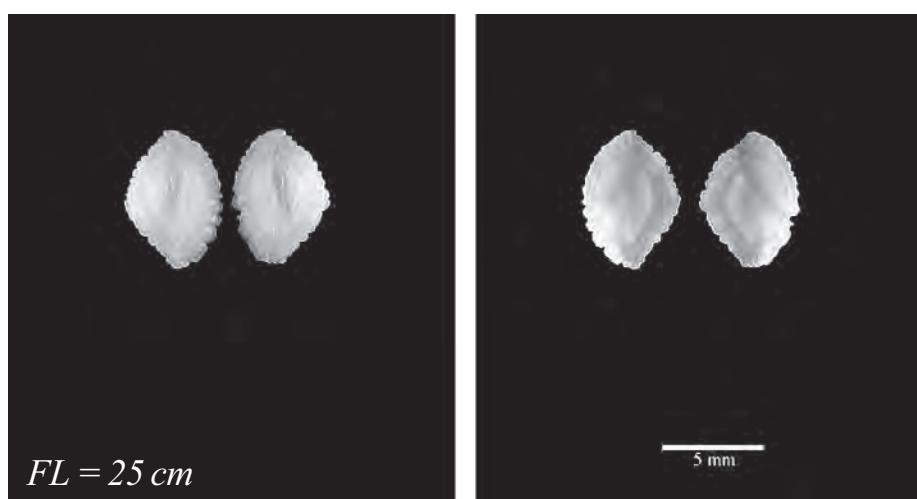
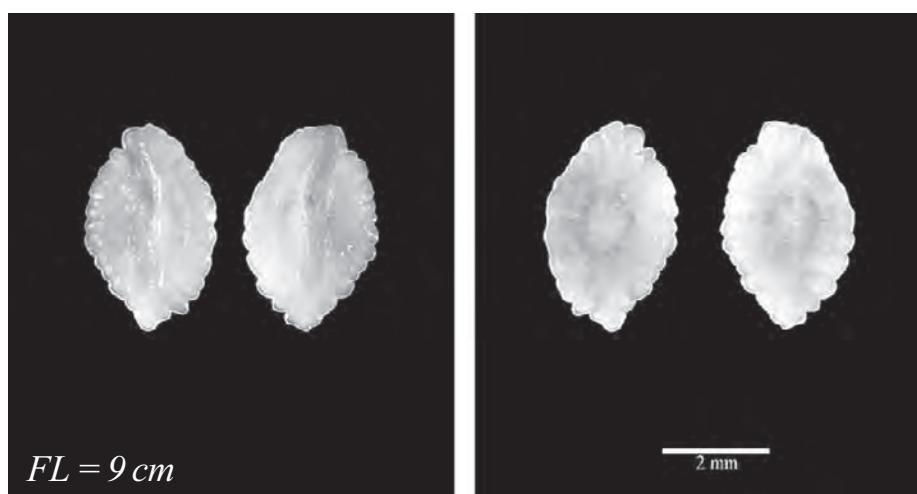
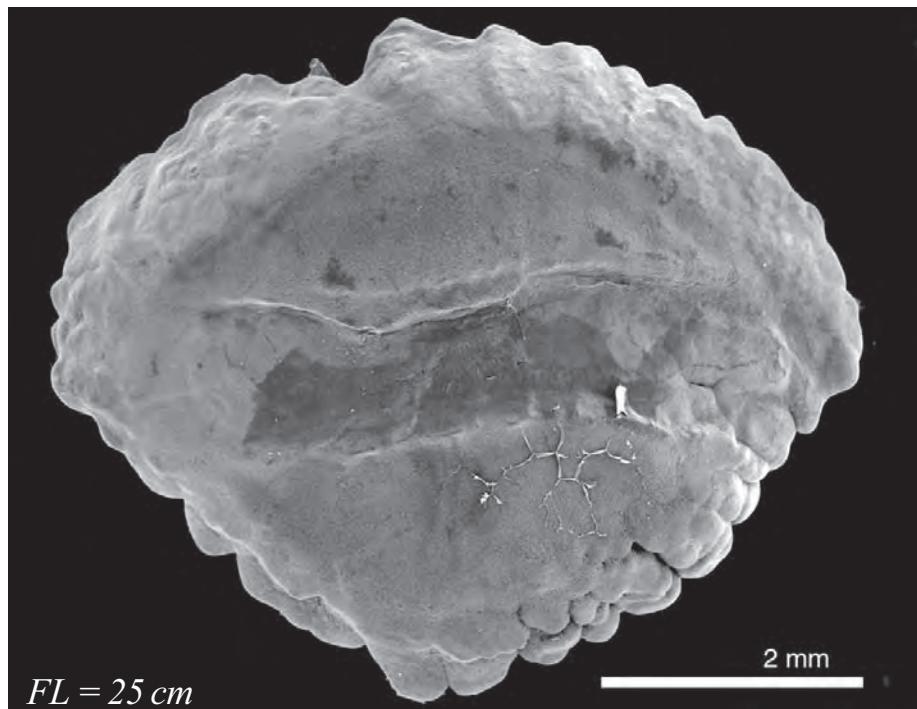
Scorpaeniformes

Triglidae: *Prionotus carolinus*

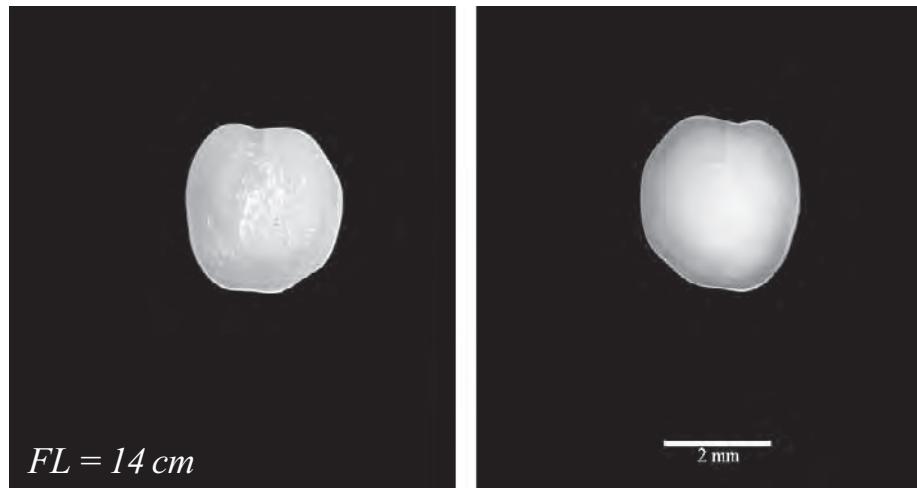
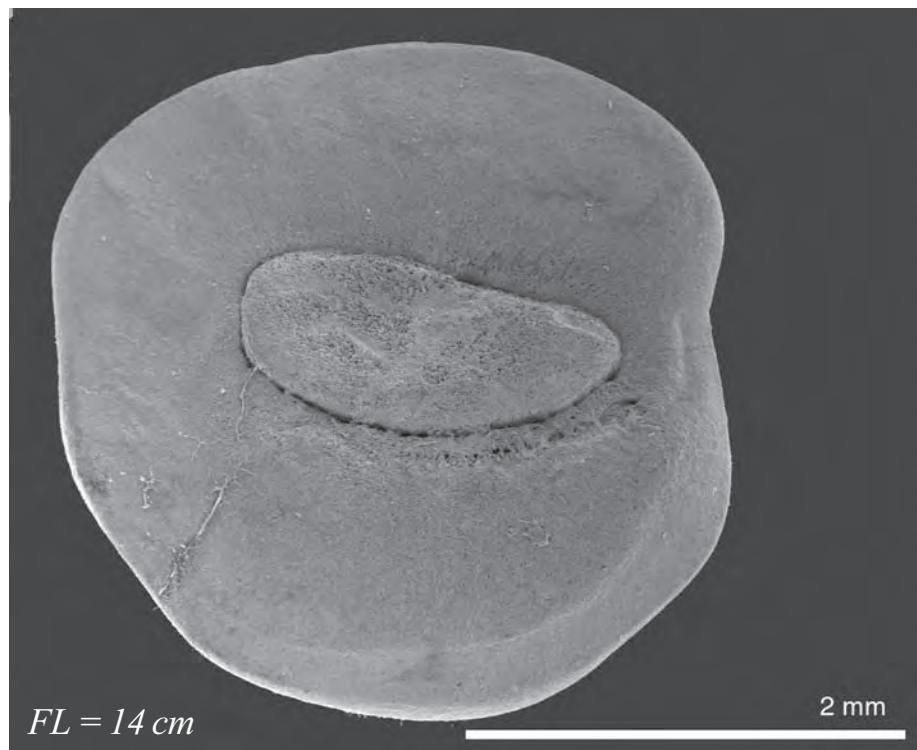


Scorpaeniformes
Triglidae: *Prionotus evolans*

257

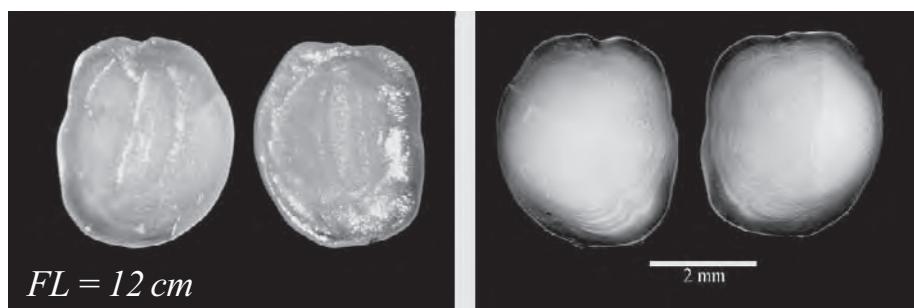
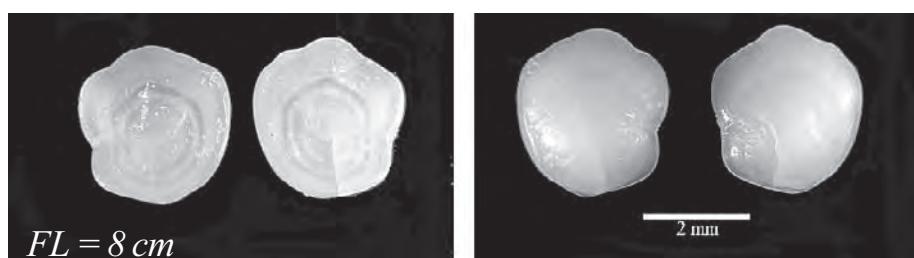
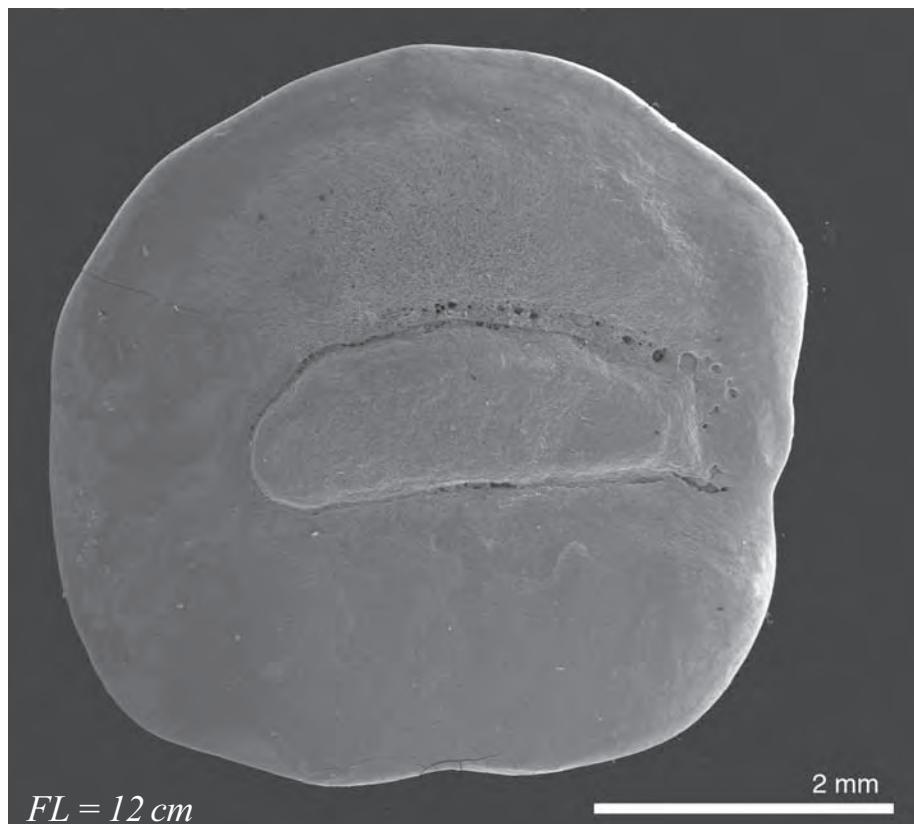


Stephanoberyciformes
Melamphaidae: *Poromitra megalops*

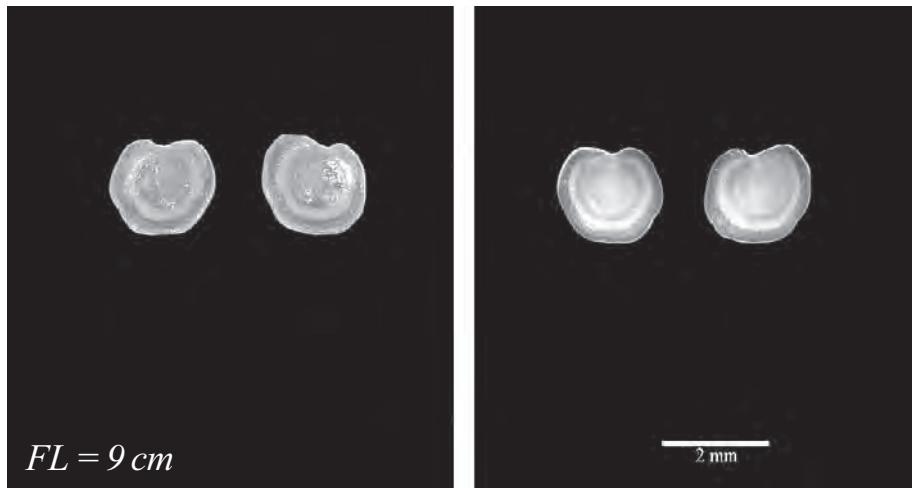
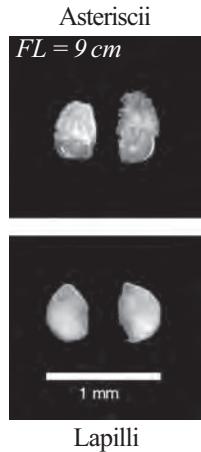
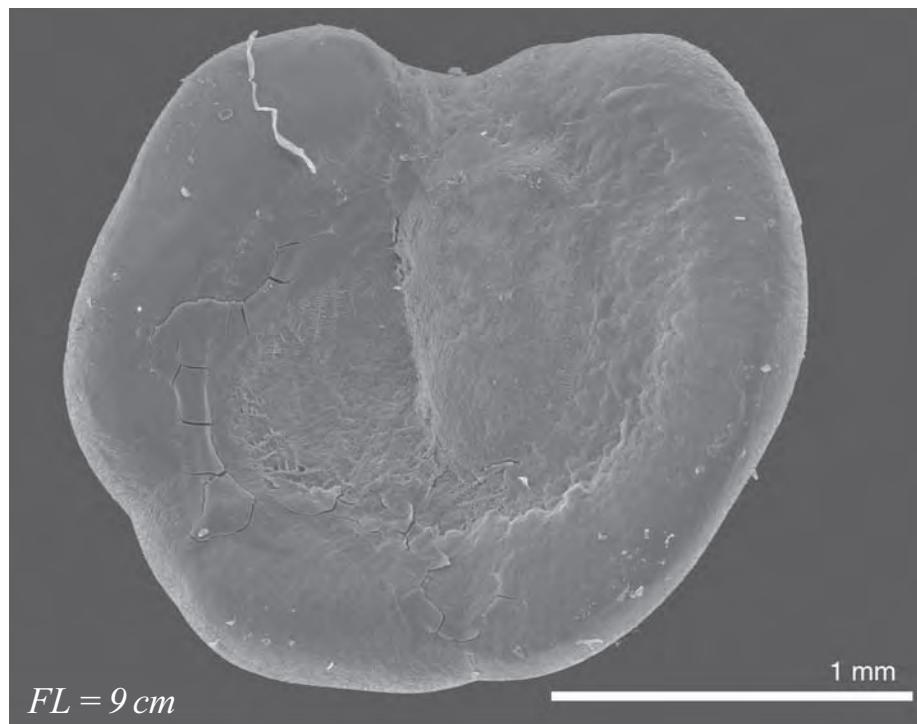


Stephanoberyciformes
Melamphaidae: *Scopelogadus beanii*

259



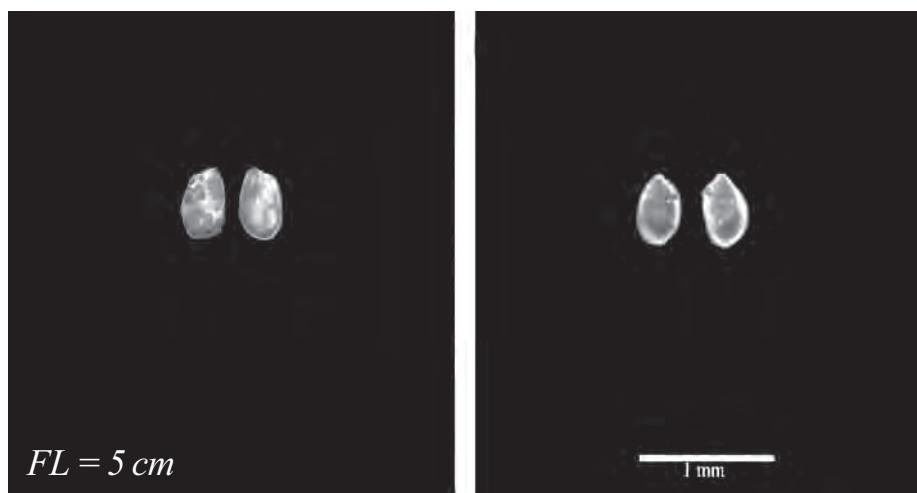
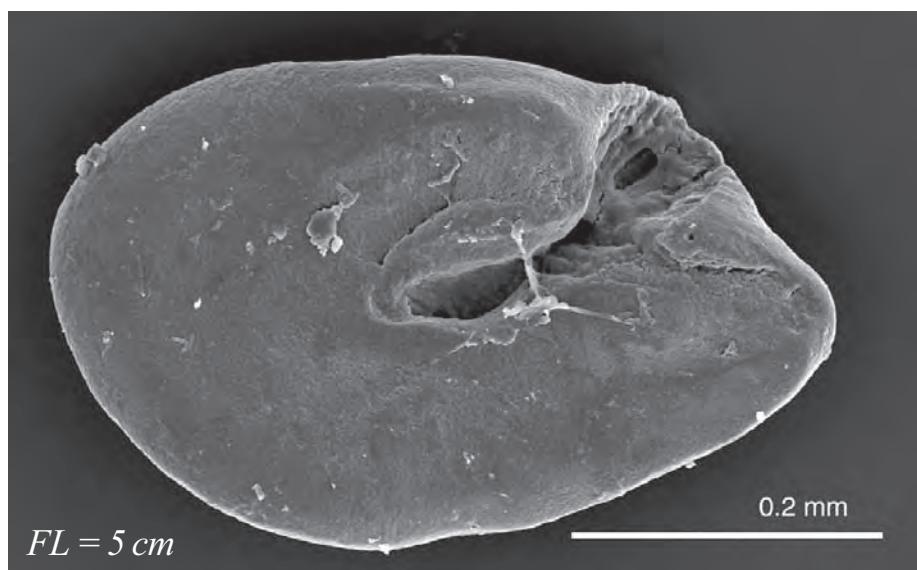
Stephanoberyciformes
Rondeletiidae: *Rondeletia loricata*



Stomiiformes

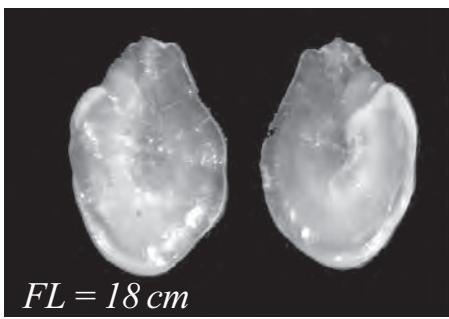
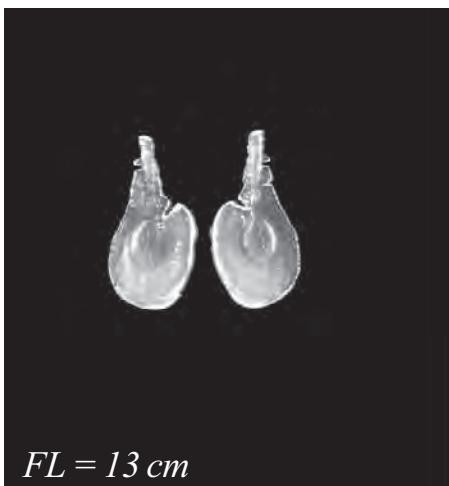
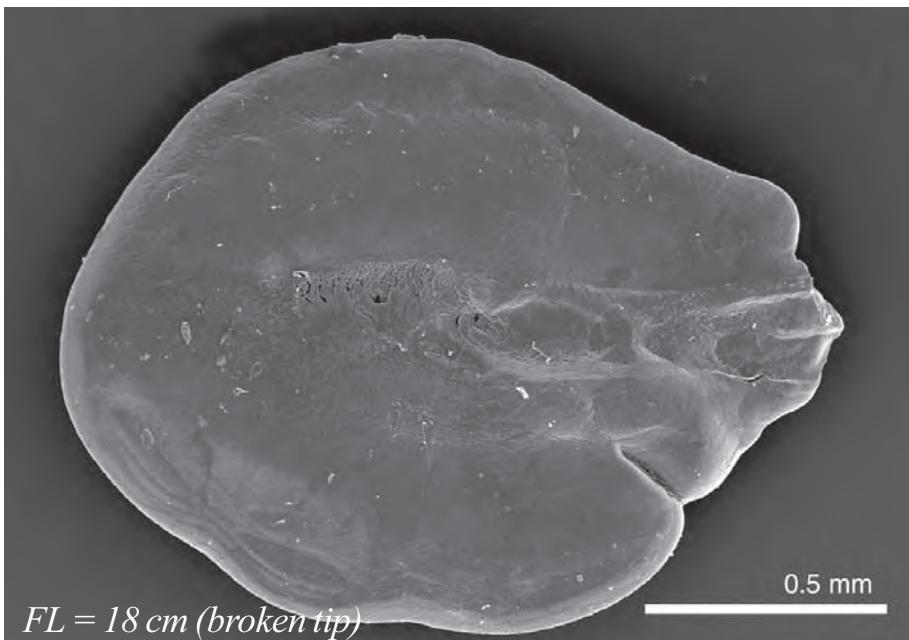
261

Gonostomatidae: *Cyclothona microdon*



Stomiiformes

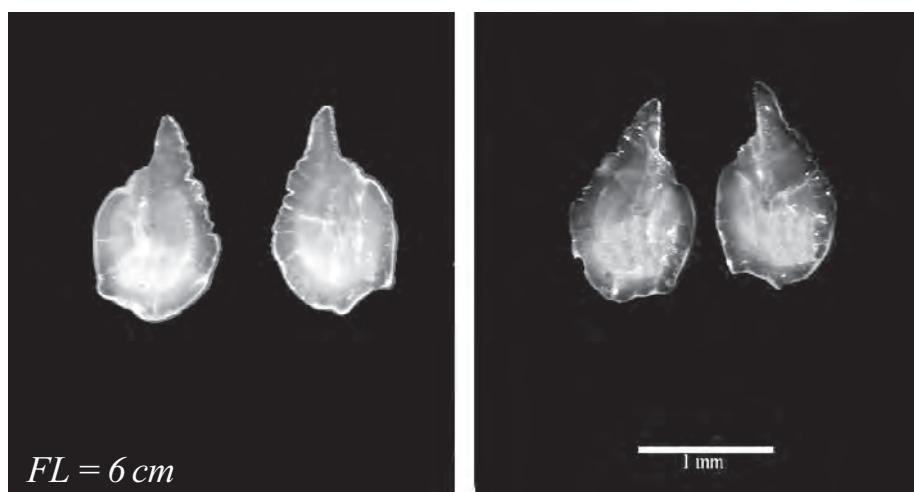
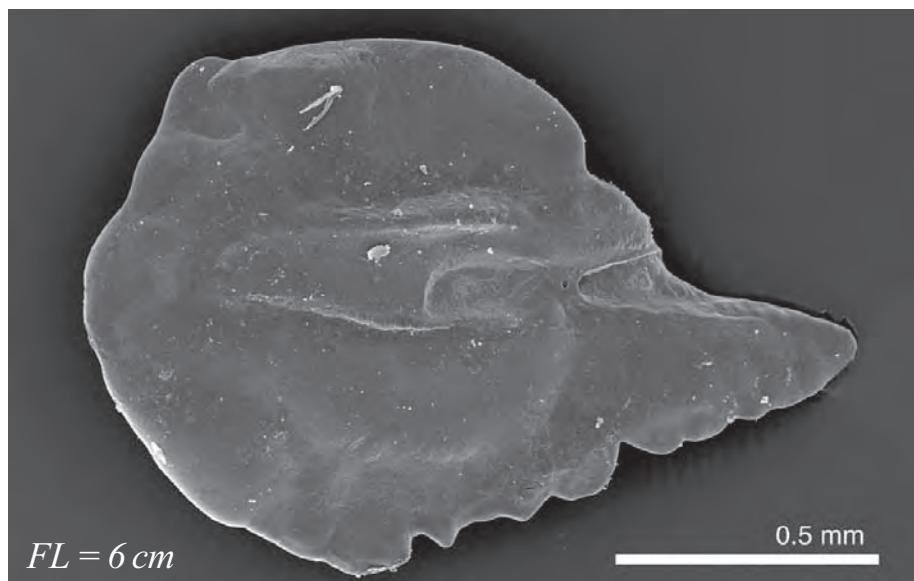
Gonostomatidae: *Gonostoma elongatum*



Stomiiformes

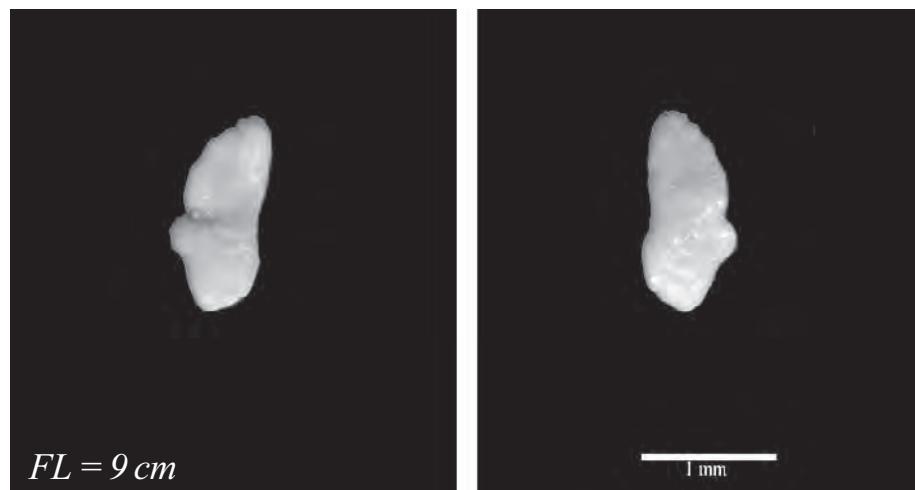
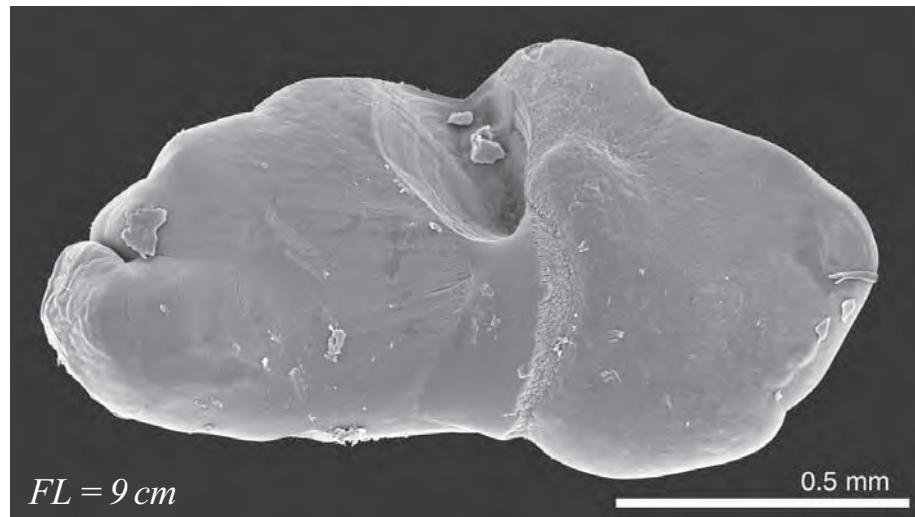
263

Gonostomatidae: *Vinciguerria nimbaria*

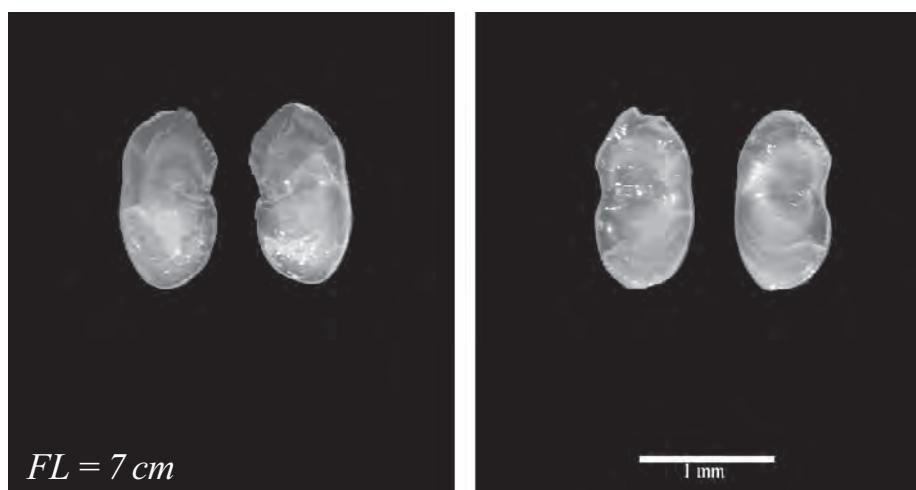
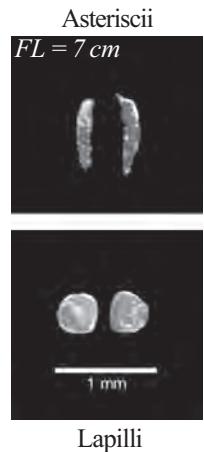
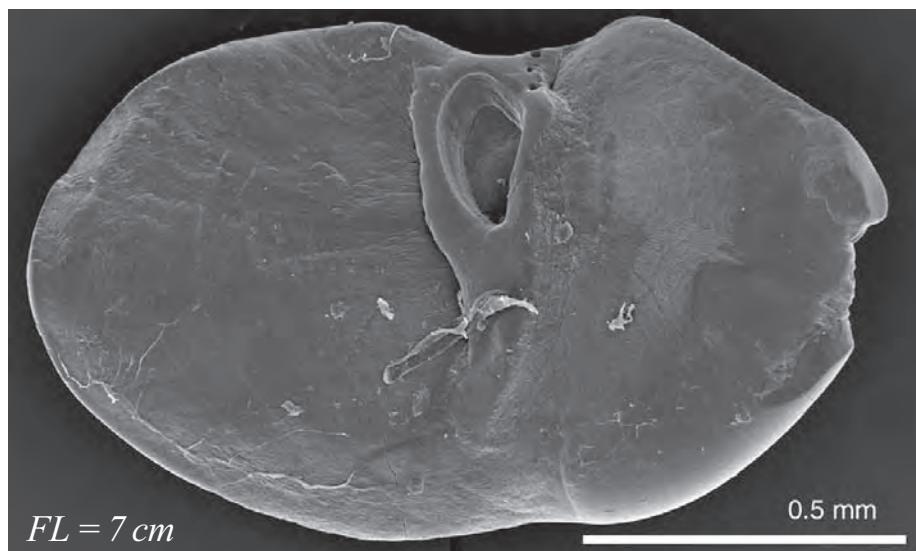


Stomiiformes

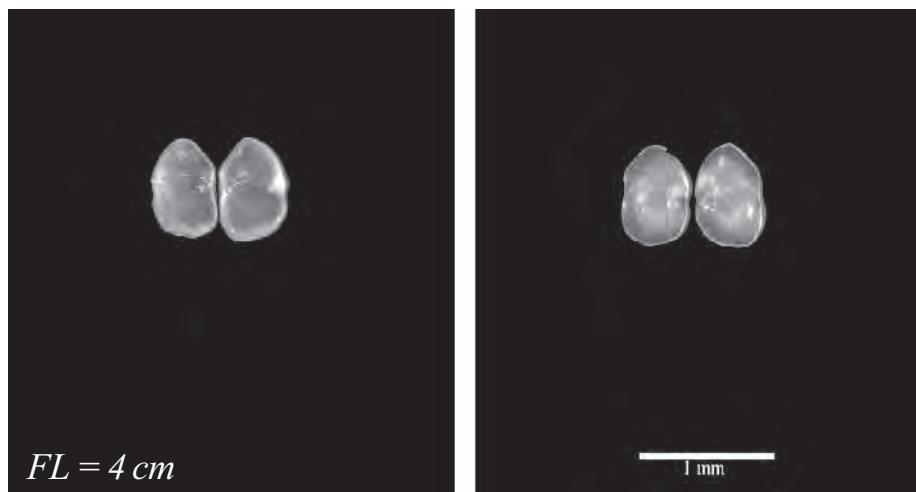
Sternopychidae: *Argyropelecus aculeatus*



Sternopychidae: *Argyropelecus gigas*

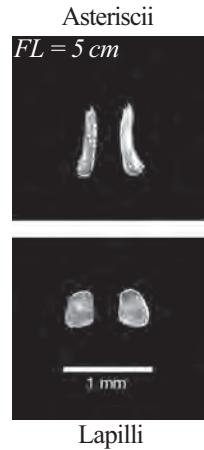
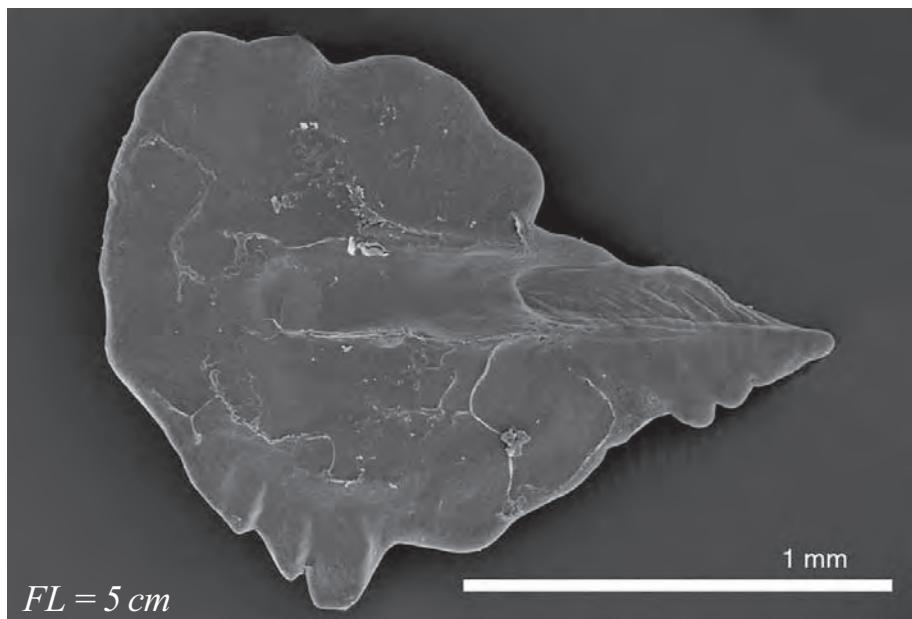


Sternopychidae: *Argyropelecus hemigymnus*



Stomiiformes

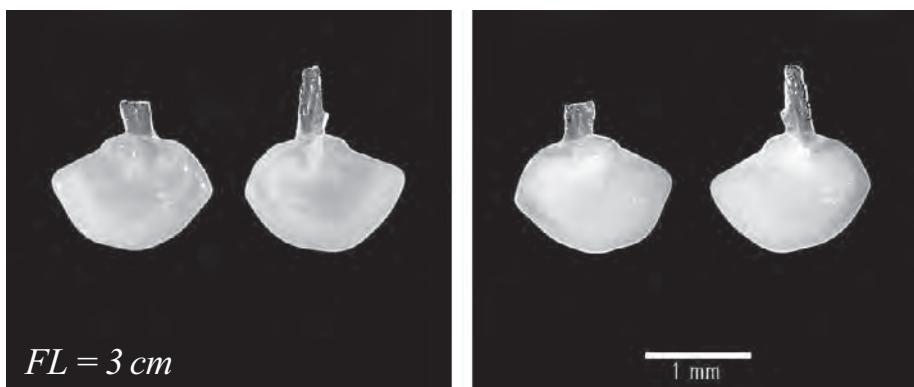
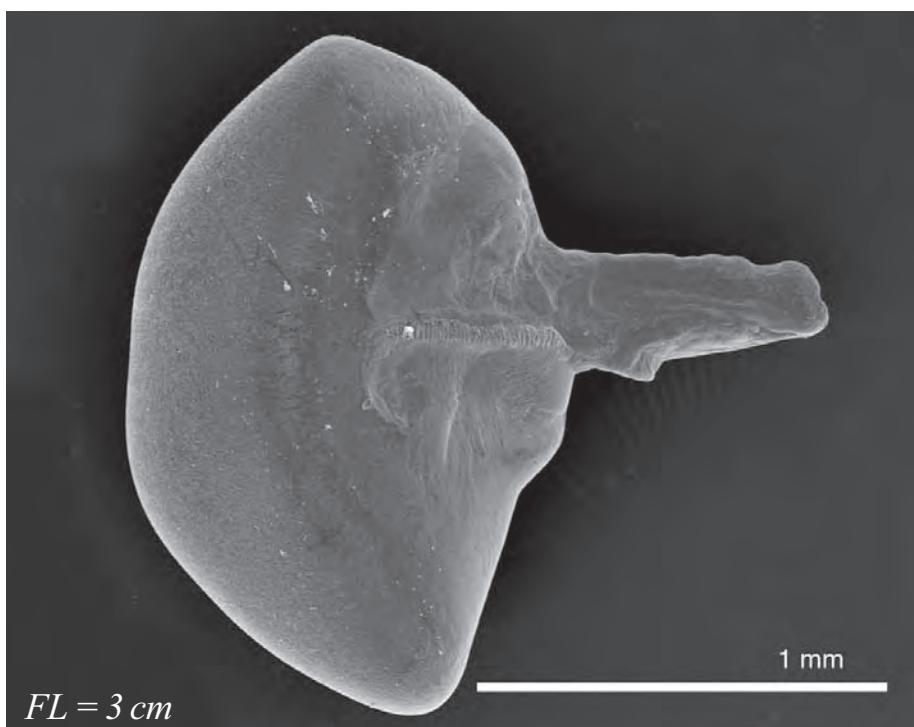
Sternopychidae: *Maurolicus muelleri*



Stomiiformes

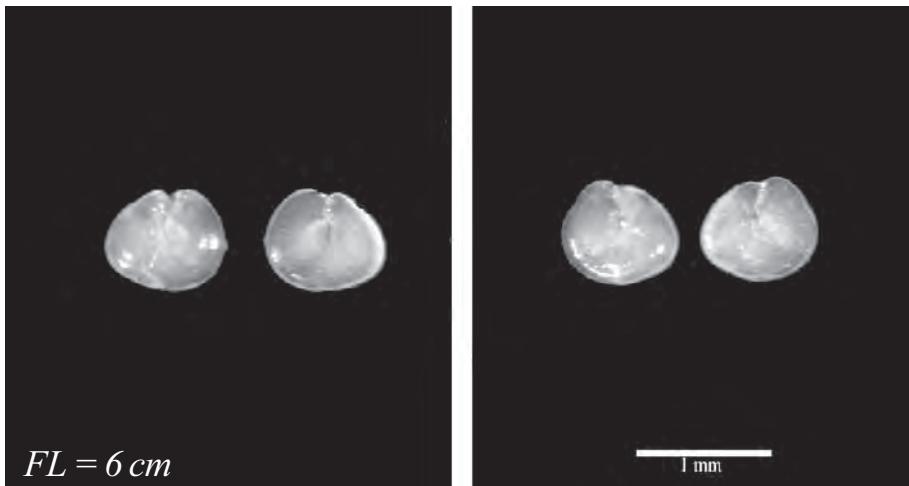
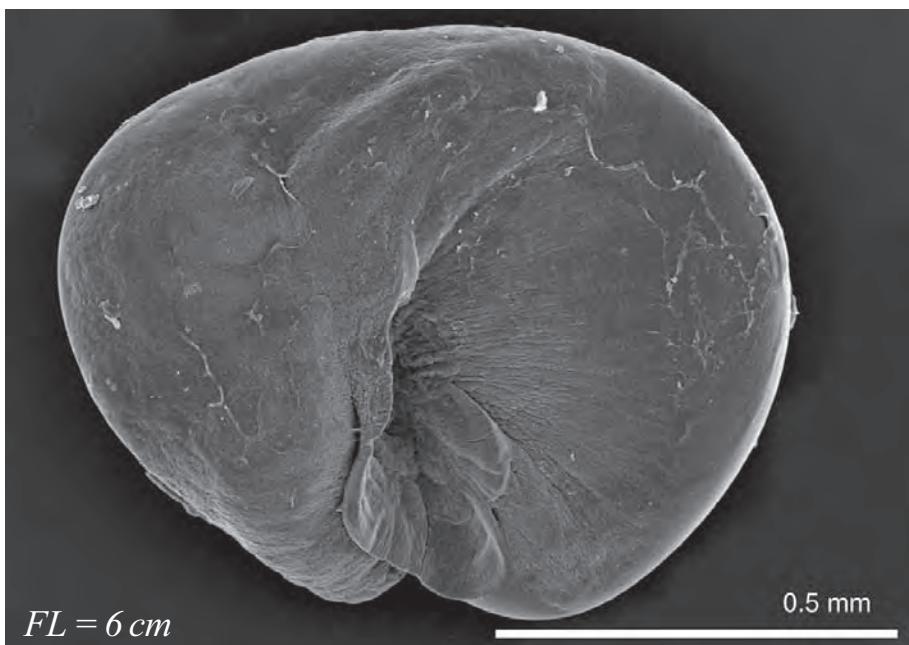
267

Sternoptychidae: *Polyipnus asteroides*



Stomiiformes

Sternopychidae: *Sternopyx diaphana*



Stomiiformes

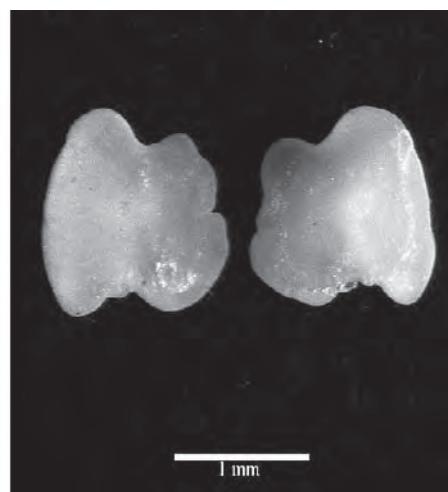
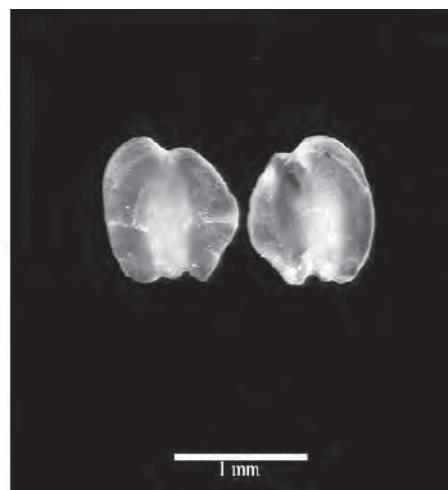
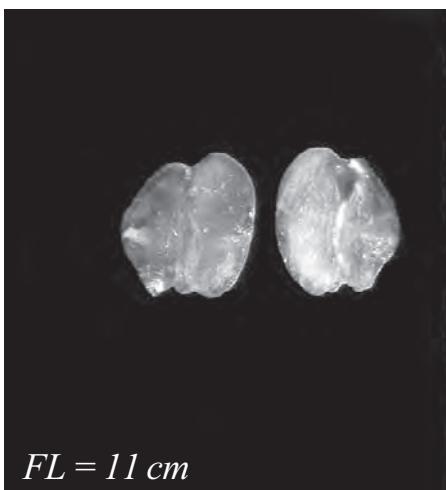
269

Stomiidae: *Chauliodus sloani*



Stomiiformes

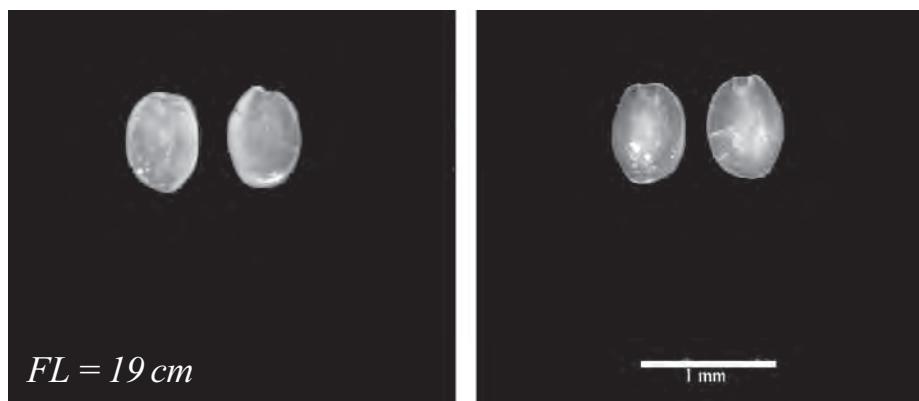
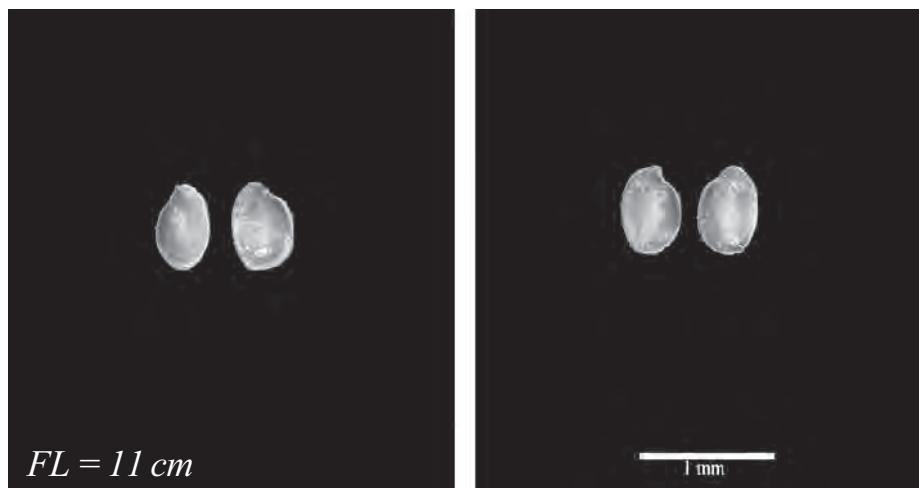
Stomiidae: *Malacosteus niger*



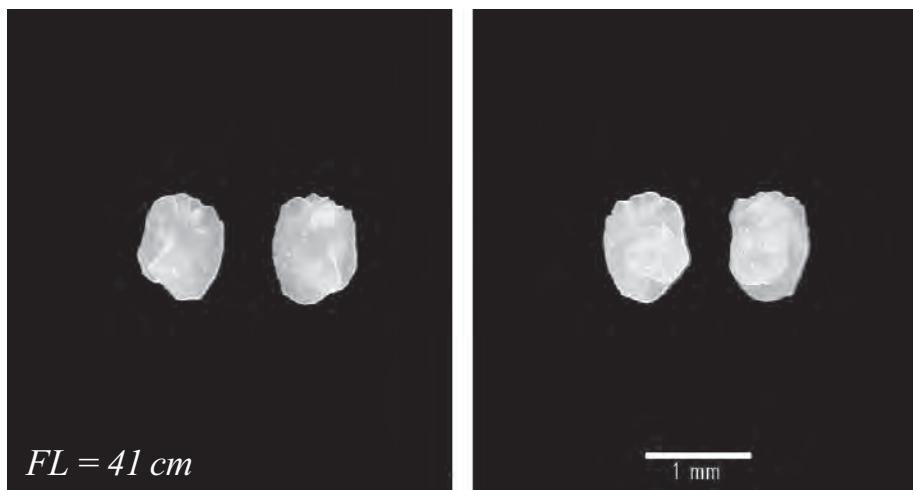
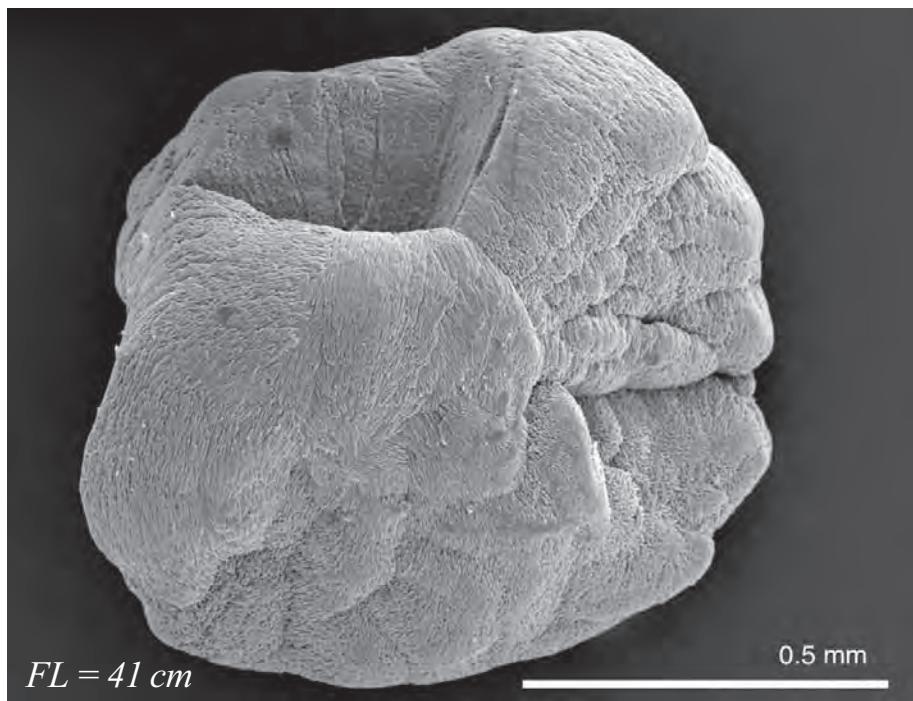
Stomiiformes

271

Stomiidae: *Stomias boa*



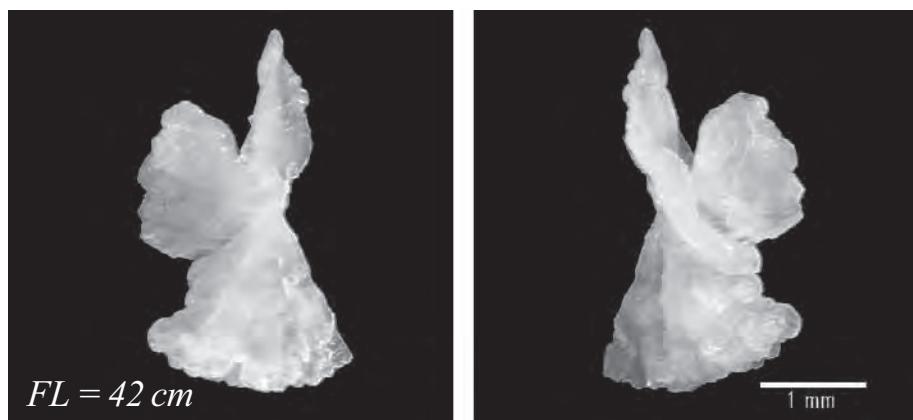
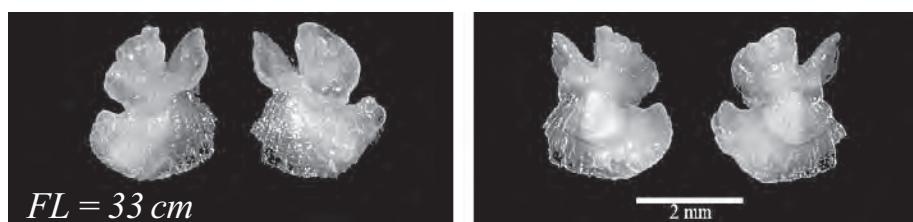
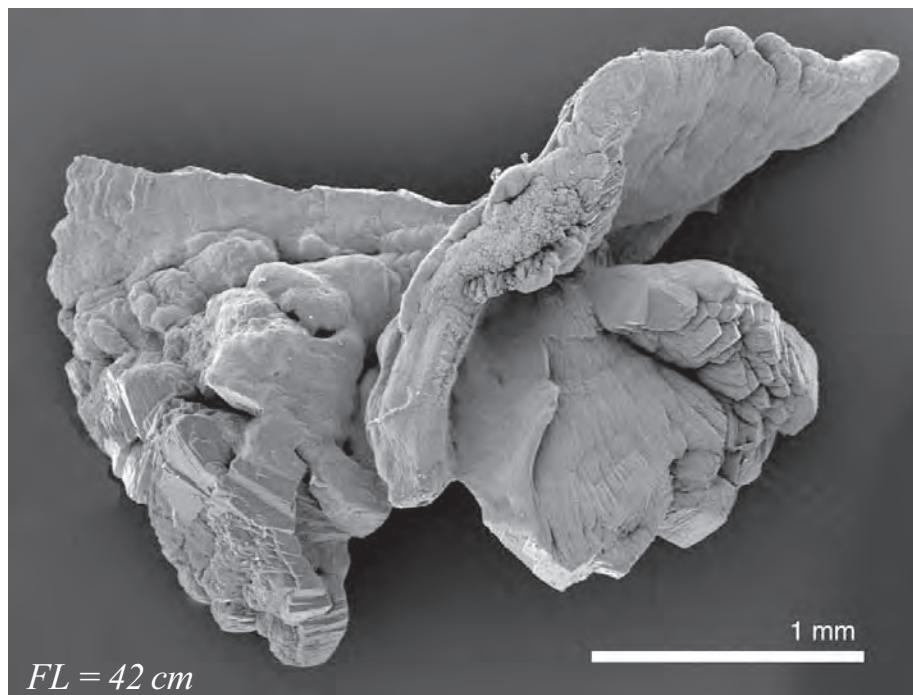
Stomiiformes

Stomiidae: *Trigonolampa miriceps*

Tetraodontiformes

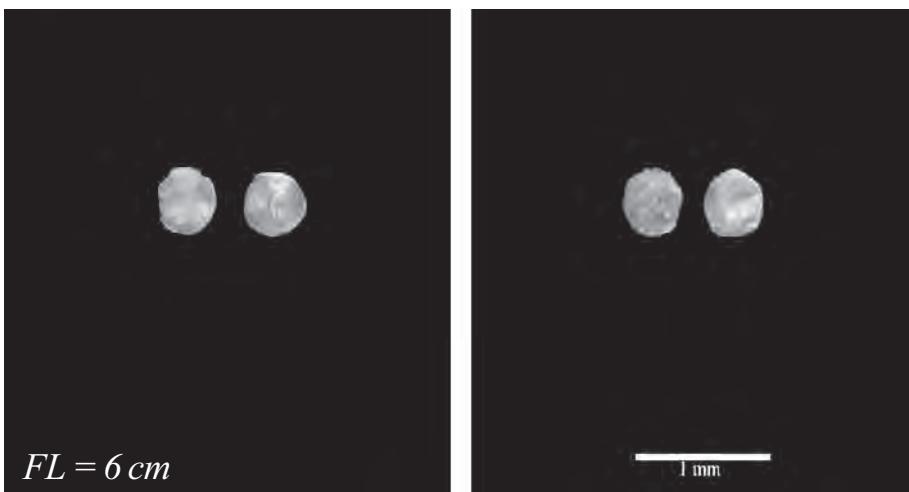
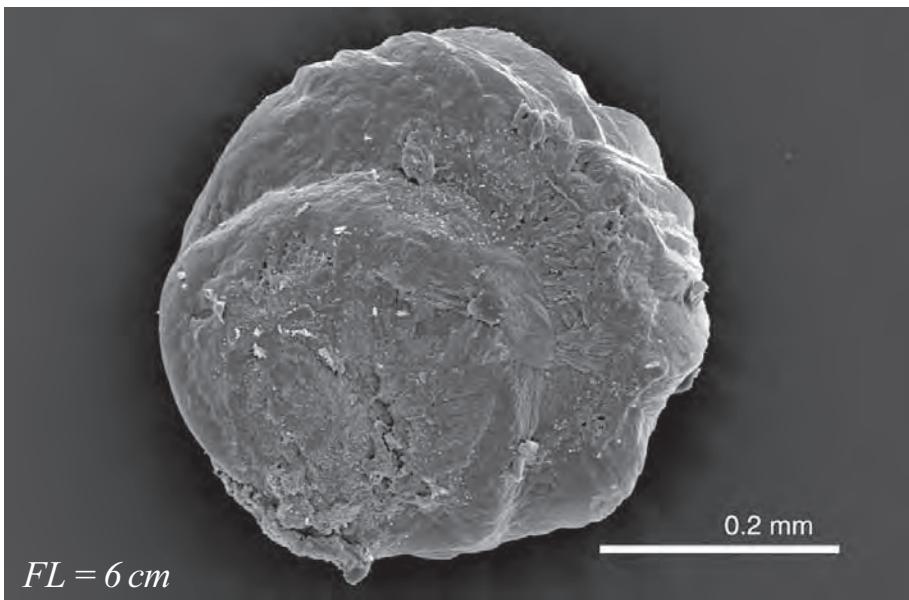
Balistidae: *Balistes capriscus*

273



Tetraodontiformes

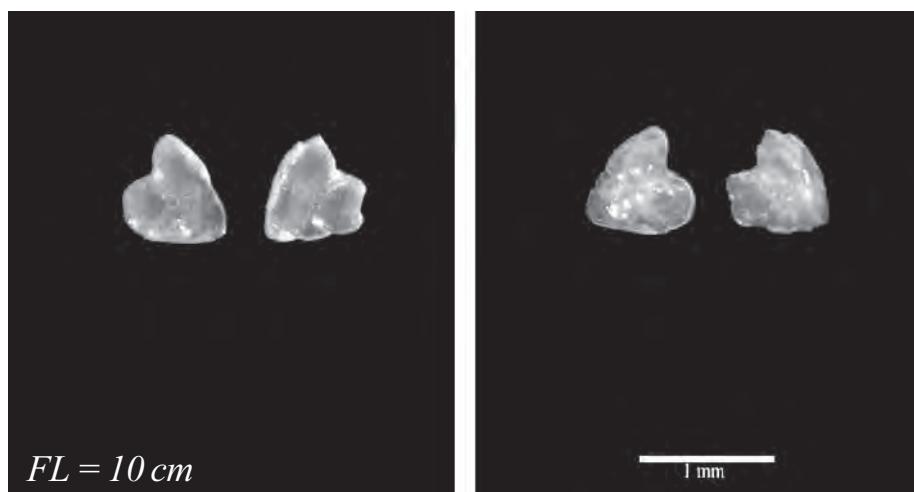
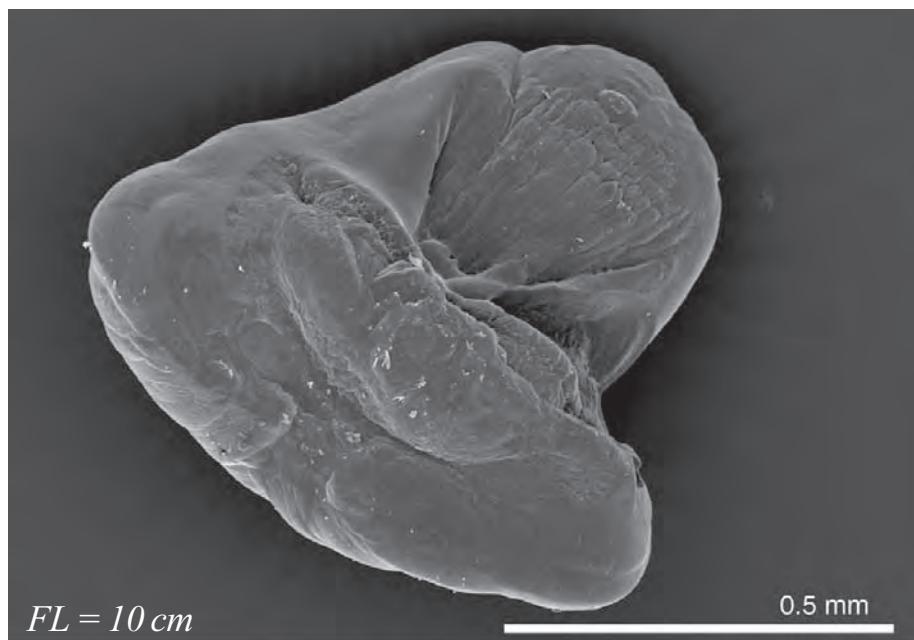
Diodontidae: *Chilomycterus schoepfi*



Tetraodontiformes

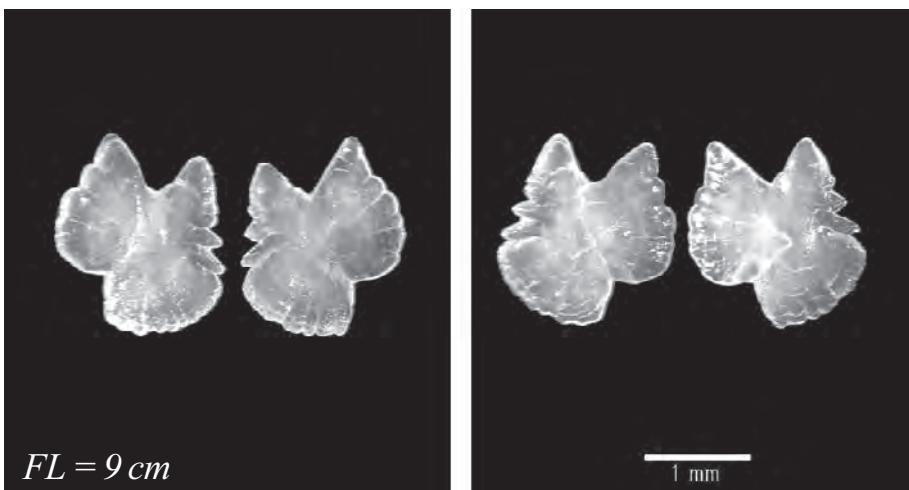
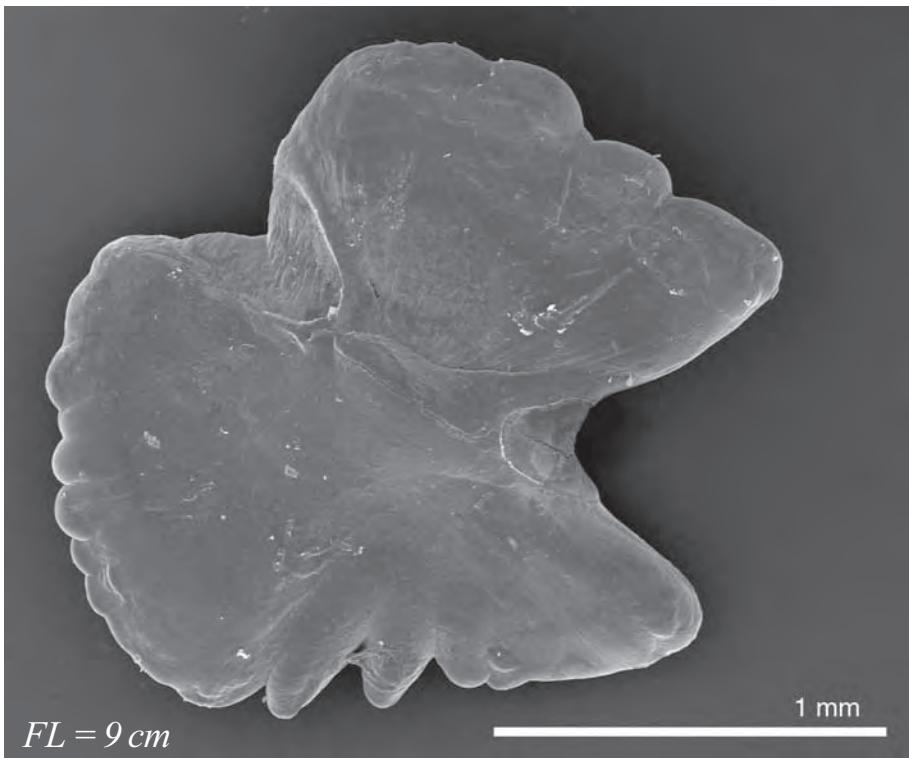
275

Tetraodontidae: *Sphoeroides maculatus*

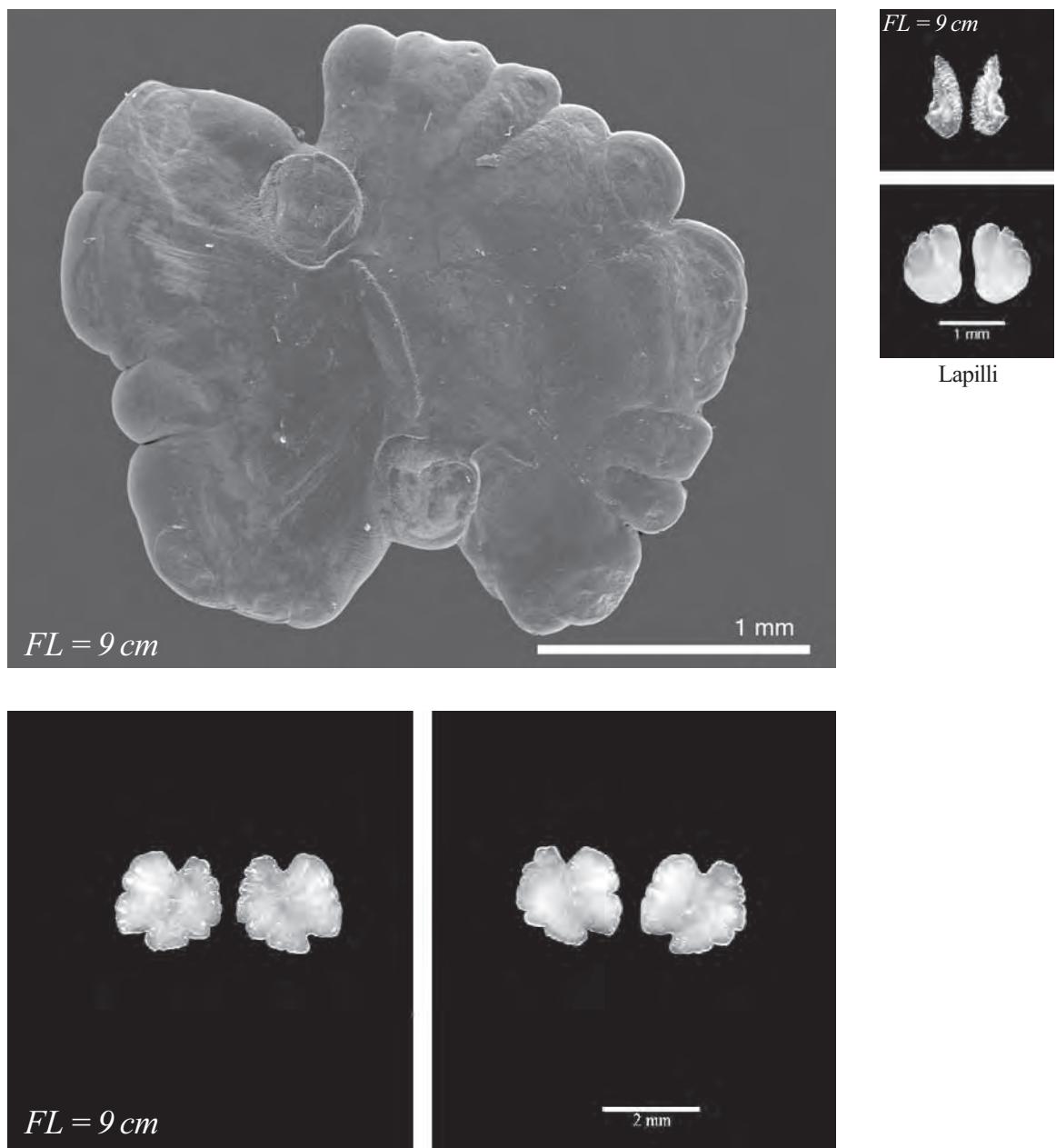


Zeiformes

Grammicolepididae: *Daramattus americanus*

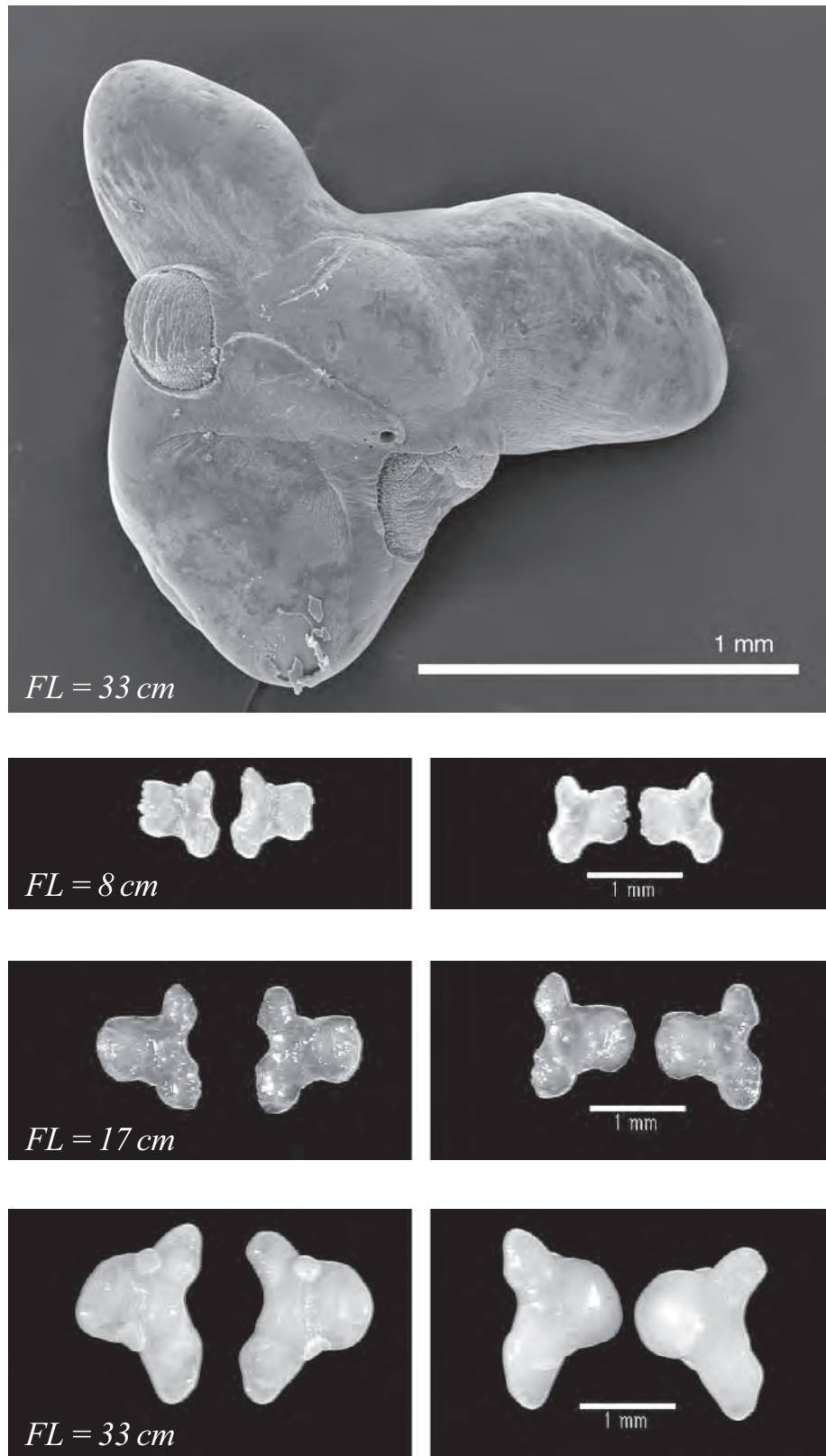


Zeidae: *Cytopsis roseus*



Zeiformes

Zeidae: *Zenopsis conchifera*



Alphabetical species list

Species	Family	Order	Page number
<i>Acanthocybium solandri</i>	Scombridae	Perciformes	172
<i>Acipenser oxyrinchus</i>	Acipenseridae	Acipenseriformes	15
<i>Agonus decagonus</i>	Agonidae	Scorpaeniformes	225
<i>Alepisaurus brevirostris</i>	Alepisauridae	Aulopiformes	30
<i>Alepisaurus ferox</i>	Alepisauridae	Aulopiformes	31
<i>Alepocephalus agassizii</i>	Alepocephalidae	Osmeriformes	128
<i>Alepocephalus bairdii</i>	Alepocephalidae	Osmeriformes	129
<i>Alosa aestivalis</i>	Clupeidae	Clupeiformes	43
<i>Alosa pseudoharengus</i>	Clupeidae	Clupeiformes	44
<i>Alosa sapidissima</i>	Clupeidae	Clupeiformes	45
<i>Ammodytes americanus/dubius</i>	Ammodytidae	Perciformes	143
<i>Anarhichas denticulatus</i>	Anarhichadidae	Perciformes	144
<i>Anarhichas lupus</i>	Anarhichadidae	Perciformes	145
<i>Anarhichas minor</i>	Anarhichadidae	Perciformes	146
<i>Anguilla rostrata</i>	Anguillidae	Anguilliformes	20
<i>Anoplogaster cornuta</i>	Anoplogasteridae	Beryciformes	40
<i>Antimora rostrata</i>	Moridae	Gadiformes	70
<i>Apeltes quadratus</i>	Gasterosteidae	Gasterosteiformes	82
<i>Aphanopus carbo</i>	Trichiuridae	Perciformes	190
<i>Argentina silus</i>	Argentinidae	Osmeriformes	134
<i>Argyropelecus aculeatus</i>	Sternopychidae	Stomiiformes	264
<i>Argyropelecus gigas</i>	Sternopychidae	Stomiiformes	265
<i>Argyropelecus hemigymnus</i>	Sternopychidae	Stomiiformes	265
<i>Artemiellus atlanticus</i>	Cottidae	Scorpaeniformes	228
<i>Aspidophoroides monopterygius</i>	Agonidae	Scorpaeniformes	226
<i>Aspidophoroides olrikii</i>	Agonidae	Scorpaeniformes	227
<i>Bajacalifornia megalops</i>	Alepocephalidae	Osmeriformes	130
<i>Balistes capriscus</i>	Balistidae	Tetraodontiformes	273
<i>Bathylagus euryops</i>	Bathylagidae	Osmeriformes	135
<i>Bathypterois quadrifilis</i>	Ipnopidae	Aulopiformes	34
<i>Bathysaurus ferox</i>	Synodontidae	Aulopiformes	38
<i>Benthodesmus elongatus</i>	Trichiuridae	Perciformes	191
<i>Benthosema glaciale</i>	Myctophidae	Myctophiformes	96
<i>Benthosema suborbitale</i>	Myctophidae	Myctophiformes	97
<i>Bolinichthys photothorax</i>	Myctophidae	Myctophiformes	97
<i>Boreogadus saida</i>	Gadidae	Gadiformes	51
<i>Brama brama</i>	Bramidae	Perciformes	147
<i>Brevoortia tyrannus</i>	Clupeidae	Clupeiformes	46
<i>Brosme brosme</i>	Gadidae	Gadiformes	52
<i>Brosmiculus imberbis</i>	Moridae	Gadiformes	71
<i>Callionymus agassizi</i>	Callionymidae	Perciformes	148
<i>Caranx cryos</i>	Carangidae	Perciformes	149
<i>Caranx hippos</i>	Carangidae	Perciformes	149
<i>Careproctus reinhardtii</i>	Liparidae	Scorpaeniformes	242
<i>Caristius groenlandicus</i>	Caristiidae	Perciformes	154
<i>Centropristes striata</i>	Serranidae	Perciformes	182
<i>Ceratias holboelli</i>	Ceratiidae	Lophiiformes	90
<i>Ceratoscopelus maderensis</i>	Myctophidae	Myctophiformes	98
<i>Ceratoscopelus warmingii</i>	Myctophidae	Myctophiformes	98
<i>Chalinura brevibarbis</i>	Macrouridae	Gadiformes	61
<i>Chauliodus sloani</i>	Stomiidae	Stomiiformes	269

Species	Family	Order	Page number
<i>Chiasmodon niger</i>	Chiasmodontidae	Perciformes	156
<i>Chilomycterus schoepfi</i>	Diodontidae	Tetraodontiformes	274
<i>Chlorophthalmus agassizi</i>	Chlorophthalmidae	Aulopiformes	32
<i>Citharichthys arctifrons</i>	Paralichthyidae	Pleuronectiformes	204
<i>Clupea harengus harengus</i>	Clupeidae	Clupeiformes	47
<i>Coregonus clupeaformis</i>	Salmonidae	Salmoniformes	218
<i>Coregonus huntsmani</i>	Salmonidae	Salmoniformes	219
<i>Coryphaena hippurus</i>	Coryphaenidae	Perciformes	157
<i>Coryphaenoides guentheri</i>	Macrouridae	Gadiformes	61
<i>Coryphaenoides rupestris</i>	Macrouridae	Gadiformes	62
<i>Cottunculus microps</i>	Psychrolutidae	Scorpaeniformes	248
<i>Cottunculus thomsonii</i>	Psychrolutidae	Scorpaeniformes	249
<i>Cryptacanthodes maculatus</i>	Cryptacanthodidae	Perciformes	158
<i>Cryptopsaras couesi</i>	Ceratiidae	Lophiiformes	91
<i>Cyclopterus lumpus</i>	Cyclopteridae	Scorpaeniformes	239
<i>Cyclothona microdon</i>	Gonostomatidae	Stomiiformes	261
<i>Cytopsis roseus</i>	Zeidae	Zeiformes	277
<i>Daramattus americanus</i>	Grammicolepididae	Zeiformes	276
<i>Decapterus macarellus</i>	Carangidae	Perciformes	150
<i>Derichthys serpentinus</i>	Derichthyidae	Anguilliformes	21
<i>Diaphus dumerili</i>	Myctophidae	Myctophiformes	99
<i>Diaphus effulgens</i>	Myctophidae	Myctophiformes	100
<i>Diaphus metopoclampus</i>	Myctophidae	Myctophiformes	101
<i>Diaphus mollis</i>	Myctophidae	Myctophiformes	102
<i>Diaphus perspicillatus</i>	Myctophidae	Myctophiformes	103
<i>Diaphus rafinesquii</i>	Myctophidae	Myctophiformes	104
<i>Diaphus termophilus</i>	Myctophidae	Myctophiformes	105
<i>Dibranchus atlanticus</i>	Ogcocephalidae	Lophiiformes	93
<i>Dicrolene intronigra</i>	Ophidiidae	Ophidiiformes	125
<i>Diogenichthys atlanticus</i>	Myctophidae	Myctophiformes	105
<i>Echeneis naucrates</i>	Echeneidae	Perciformes	159
<i>Ectrepobaste imus</i>	Scorpaenidae	Scorpaeniformes	250
<i>Electrona risso</i>	Myctophidae	Myctophiformes	106
<i>Enchelyopus cimbrius</i>	Phycidae	Gadiformes	75
<i>Epigonus telescopus</i>	Epigonidae	Perciformes	160
<i>Epinephelus niveatus</i>	Serranidae	Perciformes	183
<i>Etropus microstomus</i>	Paralichthyidae	Pleuronectiformes	205
<i>Etrumeus teres</i>	Clupeidae	Clupeiformes	48
<i>Eumesogrammus praecisus</i>	Stichaeidae	Perciformes	185
<i>Eumicrotremus spinosus</i>	Cyclopteridae	Scorpaeniformes	240
<i>Eurypharynx pelecanoides</i>	Eurypharyngidae	Saccopharyngiformes	217
<i>Fundulus diaphanus</i>	Fundulidae	Cyprinodontiformes	49
<i>Fundulus heteroclitus</i>	Fundulidae	Cyprinodontiformes	50
<i>Gadus morhua</i>	Gadidae	Gadiformes	53
<i>Gadus ogac</i>	Gadidae	Gadiformes	54
<i>Gaidropsarус argentatus</i>	Phycidae	Gadiformes	76
<i>Gaidropsarус ensis</i>	Phycidae	Gadiformes	77
<i>Gasterosteus aculeatus</i>	Gasterosteidae	Gasterosteiformes	83
<i>Gasterosteus wheatlandi</i>	Gasterosteidae	Gasterosteiformes	84
<i>Glyptocephalus cynoglossus</i>	Pleuronectidae	Pleuronectiformes	208
<i>Gonichthys cocco</i>	Myctophidae	Myctophiformes	107
<i>Gonostoma elongatum</i>	Gonostomatidae	Stomiiformes	262
<i>Gymnelus viridis</i>	Zoarcidae	Perciformes	194

Species	Family	Order	Page number
<i>Gymnophanrus tricuspidis</i>	Cottidae	Scorpaeniformes	229
<i>Halargyreus johnsoni</i>	Moridae	Gadiformes	72
<i>Halosauropsis macrochir</i>	Halosauridae	Albuliformes	16
<i>Helicolenus dactylopterus</i>	Scorpaenidae	Scorpaeniformes	251
<i>Hemitripterus americanus</i>	Hemitripteridae	Scorpaeniformes	241
<i>Hippocampus erectus</i>	Syngnathidae	Gasterosteiformes	87
<i>Hippoglossoides platessoides</i>	Pleuronectidae	Pleuronectiformes	209
<i>Hippoglossus hippoglossus</i>	Pleuronectidae	Pleuronectiformes	210
<i>Holtbyrnia macrops</i>	Platytroctidae	Osmertiformes	138
<i>Hoplostethus atlanticus</i>	Trachichthyidae	Beryciformes	41
<i>Hoplostethus mediterraneus</i>	Trachichthyidae	Beryciformes	42
<i>Howella sherborni</i>	Acropomatidae	Perciformes	139
<i>Hygophum benoiti</i>	Myctophidae	Myctophiformes	107
<i>Hygophum hygomii</i>	Myctophidae	Myctophiformes	108
<i>Hyperoglyphe perciformis</i>	Centrolophidae	Perciformes	155
<i>Icelus bicornis</i>	Cottidae	Scorpaeniformes	230
<i>Icelus spatula</i>	Cottidae	Scorpaeniformes	231
<i>Ilyophis brunneus</i>	Synaphobranchidae	Anguilliformes	26
<i>Katsuwonus pelamis</i>	Scombridae	Perciformes	172
<i>Laemonema barbatula</i>	Moridae	Gadiformes	73
<i>Lampadena luminosa</i>	Myctophidae	Myctophiformes	108
<i>Lampadena speculigera</i>	Myctophidae	Myctophiformes	109
<i>Lampanyctus ater</i>	Myctophidae	Myctophiformes	110
<i>Lampanyctus crocodilus</i>	Myctophidae	Myctophiformes	111
<i>Lampanyctus festivus</i>	Myctophidae	Myctophiformes	112
<i>Lampanyctus intracarius</i>	Myctophidae	Myctophiformes	112
<i>Lampanyctus macdonaldi</i>	Myctophidae	Myctophiformes	113
<i>Lampanyctus photonotus</i>	Myctophidae	Myctophiformes	114
<i>Lampanyctus pusillus</i>	Myctophidae	Myctophiformes	114
<i>Lampris guttatus</i>	Lamprididae	Lampridiformes	89
<i>Lepidion eques</i>	Moridae	Gadiformes	74
<i>Lepidocybium flavobrunneum</i>	Gempylidae	Perciformes	161
<i>Lepidophanes guentheri</i>	Myctophidae	Myctophiformes	114
<i>Lepophidium cervinum</i>	Ophidiidae	Ophidiiformes	126
<i>Lestidiops affinis</i>	Paralepididae	Aulopiformes	35
<i>Limanda ferruginea</i>	Pleuronectidae	Pleuronectiformes	211
<i>Lionurus carapinus</i>	Macrouridae	Gadiformes	63
<i>Liopsetta putnami</i>	Pleuronectidae	Pleuronectiformes	212
<i>Liparis atlanticus</i>	Liparidae	Scorpaeniformes	243
<i>Liparis fabricii</i>	Liparidae	Scorpaeniformes	244
<i>Liparis gibbus</i>	Liparidae	Scorpaeniformes	245
<i>Lipogenys gilli</i>	Notacanthidae	Albuliformes	17
<i>Lobianchia dofleini</i>	Myctophidae	Myctophiformes	115
<i>Lobianchia gemellarii</i>	Myctophidae	Myctophiformes	116
<i>Lophius americanus</i>	Lophiidae	Lophiiformes	92
<i>Lopholatilus chamaeleonticeps</i>	Malacanthidae	Perciformes	165
<i>Lumpenus lumpretaeformis</i>	Stichaeidae	Perciformes	186
<i>Lumpenus maculatus</i>	Stichaeidae	Perciformes	187
<i>Lycenchelys paxillus</i>	Zoarcidae	Perciformes	195
<i>Lycenchelys verrilli</i>	Zoarcidae	Perciformes	196
<i>Lycodes esmarki</i>	Zoarcidae	Perciformes	197
<i>Lycodes lavalaei</i>	Zoarcidae	Perciformes	198
<i>Lycodes pallidus</i>	Zoarcidae	Perciformes	199

Species	Family	Order	Page number
<i>Lycodes reticulatus</i>	Zoarcidae	Perciformes	199
<i>Lycodes vahlii</i>	Zoarcidae	Perciformes	200
<i>Macrorhamphosus scolopax</i>	Macrorhamphosidae	Gasterosteiformes	86
<i>Macrourus berglax</i>	Macrouridae	Gadiformes	64
<i>Macrozoarces americanus</i>	Zoarcidae	Perciformes	201
<i>Malacosteus niger</i>	Stomiidae	Stomiiformes	270
<i>Mallotus villosus</i>	Osmeridae	Osmeriformes	136
<i>Maurolicus muelleri</i>	Sternopychidae	Stomiiformes	266
<i>Melanogrammus aeglefinus</i>	Gadidae	Gadiformes	55
<i>Melanonus zugmayeri</i>	Melanonidae	Gadiformes	67
<i>Melanostigma atlanticum</i>	Zoarcidae	Perciformes	202
<i>Menidia menidia</i>	Atherinidae	Atheriniformes	29
<i>Merluccius albidus</i>	Merlucciidae	Gadiformes	68
<i>Merluccius bilinearis</i>	Merlucciidae	Gadiformes	69
<i>Microgadus tomcod</i>	Gadidae	Gadiformes	56
<i>Micromesistius poutassou</i>	Gadidae	Gadiformes	57
<i>Molva dypterygia</i>	Gadidae	Gadiformes	58
<i>Molva molva</i>	Gadidae	Gadiformes	59
<i>Morone americana</i>	Moronidae	Perciformes	166
<i>Morone saxatilis</i>	Moronidae	Perciformes	167
<i>Mugil curema</i>	Mugilidae	Mugiliformes	95
<i>Mullus auratus</i>	Mullidae	Perciformes	168
<i>Myctophum asperum</i>	Myctophidae	Myctophiformes	117
<i>Myctophum punctatum</i>	Myctophidae	Myctophiformes	118
<i>Myctophum selenops</i>	Myctophidae	Myctophiformes	119
<i>Myoxocephalus aenaeus</i>	Cottidae	Scorpaeniformes	232
<i>Myoxocephalus octodecemspinosis</i>	Cottidae	Scorpaeniformes	233
<i>Myoxocephalus scorpioides</i>	Cottidae	Scorpaeniformes	234
<i>Myoxocephalus scorpius</i>	Cottidae	Scorpaeniformes	235
<i>Narcetes stomias</i>	Alepocephalidae	Osmeriformes	131
<i>Nemichthys scolopaceus</i>	Nemichthyidae	Anguilliformes	23
<i>Nessorhamphus ingolfianus</i>	Derichthyidae	Anguilliformes	22
<i>Nezumia bairdi</i>	Macrouridae	Gadiformes	65
<i>Nezumia sclerorhynchus</i>	Macrouridae	Gadiformes	66
<i>Notacanthus chemnitzii</i>	Notacanthidae	Albuliformes	18
<i>Notolepis rissoii</i>	Paralepididae	Aulopiformes	36
<i>Notolychnus valdiviae</i>	Myctophidae	Myctophiformes	120
<i>Notoscopelus bolini</i>	Myctophidae	Myctophiformes	120
<i>Notoscopelus caudispinosus</i>	Myctophidae	Myctophiformes	121
<i>Notoscopelus elongatus kroeyerii</i>	Myctophidae	Myctophiformes	121
<i>Notoscopelus resplendens</i>	Myctophidae	Myctophiformes	122
<i>Oncorhynchus mykiss</i>	Salmonidae	Salmoniformes	220
<i>Oneirodes</i> sp.	Oneirodidae	Lophiiformes	94
<i>Osmerus mordax</i>	Osmeridae	Osmeriformes	137
<i>Paralepis atlantica</i>	Paralepididae	Aulopiformes	37
<i>Paralichthys dentatus</i>	Paralichthyidae	Pleuronectiformes	206
<i>Paralichthys oblongus</i>	Paralichthyidae	Pleuronectiformes	207
<i>Paraliparis calidus</i>	Liparidae	Scorpaeniformes	246
<i>Paraliparis copei</i>	Liparidae	Scorpaeniformes	247
<i>Parasudis triculenta</i>	Chlorophthalmidae	Aulopiformes	33
<i>Peprilus tricanthus</i>	Stromateidae	Perciformes	189
<i>Peristedion miniatum</i>	Triglidae	Scorpaeniformes	255
<i>Pholis gunnellus</i>	Pholidae	Perciformes	169

Species	Family	Order	Page number
<i>Pogonias cromis</i>	Sciaenidae	Perciformes	171
<i>Pollachius virens</i>	Gadidae	Gadiformes	60
<i>Polyacanthonotus rissoanus</i>	Notacanthidae	Albuliformes	19
<i>Polyipnus asteroides</i>	Sternopychidae	Stomiiformes	267
<i>Polymixia lowei</i>	Polymixiidae	Polymixiiformes	216
<i>Polyprion americanus</i>	Acropomatidae	Perciformes	140
<i>Pomatomus saltatrix</i>	Pomatomidae	Perciformes	170
<i>Poromitra megalops</i>	Melamphaidae	Stephanoberyciformes	258
<i>Prionotus carolinus</i>	Triglidae	Scorpaeniformes	256
<i>Prionotus evolans</i>	Triglidae	Scorpaeniformes	257
<i>Protomyctophum arcticum</i>	Myctophidae	Myctophiformes	122
<i>Pseudopleuronectes americanus</i>	Pleuronectidae	Pleuronectiformes	213
<i>Pungitius pungitius</i>	Gasterosteidae	Gasterosteiformes	85
<i>Reinhardtius hippoglossoides</i>	Pleuronectidae	Pleuronectiformes	214
<i>Rondeletia loricata</i>	Rondeletiidae	Stephanoberyciformes	260
<i>Rouleina attrita</i>	Alepocephalidae	Osmeriformes	132
<i>Ruvettus pretiosus</i>	Gempylidae	Perciformes	162
<i>Salmo salar</i>	Salmonidae	Salmoniformes	221
<i>Salmo trutta</i>	Salmonidae	Salmoniformes	222
<i>Salvelinus alpinus</i>	Salmonidae	Salmoniformes	223
<i>Salvelinus fontinalis</i>	Salmonidae	Salmoniformes	224
<i>Sarda sarda</i>	Scombridae	Perciformes	173
<i>Scomber japonicus</i>	Scombridae	Perciformes	174
<i>Scomber scombrus</i>	Scombridae	Perciformes	175
<i>Scomberesox saurus</i>	Scomberesocidae	Beloniformes	39
<i>Scomberomorus brasiliensis</i>	Scombridae	Perciformes	176
<i>Scomberomorus cavalla</i>	Scombridae	Perciformes	176
<i>Scomberomorus maculatus</i>	Scombridae	Perciformes	177
<i>Scopelogadus beanii</i>	Melamphaidae	Stephanoberyciformes	259
<i>Scophthalmus aquosus</i>	Scophthalmidae	Pleuronectiformes	215
<i>Sebastes fasciatus</i>	Scorpaenidae	Scorpaeniformes	252
<i>Sebastes marinus</i>	Scorpaenidae	Scorpaeniformes	253
<i>Sebastes mentella</i>	Scorpaenidae	Scorpaeniformes	254
<i>Selene setapinnis</i>	Carangidae	Perciformes	151
<i>Seriola dumerili</i>	Carangidae	Perciformes	152
<i>Seriola zonata</i>	Carangidae	Perciformes	153
<i>Serrivomer beani</i>	Serrivomeridae	Anguilliformes	25
<i>Simenchelys parasiticus</i>	Synaphobranchidae	Anguilliformes	27
<i>Spectrunculus grandis</i>	Ophidiidae	Ophidiiformes	127
<i>Sphoeroides maculatus</i>	Tetraodontidae	Tetraodontiformes	275
<i>Stenotomus chrysops</i>	Sparidae	Perciformes	184
<i>Sternopyx diaphana</i>	Sternopychidae	Stomiiformes	268
<i>Stichaeus punctatus</i>	Stichaeidae	Perciformes	187
<i>Stomias boa</i>	Stomiidae	Stomiiformes	271
<i>Symbolophorus veranyi</i>	Myctophidae	Myctophiformes	123
<i>Sympodus pterospilotus</i>	Cynoglossidae	Pleuronectiformes	203
<i>Synagrops bellus</i>	Acropomatidae	Perciformes	141
<i>Synagrops spinosa</i>	Acropomatidae	Perciformes	142
<i>Synaphobranchus kaupi</i>	Synaphobranchidae	Anguilliformes	28
<i>Syngnathus fuscus</i>	Syngnathidae	Gasterosteiformes	88
<i>Taaningichthys bathyphilus</i>	Myctophidae	Myctophiformes	124
<i>Taaningichthys minimus</i>	Myctophidae	Myctophiformes	124
<i>Taractichthys longipinnis</i>	Bramidae	Perciformes	147

Species	Family	Order	Page number
<i>Tautoga onitis</i>	Labridae	Perciformes	163
<i>Tautogolabrus adspersus</i>	Labridae	Perciformes	164
<i>Tetrapturus albidus</i>	Xiphiidae	Perciformes	192
<i>Thunnus alalunga</i>	Scombridae	Perciformes	178
<i>Thunnus albacares</i>	Scombridae	Perciformes	179
<i>Thunnus obesus</i>	Scombridae	Perciformes	180
<i>Thunnus thynnus</i>	Scombridae	Perciformes	181
<i>Trachyrhynchus murrayi</i>	Macrouridae	Gadiformes	66
<i>Triglops murrayi</i>	Cottidae	Scorpaeniformes	236
<i>Triglops nybelini</i>	Cottidae	Scorpaeniformes	237
<i>Triglops pingeli</i>	Cottidae	Scorpaeniformes	238
<i>Trigonolampa miriceps</i>	Stomiidae	Stomiiformes	272
<i>Ulvaria subbifurcata</i>	Stichaeidae	Perciformes	188
<i>Urophycis chesteri</i>	Phycidae	Gadiformes	78
<i>Urophycis chuss</i>	Phycidae	Gadiformes	79
<i>Urophycis regia</i>	Phycidae	Gadiformes	80
<i>Urophycis tenuis</i>	Phycidae	Gadiformes	81
<i>Venefica procera</i>	Nettastomatidae	Anguilliformes	24
<i>Vinciguerria nimbaria</i>	Gonostomatidae	Stomiiformes	263
<i>Xenodermichthys copei</i>	Alepocephalidae	Osmeriformes	133
<i>Xiphias gladius</i>	Xiphiidae	Perciformes	193
<i>Zenopsis conchifera</i>	Zeidae	Zeiformes	278



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